

**CHARACTERIZATION OF CANDIDATE GENES IN ENGLISH COCKER
SPANIEL HEREDITARY NEPHRITIS**

A Dissertation

by

ZENAIDO TRES CAMACHO

Submitted to the Office of Graduate Studies of
Texas A&M University
in partial fulfillment of the requirements for the degree of

DOCTOR OF PHILOSOPHY

December 2003

Major Subject: Veterinary Microbiology

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ABSTRACT

Characterization of Candidate Genes in English Cocker Spaniel Hereditary Nephritis.
(December 2003)

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Six different isoforms of Type IV collagen (colIV α 1-6) have been identified. The individual isoforms of colIV are termed alpha chains and are translated from six different COLIV genes (COLIVA1-A6). Collagen Type IV gene products compose the structural framework of basement membranes. The glomerular basement membrane (GBM) is a specialized basement membrane involved in the ultrafiltration processes of the kidney. The colIV α 1- α 5 chains are expressed in the human GBM while the colIV α 1- α 6 chains are expressed in the canine GBM. Many inherited diseases of the kidney have been reported and mutations in genes regulating kidney function have been identified. Alport syndrome (AS) is the most common form of human hereditary nephritis (HN). AS is defined as an inherited progressive kidney disorder associated with sensorineural deafness and is characterized by extensive thickening and multilaminar splitting of the GBM when examined by electron microscopy. AS has both X-linked (XLAS) and autosomal (ARAS) modes of inheritance. Mutations in the COLIVA5 gene are responsible for XLAS. A form of HN with characteristic splitting of the GBM with X-linked inheritance has been described in Samoyed dogs. A specific mutation in the

COLIVA5 gene has been identified in Samoyed dogs affected with HN. Mutations in the COLIVA3 and COLIVA4 genes are responsible for ARAS. A form of HN has been identified in English cocker spaniel dogs (ECS) that has been described as autosomal in inheritance and includes GBM abnormalities including extensive lamination characteristic of ARAS. Both ARAS and ECS-HN show loss of the colIVA3 and colIVA4 chains in the GBM when examined with monoclonal antibodies. ECS-HN has been hypothesized to have the same molecular basis of disease as ARAS. As such, we have isolated and characterized canine COLIVA3 and COLIVA4 sequences from normal dogs and ECS dogs affected with HN and compared the coding regions of these candidate genes.

TABLE OF CONTENTS

	Page
ABSTRACT.....	iii
TABLE OF CONTENTS.....	v
 CHAPTER	
I INTRODUCTION.....	1
The Collagens.....	1
Type IV Collagen.....	5
Extracellular Matrices.....	16
Kidney Structure and Function.....	24
Mechanisms of Glomerular Permselectivity.....	28
Alport Syndrome and Other Diseases of the GBM.....	35
Canine Hereditary Nephritis.....	44
Summary.....	50
 II THE ISOLATION AND CHARACTERIZATION OF CANINE	
COLIVA3.....	52
Introduction.....	52
Materials and Methods.....	54
Results.....	59
 III THE ISOLATION AND CHARACTERIZATION OF CANINE	
COLIVA4.....	62
Introduction.....	62
Materials and Methods.....	64
Results.....	69
 V SUMMARY AND CONCLUSIONS.....	72
REFERENCES.....	84

TABLE OF CONTENTS (continued)

	Page
APPENDIX 1.....	106
APPENDIX 2.....	114
APPENDIX 3.....	122
APPENDIX 4.....	126
APPENDIX 5.....	135
APPENDIX 6.....	145
APPENDIX 7.....	149
APPENDIX 8.....	158
APPENDIX 9.....	167
APPENDIX 10.....	170
VITA.....	171

CHAPTER I

INTRODUCTION

The Collagens

Collagens are found in all vertebrate species as well as in *Drosophila*, sea urchins, nematodes and sponges (Blumberg et al., 1988). It is suggested that all collagens evolved from a common ancestral unit of a 54 base pair (bp) repeat (Chu et al., 1984). Although diverse in their distribution and function, all collagens share features that group them as a family. In 1986 a conference was convened to discuss the biochemistry of collagens (Uitto et al., 1986). At the time of the conference 10 members of the collagen family had been described. Five years previous, only five members of the collagen family had been reported. Whereas many of the initial collagens were described on a biochemical basis of classification, many of the new collagens being reported were based on genetic sequence homologies. One of the main objectives of the conference was to derive a consensus definition of a collagen. As a result, a collagen is described as a triple helical domain that is expressed extracellularly where it performs a structural function. The word collagen is derived from the ancient Greek word *kolla* for glue and the helical nature of collagens has been known for some time.

This dissertation follows the format of Journal of Cell Biology.

X-ray crystallography was used to elucidate the double helical nature of DNA and this method was also used to determine the triple helical nature of collagen fibers. In 1954, Ramachandran and Kartha first used this technique to identify the interchain hydrogen bonding and the importance of glycine in the amino acid composition of collagen (Ramachandran, 1954). In 1955, they later reported a more detailed spatial analysis, emphasized the importance of proline residues and described the coiled coil nature of collagen (Ramachandran, 1955). In the same year Rich and Crick, using the same methodology, added information on the primary amino acid sequence and proposed a refined structure (Rich, 1955). The helical structure of collagens is essential for the association of the three alpha chains and is termed the collagenous domain of the molecule. The helical structure is made possible by the incorporation of the amino acid glycine at every third residue creating the characteristic collagenous sequence. Collagens are triple helical molecules with the sequence Gly-X-Y where Gly represents the amino acid glycine while X and Y can represent any other amino acid. Most commonly X and Y are represented by the amino acids proline or hydroxyproline (which account for approximately 20% of the collagenous structure) or hydroxylysine which is used to create interchain stabilizing bonds with other hydroxylysine amino acids to stabilize the quaternary structure of the molecule.

The collagens undergo extensive post-translational modification. Lysine is modified by lysyl hydroxylase and the altered amino acid is then used to form interchain crosslinks used for quaternary stabilization. This enzyme was first isolated in 1958 and identified as a necessary physiological component of crosslinking collagens and elastin,

a molecule found in the basement membranes of the heart (Pinnell and Martin, 1968). The necessary nature of these crosslinks is evident by the addition of lathyrogen which prevents crosslinkage. Lathyrogens, such as β -aminoproionitrile (BAPN), are found in plants and when digested by livestock can lead to extreme illness or death. The inherited connective tissue disease Ehlos-Dandrome Syndrome type VI is also associated with a defect in lysyl hydroxylase (Pinnell et al., 1972). Hydroxylysine is also used as a site for glycosylation (Spiro, 1967a). Type IV collagen of the GBM was also shown to be glycosylated (Spiro, 1967b). The importance of the glycosylation is unknown but could possibly help the molecules become more hydrophilic, an essential characteristic of basement membranes. The collagens are synthesized intracellularly in an immature form and then transported extracellularly. The immature collagen is termed procollagen and was first identified in cultured human fibroblasts using chromatographic techniques (Layman et al., 1971) and the bones of rats using pulse-chase experiments (Bellamy and Bornstein, 1971). These reports showed that the conversion from a pre-procollagen to a procollagen occurred extracellularly by an excreted enzyme (then termed a zymogen) that had proteolytic activity. In 1979 the specific proteolytic enzyme responsible for the release of the N- and C-terminal pro-peptides was identified (Kao et al., 1979).

Glycine is necessary at every third residue in the collagenous sequence because its single hydrogen atom as its side chain is the smallest of all amino acids and causes the least possible amount of steric hindrance. Substitution of any other amino acid creates a steric hindrance that disrupts the helical structure. Thus, a base change creating a single amino acid substitution could create disturbances in the structural integrity of

the collagen molecule. This was first seen in mutations in collagen type I genes in osteogenesis imperfecta (Vuorio and de Crombrughe, 1990). Collagen molecules consist of three monomers wound together in the left-handed triple helix. The individual monomeric molecules are termed alpha chains. The collagens are an extended family of proteins and are categorized as types based on various biochemical techniques (Uitto et al., 1986). Each type of collagen is given a Roman numeral as its type designation (type I collagen, type II collagen, etc.) and at least 19 types of vertebrate collagen have been identified (Kivirikko and Myllyla, 1987; Prockop and Kivirikko, 1995; van der Rest and Garrone, 1991; Vuorio and de Crombrughe, 1990).

Although the first collagen was isolated from the tail tendon of rats by acid extraction in the widely reported experiments by Nagoette beginning in 1927, the next identified collagen was not identified until 1969 from chicken cartilage (Miller and Matukas, 1969). This second identified collagen created the need for a system of nomenclature. The present nomenclature of collagens categorizes this collagen as collagen type II and the first collagen isolated by Nagoette as collagen type I. In 1971, Kefalides isolated a unique collagen from the anterior lens capsule from bovines and identified this as type IV collagen (Kefalides, 1971). The technique of cyanogen bromide cleavage of purified collagen extract used by Kefalides was the same method used to identify other collagens including the identification of collagen type III isolated from chicken skin a year earlier (Miller et al., 1971). In 1963, with the use of ion exchange chromatography, Piez first identified the alpha chain subunits of collagen type I (Piez, 1963). He termed the most abundant of the alpha chains $\alpha 1$ and the second most

abundant alpha chain $\alpha 2$ and described the nature of collagen type I as being composed of two $\alpha 1$ chains and one $\alpha 2$ chain, thus describing a trimer of $(\alpha 1)_2(\alpha 2)_1$ (Piez, 1963). A collagen molecule could be composed of a single species of identical alpha chains or a combination of different alpha chain species. A single alpha chain species is associated with a single collagen type. The identity of the alpha chain is defined by the type of collagen with which it is associated and by its order of identity with respect to the other monomers of the same type. For example, the alpha 1 chain of type I collagen ($\text{colIV}\alpha 1$) was defined before the alpha 2 chain of type I collagen ($\text{colIV}\alpha 2$) and the alpha 1 chains of type I collagen ($\text{colI}\alpha 1$) are not the same as the alpha 1 chains of type II collagen ($\text{ColII}\alpha 1$). The role of the collagens is structural and together they are the most abundant protein in the body.

Type IV Collagen

Six different isoforms of mammalian type IV collagen ($\text{colIV}\alpha 1-6$) have been identified. The individual isoforms of colIV are termed alpha chains and are translated from six different colIV genes (COLIVA1-A6) (Hudson et al., 1993). All colIV isoforms have a similar function, molecular structure and genetic composition, but different molecular organization and patterns of expression. The characteristics of collagen type IV will be the focus of this section.

Structurally, collagen type IV is an essential component of basement membrane architecture. Type IV collagen allows the formation of a network of collagen molecules rather than a collagen fiber, because the mature colIV molecule has characteristics that make it different from the fiber forming collagens. Type IV collagen contains more than 20 interruptions in the Gly-X-Y collagenous sequence resulting in a more flexible collagen molecule (Brazel et al., 1988; Dolz et al., 1988; Hofmann et al., 1984). These interruptions in the collagenous sequences range from 21 interruptions for the $\alpha 1$ chain of type IV collagen (colIV $\alpha 1$) (Brazel et al., 1987) to 26 interruptions for the colIV $\alpha 4$ chain (Mariyama et al., 1994). The colIV $\alpha 2$ and colIV $\alpha 3$ chains each have 23 interruptions in their collagenous sequence (Brazel et al., 1988; Leinonen et al., 1994) while the colIV $\alpha 5$ and colIV $\alpha 6$ chains have 22 and 25 interruptions (Pihlajaniemi et al., 1990; Zhou et al., 1994b). The multiple interruptions of collagen type IV are different with respect to the fiber forming collagens like collagen type I which contains an uninterrupted collagen domain (Hofmann et al., 1984).

Terminal non-collagenous sequences of fiber forming collagens are cleaved, but for type IV collagen these sequences are preserved and used for self assembled networks. Three alpha chains of colIV molecules self associate in parallel fashion with their c-terminals at one end of the triple helix and their amino terminals at the other end of the molecule (Timpl et al., 1981). Both the c-terminal globular domain and the amino terminal collagenous domain of type IV collagen are involved in higher order assembly. Each alpha chain has a c-terminal non-collagenous globular domain and collectively the c-terminals of a colIV molecule are termed the NC-1 domain (Timpl et al., 1981). NC-1

domains from two triple-helical colIV molecules associate to form c-terminal linked dimers. Amino terminal sequences from four colIV molecules associate and, collectively, forms a structure termed the 7S domain (Kuhn et al., 1981).

The 7-S domain of type IV collagen was identified from multiple sources including EHS tumor cells, bovine lens capsule and human kidney (Dixit et al., 1981; Risteli et al., 1981). This novel collagen domain had a sedimentation coefficient of 7.2S and resisted bacterial collagenase but had an amino acid sequence similar to basement membrane collagen. The molecule was found to be highly glycosylated, but the origin of the molecule was unknown. The 7S domain is secured with disulfide bonds and hydrophobic interactions (Siebold et al., 1987). The 7S domain has many important features. These include a 21 residue N-terminal non-triple helical region (NH-1), a 117 residue triple helical region (TH-1) that is responsible for the aggregation of the four molecules and a 10 residue non-triple helical region (NH2) that creates an area responsible for flexibility in this region (Siebold et al., 1987). Through these associations, colIV molecules create a network and form the structural foundation of basement membranes.

The network model for the organization of type IV collagen molecules in basement membranes was first proposed after the identification of the non-collagenous c-terminal domain termed the NC-1 domain (Timpl et al., 1981). This model suggested a regular network of type IV collagen molecules generated by sites interacting at the opposite ends of each molecule. By rotary shadowing electron microscopy, it was observed that the 7S domain connected four molecules while the NC-1 domain

connected two molecules and formed a lattice network. The proposed structure indicated a network similar to chicken wire that was continuous and could explain the mechanical strength and flexibility of basement membranes. Later, it was shown by electron microscopy that type IV collagen could self assemble *in vitro* when purified collagen type IV isolated from EHS cells was heated and allowed to reconstitute (Yurchenco and Furthmayr, 1984). This report suggested a more complex network than the initial chicken wire model and indicated lateral assembly of the molecules as opposed to a strict head to head and tail to tail configuration. Lateral associations were shown to exist through similar methods of experimentation and identified the mechanism of lateral associations to be from interactions of the NC-1 domain with specific intermolecular sites (Tsilibary and Charonis, 1986). The lateral association was inhibited by anti NC-1 antibodies. This is an important finding with regards to this research because it showed that, while the NC-1 domain is important for molecular stability, there are several sites within the molecule for investigation with regards to disruption of the matrix that might result in pathogenesis. In other words, the NC-1 domain must have sites internally in associated molecules with which to bind. Further complexity of the matrix assembly was shown *in situ* by the observation of a super helical formation of different monomeric trimers of collagen type IV molecules around an established basement membrane matrix. This modified model of the structural framework of basement membrane matrices indicated a structure much more like a fabric than chicken wire (Yurchenco and Ruben, 1987).

The most common network of colIV molecules in basement membranes is composed of two colIV alpha 1 chains and one colIV alpha 2 chain (colIV(α 1)₂(α 2)₁) (Hudson et al., 1993). It was shown that these trimers are formed beginning at the NC-1 domain of the C-terminus, proceeds to the N-terminus 7S region, that the process can be reversed at 50°C, and the trimer can be reconstituted at 25°C (Dolz et al., 1988). Removal of the NC-1 domain with pepsin leads to a loss of the refolding ability. The colIV (α 1)₂(α 2)₁ trimer has been found to be present in most basement membranes and has wide tissue distribution (Trueb et al., 1982). The minor species of collagen type IV (α 3, α 4, α 5, and α 6), have been shown to have a limited tissue distribution (Kleppel et al., 1989a). These six colIV alpha chains could in theory form 56 different kinds of triple helical monomers (Hudson et al., 1993). Definitive evidence regarding all of the molecular associations of the colIV alpha chains is still an active area of research, although some molecular associations have been reported. As previously mentioned, the colIV (α 1)₂(α 2)₁ trimer has been identified in most basement membranes. Using co-precipitation techniques with monoclonal antibodies to colIV NC-1 domains, a heterodimeric complex of two colIV α 3 and one colIV α 4 chain in the GBM has been reported (Johansson et al., 1992). Earlier studies in the same tissue with the same techniques indicated a homotrimeric structure of three colIV α 3 chains (Saus et al., 1988). ColIV α 3 homodimers, (colIV α 3)₂, have also been reported in embryonic tissues (Haralson et al., 1985). More recently, investigators have reported a disulfide crosslinked complex of colIV α 3- α 4- α 5 chains of type IV collagen from bovine kidney (Gunwar et al., 1998). This finding could explain why patients with Alport Syndrome frequently

show no immunohistochemical staining of either the colIV α 3, α 4 or α 5 chains which are normally present in human GBM (Kashtan et al., 1996; Kleppel et al., 1989a; Nakanishi et al., 1994; Yoshioka et al., 1994). In these patients, genetic analysis shows that the molecular basis of disease has been identified as a mutation in the COLIVA5 gene and that the COLIVA3 and COLIVA4 genes are unaffected and transcribed at normal levels. Therefore, these results suggest that mutations in the gene encoding the colIV α 5 chain causes defective assembly of not only the colIV α 5 chains, but also the colIV α 3 and colIV α 4 chains in the GBM. For our studies the converse could also be true, mainly, that mutations in either the COLIVA3 or COLIVA4 genes that disrupt protein structure could also disrupt the assembly of colIV α 3- α 4- α 5 trimers and thus lead to the same GBM defect. The exact nature of the colIV interactions is still unknown. The molecular assembly of colIV products could involve multiple combinations the colIV α 3, colIV α 4, colIV α 5 and colIV α 6 chains. The association of these chains could also be at the transcription/translation level because, as will be discussed below, mRNA expression of the colIV α 3, α 4 and α 5 chains appears to be coordinated (Thorner et al., 1996). Research on seminiferous tubule basement membrane in bovine testes has shown three major hexameric complexes of colIV molecules (Kahsai et al., 1997). One is the classic colIV(α 1) $_1$ (α 2) $_2$ network, and the others represent two novel networks. One of the novel networks involves all of the colIV alpha chain species (colIV α 1- α 6), while the other novel network is restricted to the colIV α 3- α 6 chains. Beyond these novel networks, using monoclonal antibodies and HPLC technology the content of colIV α 3 in these

basement membrane structures was calculated to be 40% while the content of the colIV α 4 chains was 18% (Kahsai et al., 1997). The result of this research shows highly complex, interactive, coordinated and heterogeneous nature in the composition of colIV molecules in basement membrane matrices.

Collagen type IV genes are highly conserved. Our research using Genbank of comparative sequence analysis of corresponding colIV alpha chains in different mammalian species, consistently show over 80% homology at the nucleotide level. COLIV genes share similar genetic structure and all consist of many exons over large areas of genomic DNA. Each mammalian COLIV gene has over 50 exons and covers at least 100 kb of genomic DNA (Soininen et al., 1989; Zhou et al., 1994a; Zhou et al., 1994b). COLIV genes generate similar sized transcripts of over 5,000 bases and produce similar sized products. The colIV α 1 chain represents the smallest product with 1,642 amino acids (Soininen et al., 1987) and the colIV α 2 chain is the largest with 1,676 amino acid residues (Hostikka and Tryggvason, 1988). The human col IV α 3, α 4 α 5 and α 6 chains have 1,642, 1,652, 1,659 and 1,670 amino acids respectively (Leinonen et al., 1994; Mariyama et al., 1994; Zhou et al., 1992), (Zhou et al., 1994b)).

Although the colIV “classical” chains (colIV α 1 and colIV α 2) have been shown to be present in most basement membranes, the minor species of colIV (colIV α 3, colIV α 4, colIV α 5 and colIV α 6) have a limited tissue distribution. The limited distribution of these products suggests a specific functions in the tissues where they are expressed. The isolation of the minor species of collagen all arose from investigations into the molecular composition of the GBM. As such, much of the investigation into

collagen type IV tissue distribution has been focused primarily on basement membranes of the kidney. Immunohistochemical staining with monoclonal antibodies can be used to study the expression of specific proteins. Chain-specific antibodies directed against individual collagen type IV α chains can be used to detect expression of these products in specific basement membranes.

In the kidney, the colIV α 3 and colIV α 4 chains are expressed predominately in the GBM and are also detected to a minor extent in Bowman's capsule and the distal tubules (Butkowski et al., 1989; Kleppel et al., 1989b) and, extrarenally, these chains are also detected in the testis (Kahsai et al., 1997). The colIV α 5 chain is also detected in the above locations, as well as the collecting ducts of the kidney and, extrarenally, in the skin (Kleppel et al., 1992). The restricted distribution of these molecules suggests a specified role in these structures.

The homology of the collagen gene family with respect to molecular techniques can be seen in the history of the isolation of the COLIVA5 gene (Pihlajaniemi et al., 1990). In an investigation to isolate collagen type V genes (not COLIV genes), a human placental cDNA library was screened with an avian fibrillar collagen gene. This probe was expected to hybridize strongly to collagen type V genes and several colonies with positive signals were isolated. Two of the clones were COLIA1 genes and one was a COLIVA1 sequence. Another clone isolated in this study had interruptions of the collagenous sequence and was identified as a previously unidentified type IV collagen sequence, COLIVA5.

The COLIVA6 gene is X-linked and lies head to head, adjacent to the COLIVA5 gene, where they share a common promoter (Oohashi et al., 1994; Sugimoto et al., 1994; Zhou et al., 1994a). The isolation of the COLIVA6 gene followed the identification of a distinct pattern found in the COLIV genes. In the analysis of the COLIVA1-A5 sequences, it was found that the genes could be divided into two distinct groupings. Based on DNA sequence homology, the COLIVA1, COLIVA3 and COLIVA5 genes were found to belong to an $\alpha 1$ -like class and the COLIVA2 and COLIVA4 genes were classified as $\alpha 2$ -like genes. This nomenclature also identified the colIV $\alpha 1$ and colIV $\alpha 2$ as “classical” chains to distinguish them from the other tissue specific chains. Because it appeared that there was a missing member of the alpha 2-like genes, and it was shown that the COLIVA1 and COLA2 genes are arranged in a head to head configuration and shared common promoter elements, it was hypothesized that the COLIVA5 gene was likewise paired with an unidentified collagen type IV gene. Using the homology inherent in these genes, a COLIVA4 clone was used to screen a human lens library, and several overlapping clones were isolated (Oohashi et al., 1994). Characterization of these sequences showed them to contain a novel COLIV isoform that was identified and classified as the COLIVA6 gene (Oohashi et al., 1994). Mapping of COLIVA6 localized the gene to the same region as COLIVA5 on chromosome Xq22 and showed that the genes are also transcribed by a common promoter (Sugimoto et al., 1994). Although COLIVA6 is expressed in some tissues in the kidney such as Bowman's capsule and the collecting ducts, it is not expressed in the GBM of humans or in young dogs, but is expressed in older dogs (Peissel et al., 1995; Lees et al., 2000).

As mentioned previously, the $\text{colIV}\alpha 1$ and $\text{colIV}\alpha 2$ chains were first identified in 1971 by Kefalides from cyanogen bromide cleavage of bovine anterior lens capsule (Kefalides, 1971). The genes for the $\text{colIV}\alpha 1$ and $\text{colIV}\alpha 2$ monomers of type IV collagen have been isolated, sequenced and located autosomally on human chromosome 13 (Griffin et al., 1987; Saus et al., 1989; Soininen et al., 1988). These genes in the human are arranged in a head to head configuration and are transcribed by a common bi-directional promoter (Poschl et al., 1988; Soininen et al., 1988). This promoter region had no detectable transcriptional activity in transient gene expression assays after fusion to the chloramphenicol acetylase (CAT) gene in either direction indicating the necessity of additional elements for efficient and tissue specific expression of both genes (Poschl et al., 1988). No molecular defects in these genes have been identified in HN. It is unlikely that mutations in COLIVA1 and COLIVA2 genes would be found in specific diseases since these genes are expressed in embryonic development and their products are found in most basement membranes (Trueb et al., 1982). In other words, structural defects in these molecules would not be tolerated in development and would lead to wide ranging pathologies. In fact, temperature sensitive mutations in the COLIVA1 and COLIVA2 genes in *C. elegans* cause embryonic lethality (Guo 1989). It is suggested that genetic defects resulting in a loss of function in a tissue specific collagen species, such as COLIVA3 or COLIVA4, is more likely in autosomal forms of HN.

The $\text{colIV}\alpha 3$ chain was first isolated using antibodies from patients with Goodpasture syndrome (GS). GS is an autoimmune disorder in humans where patients have circulating antibodies to GBM components and develop rapidly progressive

glomerulonephritis. After collagenase digestion, preparations from bovine GBM that reacted with Goodpasture sera identified the NC-1 domain of collagen type IV as the targets of Goodpasture antibodies (Wieslander et al., 1984). Further investigation with lens basement membranes using reverse-phase HPLC, SDS-PAGE and amino acid sequencing showed the Goodpasture antigen to be a novel collagen type IV isoform designated colIV α 3 (Butkowski et al., 1987; Saus et al., 1988). Another result of this same investigation was the identification of another novel collagen type IV isoform designated colIV α 4. Although the above investigation only briefly characterized these novel proteins with respect to differential electrophoretic migration patterns and immunoreactivity, the discovery of new colIV isoforms opened the door to many other findings including the understanding of basement membrane structure and function. These findings also helped to explain the molecular basis of many inherited basement membrane pathologies that will be discussed below.

The human COLIVA3 and COLIVA4 genes have been mapped to human chromosome 2 and, similar to the genes described above, are situated in a head to head configuration (Kamagata et al., 1992; Mariyama et al., 1992; Morrison et al., 1991). They have also been mapped autosomally in the dog (personal communication). In the human genome, both the COLIVA3 and COLIVA4 genes have been suggested to cover over 100 kb and to contain over 50 exons (Boye et al., 1998; Heidet et al., 2001). Each gene produces a product of approximately 1,700 amino acids from an ~5,000 base coding region (Leinonen et al., 1994; Mariyama et al., 1994). Partial bovine COLIV cDNA sequences have also been isolated using human sequences as primers for PCR

amplification (Morrison et al., 1991). The COLIVA3 and COLIVA4 genes produce major components of the GBM and aberrations in these genes have been found in humans with autosomal HN which will be discussed in later sections (Boye et al., 1998; Lemmink et al., 1994; Mochizuki et al., 1994).

Extracellular Matrices

Extracellular matrices (ECMs) are classified into two categories. One form of ECM fills spaces between cells and has many receptors for the binding of cytokines and growth factors. The other form of ECM creates basement membranes, which are thin sheets around tissues and were initially termed reticulin (Rambourg and Leblond, 1967). Together the separate components of basement membranes create a scaffold for cells and maintain the structural integrity of many tissues. Although having similar components, basement membranes are not necessarily the same in all tissues. Early immunological assays of the canine kidney showed differences between the basement membranes of the tubules and of the GBM (Krakower, Goodman 1955). Their research showed that although these basement membranes were made predominantly from collagen, the antigenic epitopes differed between these structures although, at the time of this report, the collagens causing the immunological reactions they observed had not been identified. The ECM plays an important role in modulating the behavior of cells with which it interacts. The major molecules that constitute the ECM include collagen type

IV, entactin/nidogen (E/N), laminin and heparine sulfate glycoprotein (HSGP). A model system for the study of ECM components is the Engelbreth-Holm-Swarm (EHS) cell line, which is derived from a benign mouse tumor and produces abundant ECM molecules (Swarm, 1964). The EHS matrix consists of 60% laminin, 30% colIV, 3% heparine sulfate proteoglycans and 6% E/N (Kleinman et al., 1986).

The glomerular basement membrane (GBM) is a specialized ECM created predominately by podocytes of the kidney. Aberrations in the GBM have been shown to be associated with podocyte abnormalities including effacement and resulting kidney pathologies including proteinuria. The major components of the GBM and their molecular interactions will be discussed in this section, except for colIV whose role in the GBM has been described above and will also be discussed in subsequent sections.

Although constituting a small proportion of overall basement membrane mass, E/N will be discussed first because of its ability to bind to all of the other major components of the GBM. Two groups characterized this molecule independently from different sources. The first group described a basement membrane component isolated from a mouse carcinoma cell line and named the molecule entactin (Carlin et al., 1981). Another group isolated a similar product from the EHS matrix and named the molecule nidogen (Timpl et al., 1983). Both molecules were glycoproteins of approximately 150 kD. Amino acid and cDNA sequence analysis of entactin and nidogen in mouse and human demonstrated that both were identical molecules and thus can be referred to collectively as E/N (Durkin et al., 1988; Mann et al., 1989; Nagayoshi et al., 1989). The gene for E/N has been located autosomally on human chromosome 1 (Olsen et al.,

1989), and it has been shown that E/N expression can be detected at the 8-16 cell stage in mouse development (Dziadek and Timpl, 1985; Wu et al., 1983). Expression of E/N has also been found in the GBM (Saxena et al., 1990). Electron microscopic rotary shadowing of E/N shows a linear molecule composed of three globular domains (G1, G2 and G3) separated by two rod-like domains, and has a total length of approximately 30nm (Fox et al., 1991). E/N binds to laminin via the G3 domain (Mann et al., 1988) and to collagen IV via its G2 domain (Aumailley et al., 1989; Fox et al., 1991). E/N has also been shown to bind to the protein core of HSGPs in basement membranes (Battaglia et al., 1992). Thus, although E/N was first identified by co-immunoprecipitation with laminin, it appears that apart from being able to bind to itself, E/N can bind to all of the other major basement membrane components. Its ubiquitous and early expression indicate a strong role in basement membrane assembly.

Other isoforms of E/N have been reported in mouse and human (Kimura et al., 1998; Kohfeldt et al., 1998). The mouse isoform has been termed entactin-2 while the human isoform is termed nidogen-2. These new isoforms can be conjointly termed E/N-2. These molecules show similar structure to the original E/N-1 molecules, but share less than 50% sequence homology to the E/N-1 isoforms (Kohfeldt et al., 1998). E/N-2 is proposed to have similar function to E/N-1 because E/N-2 binds to laminin, but at a different site and with less affinity than E/N-1, and has colIV and HSGPs binding ability equal to the E/N-1 isoforms (Erickson and Couchman, 2000). The discovery of E/N-2 suggests the other E/N isoforms may exist and may be a growing family of ECM molecules with different tissue specific functions.

Many non-collagenous glycoproteins are building blocks of ECMs, but the laminins are the most ubiquitous and abundant in basement membranes. Laminin was first isolated from an EHS matrix (Timpl et al., 1979). Laminins are large heterotrimeric molecules of 400-600 kD and it has been shown that the laminins can self assemble *in vitro* into independent networks via intermolecular bonding (Yurchenco et al., 1992; Yurchenco et al., 1985). The network forming ability of laminin is thought to be important for basement membrane stability. The laminins have evolved into a family of proteins with many different isoforms. These laminin isoforms include five different α -chains ($\alpha 1$ - $\alpha 5$), three different β -chains ($\beta 1$ - $\beta 3$) and three different γ -chains ($\gamma 1$ - $\gamma 3$) (Burgeson et al., 1994). The composition of a mature laminin molecule includes one of each of the three chains. It is possible to predict which laminin trimers are present in basement membranes by co-localization with immunohistochemical staining of individual laminin chains. Theoretically, 45 trimeric formations of laminin chains can exist (5α chains x 3β chains x 3γ chains). To date, however, only 12 different laminin isoforms have been reported (Burgeson et al., 1994). The most abundant laminin in the GBM is laminin-11 composed of a trimer of $\alpha 5\beta 2\gamma 1$ chains (Miner et al., 1997). Laminin-9 ($\alpha 4\beta 2\gamma 1$) has been reported in developing GBM (Miner et al., 1997), but the $\alpha 4$ chain gradually disappears as the glomerulus matures (Miner et al., 1998). The functions of the laminins have been inferred by identification of mutations in these genes in specific diseases, or by knockout experiments of specific laminin chains in mice. It is presumed that the knockout of a specific laminin chain will result in the loss of function of the mature laminin trimeric molecule. Knockout experiments involving the $\beta 2$ chain

in mice results in early death due to massive proteinuria and show fusion of the podocyte foot processes, although, the GBM appears normal (Noakes et al., 1995). While the $\alpha 5$ chain of laminin is also present in laminin-10 ($\alpha 5\beta 1\gamma 1$) which is present in most basement membranes of the kidney (Miner et al., 1997), laminin $\alpha 5$ genetic knockouts also affects laminin-11 ($\alpha 5\beta 2\gamma 1$) in the GBM. As expected, the $\alpha 5$ knockouts have much more devastating effects on kidney development and the mice die *in utero* (Miner et al., 1998). The $\alpha 5$ chain of laminin has been shown to be essential for glomerulogenesis, and the laminin $\alpha 5$ knockout mice also have neural tube defects that indicate the importance of this chain in other tissues (Miner and Li, 2000).

Like the E/N isoforms described above, the laminins can form associations with other ECM molecules. A strong component of laminin function in basement membrane architecture involves molecular associations with integrins. Integrins are a diverse family of integral membrane glycoproteins that function to mediate interactions of cells with the ECM. Integrins also function to mediate cell:cell interactions and cell signaling (DeSimone et al., 1987). Integrins are dimeric proteins that contain α and β chains. There have been 19 integrin α and 8 integrin β chains reported and 25 different combinations of these chains have been described (Humphries, 2000). The molecular association of laminins and integrins directly links the ECM to the surface of the cells. Many regions of laminin have been reported to associate with integrins and many integrin–laminin associations have been reported (Belkin and Stepp, 2000).

As mentioned previously, the laminins bind E/N in a 1:1 equimolar association; that is, one molecule of laminin binds one molecule of E/N. Unlike E/N, however,

laminin can self assemble to form an independent network in basement membranes (Yurchenco et al., 1992). The specific epitope of laminin that binds E/N has been located on the γ chains with the $\gamma 1$ chain having the strongest association, and antibodies to this region inhibit laminin:E/N binding (Mayer et al., 1993). Thus, the laminin-E/N interaction connects the colIV and laminin networks as well as other ECM components such as the HSPGs. The direct binding of laminin to HSGPs can increase these interactions. The triple helical core of laminin has been shown to bind to agrin, a specific HSGP in the in the GBM (Denzler et al., 1997).

The carbohydrate content of basement membranes was first demonstrated by periodic acid-shiff staining techniques and attributed to glycoproteins in 1957 (Leblond and Inoue, 1989). Heparine sulfate glycoproteins (HSPGs) are found in all basement membranes and consist of a protein core to which several carbohydrate chains are attached (Paulsson et al., 1987). HSPGs contain heparine sulfate as its glycosaminoglycan (GAG) side chain that is composed of unbranching repeating disaccharide units with the aminosugar glucosamine. The glucosamine component of HSPG is extremely sulfated and creates a hydrophilic positively charged environment in basement membranes (Kanwar and Farquhar, 1979b). The anionic nature of the GBM created by the presence of HSPG is extremely important to GBM permselectivity and will be discussed in later sections.

Based on immunoflourecent studies with antibodies directed against the core proteins of HSGPs, the most abundant HSGP in the GBM is agrin (Groffen et al., 1997; Groffen et al., 1998). Low levels of agrin are also detected in other basement membranes

in the kidney in the rat, but are attributed to different agrin isoforms (Raats et al., 1998). The 95nm protein core of agrin has many domains but its most N-terminal region has been shown to be necessary for the binding of laminin-1 (Denzer et al., 1997; Denzer et al., 1995). Agrin has also been shown to bind laminin-2 (Cotman et al., 1999) and integrin $\alpha 5 \beta 1$ (Martin and Sanes, 1997) and, although these specific isoforms have not been identified in the GBM, as has been discussed, intermolecular associations are essential for GBM architecture. Although this investigation focuses on the components of the GBM, agrin is also known as a key component of the neuromuscular junction (Denzer et al., 1998). Agrin was first identified as a synaptic organizing protein associated with aggregation of acetylcholine receptors and acetylcholinesterase at regenerating neuromuscular junctions *in vivo* in Torpedo (an oceanic ray) (Nitkin et al., 1987). Initially identified as an aggregating factor, the molecule was termed agrin. Agrin has been characterized as a large glycoprotein of over 400 kD when fully glycosylated (Tsen et al., 1995) and has been mapped autosomally in both human and mouse (Kallunki et al., 1991; Rupp et al., 1991). The importance of agrin has been demonstrated with knockout experiments in mice which result in aberrant kidney development and death at birth (Gautam et al., 1996).

Other proteoglycans in basement membranes of the kidney include perlecan, bamacan and colXVIII. Perlecan is widely expressed in the kidney, but has restricted distribution and is not predominant in the GBM (Groffen et al., 1998). Bamacan is a chondroitin sulfate glycoprotein which also has widespread distribution in the kidney, but has not been detected in the GBM (Wu and Couchman, 1997). ColXVIII has been

shown to be a glycoprotein (Halfter et al., 1998) and expression of colXVIII α 1 has been reported in the GBM (Muragaki et al., 1995). ColXVIII has been classified into a subfamily of non-fibrillar collagens with multiple interrupted triple helical domains, but any role in any pathology has yet to be determined (Oh et al., 1994).

One of the fascinating aspects of basement membrane biology is the ability of the individual components to self assemble *in vitro*. Individual basement membrane components from EHS cells (including laminin, E/N, collagen type IV and HSPG) were isolated from purified extracts and, when combined, at physiologic temperatures, spontaneously assembled into structures that resembled the lamina densa of basement membranes (Grant et al., 1989).

The increased knowledge of basement membranes, including the above studies, indicates a complexity of basement membrane architecture through specific molecular associations that reflect the diverse nature and function of all ECMs. For example, it is now problematic to discern the exact point of transition from the ECM, the cell membrane and the cytoskeleton. The specific focus of this study is directed toward the GBM, a specialized ECM. The GBM is subjected to tremendous forces and constant pressure from birth until death. The delicate nature of this structure can be revealed if any component has a loss of function from either environmental influences or genetic aberrations. The composition of basement membrane structure that has been described above is based on specific intramolecular and intermolecular associations and has been focused primarily on molecules expressed in the GBM. Interruptions in these molecular

associations result in decreased performance of the GBM and, as will be discussed below, an overall loss of kidney function.

Kidney Structure and Function

The kidneys regulate arterial pressure, extracellular fluid osmolarity, blood and extracellular fluid volumes, acid-base balances in the body and the excretion of nitrogenous waste products. Although the kidneys perform many physiological functions, the focus of this investigation will concentrate on the kidney as the site for the ultrafiltration of plasma.

Many aspects of human kidney function can be found in numerous textbooks on physiology including “The Textbook of Medical Physiology” (Guyton, 1991) and “Physiology” [Sperelakis, 1996]. Other resources written with specific reference to kidney function include “(The) Kidney” [Brenner 1991] and “Pathology of the Kidney” [Heppinstall, 1992]. These references will be used to describe the basic nature of kidney physiology.

Normally, there are both left and right kidneys and each kidney is profused with one renal artery and drained with one renal vein. In between, the flow from the renal artery is divided into individual arterioles, many of which introduce blood to individual nephrons. Although there are two kinds of nephrons, the cortical nephrons and the juxtaglomerular nephrons, each nephron is capable of formation of urine. Therefore, although nephrons in mammalian kidneys number in the hundreds of thousands, urine

formation can be described using a single nephron. Each glomerulus, its associated tubule and vascular structure defines a nephron. Blood flows to the glomerulus, the main filter of the kidney, through the afferent arteriole and exits through the efferent arteriole. Every component of blood is passed through the capillary structure of the glomerulus. The afferent arteriole is divided into approximately fifty capillaries in each glomerulus. The capillaries of the glomerulus are uniquely fenestrated, that is, the capillaries of the glomerulus have thousands of small holes averaging from 500 Å to 100 Å in diameter. This allows the blood of the capillary structure of the glomerulus to come into direct contact with the underlying basement membrane (GBM). The GBM is sandwiched between the endothelial capillaries of the glomerulus and the epithelial cells that surround the glomerular capillaries. The epithelial cells are called podocytes. Podocytes are specialized epithelial cells that interdigitate with one another to form the foot processes. The interdigitation of podocytes forms slit diaphragms (also known as filtration slits) in the glomerular epithelium. The podocytes of the glomerulus are surrounded by another epithelial structure known as Bowman's capsule. Bowman's capsule forms the first structure of the urinary space and is the collecting site for the primary filtrate that is created by the ultrafiltration of plasma.

Thus, ultrafiltration of plasma is directed in the glomerulus from the fenestrated capillary across the GBM and through the slit diaphragms created by the epithelial podocytes and into Bowman's capsule. From Bowman's capsule the primary filtrate flows into the tubular structure of the nephron where further processing of the primary filtrate occurs. The active participation of the tubular epithelial cells contribute to the

processing of the primary filtrate and is an essential process for the ultimate formation of urine.

One of the basic functions of urine formation can be described as the excretion of unwanted substances. Unwanted substances in the blood include excesses of sodium, potassium and chloride ions as well as end products of metabolism including urea and creatinine. In as it is a basic function of the nephron to clear the plasma of unwanted substances, the nephron must be selective and not allow cells or large proteins to pass into the urinary space. While the average human has about 40 liters of total body fluid, 180 liters of plasma is filtered per day, creating 14 liters of primary filtrate and ultimately 1.4 liters of urine per day. Therefore, the glomerulus filters a large amount of plasma to create a primary filtrate that flows into the urinary space and then into the tubular portion of the nephron. In the tubular structure, the unwanted substances are passed in the formation of urine, while most of the water and many other solutes are reabsorbed. Most of the soluble components of the blood pass through the filtration mechanisms of the glomerulus and are present in the primary filtrate. Electrolytes are reabsorbed with water according to homeostatic demands. Larger serum proteins are normally not allowed passage into the tubular space. One of the smallest of plasma proteins, as compared to fibrinogen at 340 kD or immunoglobulins at 150-900 kD, is albumin with a molecular weight of 69 kD. Albumin is normally not allowed passage through the GBM into the urinary space. While small amounts of proteins are present in the proximal tubule, with normal kidney function, these proteins are cleared during urine formation. Small proteins that may pass through the ultrafiltration processes undergo

pinocytosis via cells of the proximal tubule. Proteinuria refers to the presence of protein in the urine. Hematuria refers to the presence of erythrocytes or cell structures (cell casts) in the urine. Neither is present in normal urine. A wide range of pathologies, many of which will be described in later sections, are associated with abnormalities in the filtration process.

The structure and function of the mammalian kidney has long been a subject of fascination and experimentation. An experimental model of nephritis was established in 1900 by Lindemann by introducing serum from a guinea pig immunized with rat kidney that became nephrotoxic when injected into rabbits. This experimental model was refined further when sonicated preparations of GBM were used in the initial immunization helping to establish the GBM at the target site for nephrogenic antigens (Rohrbach, 1991). Electron microscopy had been used for many purposes, but it was not until 1948 when a method described by Pease and Baker was used to make fine sectioning of biological tissues that this powerful tool could be used to examine fine tissue structure (Pease and Baker, 1950). Their method allowed an ultrathin sectioning of .05 microns and their survey resulted in many findings. They were able to describe the “brush border” of the proximal tubule, the fenestrated nature of the glomerular endothelium and detailed descriptions of the glomerular basement membrane. They describe the GBM as “pictorially most spectacular”. They described the GBM as thin (300-350 nm) and existing as a continuous layer around the fenestrated endothelium that is in direct contact with the epithelial cells of the glomerulus. These studies, among others, have shown the GBM to be a specialized structure in the kidney. The molecular

composition of the GBM has been described above. The role of the GBM with respect to ultrafiltration and the importance of changes in the GBM associated with disease will be discussed below.

Mechanisms of Glomerular Permselectivity

The mechanisms of glomerular permeability have been an object of discussion for some time. In 1960, a seminal study was done by Farquhar et al. describing ferritin transfer across the glomerular capillary wall to explore the mechanisms of glomerular ultrafiltration (Farquhar et al., 1961). Although the structure of the renal glomerulus had been described, little was known about the mechanisms involved in the transport of substances from the capillary to the urinary space. At that time two prevailing hypotheses had been established for glomerular filtration. One was that the glomerular basement membrane acted as the main filter of the kidney since the majority of the electron microscopists had described the GBM as the only continuous layer around the capillary wall. The other hypothesis indicated that the spaces between the foot processes of the epithelial cells acted as slit pores allowing for the free passage of water and small molecules but restricted the passage of proteins. The approach of Farquhar et al to explore the mechanisms of glomerular permeability was to directly observe the pathways of glomerular filtration across the capillary wall by introducing a particulate tracer able to be seen by electron microscopy. Their main conclusion, held for many decades, was

that the glomerular basement membrane was the main filter of the kidney in that there were no identified channels across the foot processes. They observed, however, that the filtration was not perfect and that any small molecules, especially proteins, would be absorbed through pinocytosis in the epithelium. They further concluded that the function of the GBM is probably dependent on both the epithelium and endothelial layers and any agent that would interfere with these cells could affect the chemical composition and therefore the permeability of the basement membrane. This group reinforced their conclusions with a report using graded dextrans to determine the molecular weight limitations of glomerular filtration (Caulfield and Farquhar, 1974). These results found that the size limitations of glomerular filtration using dextran sulfate were 40-50 kD. While 14 kD dextran was cleared from the bloodstream easily, progressive size resulted in decreased clearance until approximately 55-60 kD where there was a sharp decline of clearance. This corresponded to the clearance of albumin, which at a molecular weight of 68 kD does not cross the GBM. In other words, they identified the size limitation of the filtration of the GBM at 10A° while the size of albumin is 36A°. Their results helped to reinforce that the GBM is the primary filtration barrier to plasma proteins. Thus, the size and weight of molecules is important for glomerular permselectivity and further studies showed that the charge of a molecule is also significant in determining “charge-selectivity” (Chang et al., 1975). Again, using the normally neutrally charged dextran sulfate in the previous research, charges were added to the molecules to make it either positive or negatively charged. Their results showed that positively charged molecules pass through the GBM much faster than a neutrally charged molecule and that a

negatively charged molecule is inhibited from crossing. It is interesting to note that albumin is a negatively charged molecule. The mechanisms of charge and filtration is based on the fact that the heparine sulfate glycosaminoglycans (GAGs) that are imbedded in the GBM are anionic (Kanwar and Farquhar, 1979a). In fact, the loss of these anionic sites is associated with some forms of nephrosis (Caulfield and Farquhar, 1978; Rohrbach et al., 1983) and the removal of GAGs using GAG-degrading enzymes allows albumin to pass through the GBM (Rosenzweig and Kanwar, 1982). In more recent reports, an acute, selective proteinurea could be induced in rats by a single intraveinuous injection of a monoclonal antibody directed against the most abundant HSPG in the GBM, agrin (van den Born et al., 1992).

Some of the first evidence for an alternative view of glomerular ultrafiltration came from studies using peroxidase molecules as tracers (Graham and Karnovsky, 1966). The peroxidase molecules were twice the size of human albumin and large enough to be retained by the glomerular structure. These studies found that a greater concentration of reaction product was retained in the portion of the basement membrane adjacent to the epithelial foot processes. They further reported that much of the product was retained on the epithelial membranes and between the foot processes adjacent to mesangial areas and more specifically in the epithelial slits. Only after some time did the peroxidase product stain the glomerular basement membrane. There was little disagreement that filtrate passed from the capillary lumen through the endothelial fenestrae, across the basement membrane, and through the epithelial slits into the urinary space, but these studies suggested the restriction of passage occurred at the epithelial

slits. This challenged the hypothesis that the GBM was the principle glomerular filter and suggested that the main filter barrier is distal to the GBM located at the epithelial slits; that the GBM was regulated in function as a “course filter” and that the epithelial slits are the principle filtration barrier.

During this early stage of research, however, the structural evidence for a mechanism at the slit pore was unknown, but the use of new staining techniques identified structures in support of this hypothesis. With the use of tannic acid fixative for electron microscopy, a structure termed the “slit diaphragm” was identified at the epithelial slits between the foot processes (Rodewald and Karnovsky, 1974). The slit diaphragm was identified as an isoporous structure with rectangular pores of approximately 40A° across, 140A° through the slit diaphragm and 70A° in length. The dimensions of the pores found in the slit diaphragm were smaller than the 70A° width of albumin, and supported the hypothesis that the slit diaphragm was the principal filtration barrier to plasma proteins. Furthermore, these studies showed that the slit diaphragm was a continuous, two-dimensional band that extended to all foot processes so it is a necessary structure for molecules to cross this in the filtration process and diminished the uniqueness of the GBM in this regard. Detailed analysis of the electro-imagery reported in this same investigation described another new “zipper-like” spacing of cross bridges within the slit diaphragm although the molecular nature of these novel structures could not be revealed until recently.

Mapping the genetic defect in congenital nephrotic syndrome of the Finnish (NPHS1) type, an autosomal recessive disorder in humans, identified the first of these

molecules in the slit diaphragm. Congenital nephrotic syndrome (CNS) is a heterogeneous group of renal diseases that result in increased postnatal glomerular permeability and proteinuria early in life. Mapping of NSPH1 using positional cloning localized the gene to a 150 kb region on chromosome 19q13.1 and sequencing of this region revealed 11 candidate genes, one of which was mutated in affected families (Kestila et al., 1998). Over 30 different mutations in the NSPH1 gene (*Nphs1*) have been identified (Lenkkeri et al., 1999). *Nphs1* codes for nephrin, a large transmembrane glycoprotein, that is 26 kilobases, has 29 exons, and codes for a 4.3 kb mRNA and is a member of the Ig superfamily (Kestila et al., 1998). Rat and mouse nephrin genes have also been characterized and show high homology and similar organization to the human genes (Putala et al., 2000). Nephrin has been specifically located to the slit diaphragm in human (Ruotsalainen et al., 1999) and mouse (Holzman et al., 1999). It is postulated that pairs of nephrin molecules compose the molecular zipper between the foot processes of podocytes. A model for NPHS1 has been created by inactivation of the *Nphs1* gene in mice that will provide a powerful system for future studies (Putala et al., 2001). Also, the glomerular specific promoter for the nephrin gene has been inserted into two systems that may facilitate the identification of transacting factors that are required for podocyte-specific expression of GBM molecules. (Moeller et al., 2000; Wong et al., 2000)

Another necessary component for slit diaphragm assembly was previously well characterized, and yet its function in the kidney has only recently been revealed. The CD2-associated protein (CD2AP) is critical for stabilizing contacts between T cells and antigen presenting cells and is termed CMS in humans. Knockout mice in this gene were

developed in an effort to determine the loss of CD2-associated protein (CD2AP) on immune function (Shih et al., 1999). A knockout of the first intracellular SRC homology domain (SH3) in blastocysts resulted in a diminished immune capacity, but these mice also developed a form of congenital nephrotic syndrome. The cause of renal failure was determined to be due to the loss of foot processes in the glomerulus. Further, it was shown that the CD2AP co-immunoprecipitated with nephrin (Shih et al., 1999). CD2AP is expressed in the slit diaphragm and binds nephrin via a novel c-terminal domain (Shih et al., 2001). Because of the known function of CD2AP in T cells, it is suggested that CD2AP anchors nephrin to the internal protein fibers that form the podocyte cytoskeleton to help form and stabilize the slit diaphragm (Yuan et al., 2002).

The study of pathologies with apparent filtration barrier defects can provide useful models for the discovery of the genes and molecules responsible for normal glomerular filtration and the molecular basis of disease. This has been demonstrated in the study of focal segmental glomerulosclerosis (FSGS). FSGS is heterogeneous group of disorders manifested as non-specific renal lesions secondary to other disorders or as a specific inherited condition. FSGS may be defined by the presence of segmental sclerosis in some, but not necessarily all glomeruli, and is characterized by increased urinary protein excretion and decreased kidney function leading to end-stage renal failure (Ichikawa and Fogo, 1996).

The steroid-resistant form of FSGS in humans (SRN) is an autosomal recessive disorder and has been mapped to chromosome 1q25-31 (Fuchshuber et al., 1995). Recently, mutations in the *Nsph2* gene have been reported in families with SNR (Boute

et al., 2000). The product of *Nsph2* has been termed podocin because the expression of this gene is restricted to fetal and adult podocytes. The podocin gene codes for a 42 kD protein from a 1,149bp open reading frame. It is a putative integral membrane protein and shows homology to human stromatin, which appears to be an ion channel associated with the cytoskeleton. Although the actual function of podocin is unknown, it is possible that it may be involved in organizing the cytoskeleton of podocyte foot processes at the slit diaphragm and may also interact with other podocyte proteins necessary for slit diaphragm assembly. It has been reported that podocin co-localizes with nephrin to the actin cytoskeleton protruding from the tips of actin filaments (Saleem et al., 2002). The cytoplasmic side of podocin has molecular homology for binding to an SH-3 binding site similar to the structure found in the CD2AP above.

The gene responsible for the autosomal dominant form of FSGS has been mapped to human chromosome 19q13 and mutations in the gene encoding α -actin-4 (ACTN4), which resides at this locus, have been reported in three families (Kaplan et al., 2000). Although the exact function of ACTN4 with regards to FSGS is unknown, expression of this gene occurs in capillaries, glomerular podocytes and in other tissues such as circulating lymphocytes. ACTN4 is an actin crosslinking protein and it is postulated that regulation of the actin cytoskeleton of glomerular podocytes may be altered in FSGS thus interfering in normal filament assembly in the foot processes. This indicates that the actin cytoskeleton in podocytes may be involved in normal glomerular ultrafiltration and in the pathogenesis of proteinuria. Because ACTN4 is also expressed in capillaries, it is possible that a loss of function in this gene could also change the renal

vasculature cytoskeleton and thus alter glomerular hemodynamics leading to the described phenotype. Regions relating to other forms of inherited FSGS have been mapped in the human, but the specific genes for these syndromes have not yet been identified.

These studies show a complex design of glomerular ultrafiltration that includes the coordinate function of the slit diaphragm and the GBM involving many gene products. The function of each of these genes is associated and/or dependent on intermolecular and/or intramolecular associations of other GBM components. The loss of function of any of these components can, therefore, lead to molecular dissociation and subsequent failure of overall glomerular ultrafiltration processes and, hence, an overall loss of kidney function. As will be discussed, the products of the COLIV genes have been shown to be specifically involved in these processes.

Alport Syndrome and Other Diseases of the GBM

Although there are many diseases that are aimed at various parts of the nephron, like the slit diaphragm pathologies described above, diseases referred to as HN can be located directly to components of the GBM. Owing to its similarities to the form of hereditary nephritis in English cocker spaniel dogs which is the focus of this investigation, Alport Syndrome (AS) will be discussed more fully than other diseases of the GBM.

In humans, AS is the most common form of HN with an estimated gene frequency of 1:5000 (Atkin, 1988). Dr. Cecil Alport in 1927 first reported the symptoms associated with a new clinically relevant syndrome, which is now referred to as Alport Syndrome (AS) (Alport, 1927). Dr. Alport described an inherited nephrotic condition that was associated with deafness where male members of a family progressed to end stage renal disease (ESRD). The characteristics of AS have since been shown to be very heterogeneous. In 1988 AS was classified into six different types based on clinical criteria (Atkin, 1988). Type I was classified as juvenile Alport with deafness, type II was X-linked with deafness, type III was X-linked adult with deafness, type IV was X-linked adult without deafness, type V was autosomal with deafness and thrombocytopathy and type VI was autosomal with deafness.

The most common and identifiable characteristic of AS are renal abnormalities. In individuals with hereditary nephritis, the lamina of the GBM show structural abnormalities including overall thickening and splitting that can be seen by electron microscopy (Churg and Grishman 1972; Hinglais et al. 1972; Spear and Slusser 1972). AS can be diagnosed when the characteristic splitting of the GBM is observed (Heptinstall, 1992).

The most common extra-renal defect associated with AS is deafness, which is frequent, but not universal, and occurs in approximately 50% of all patients (Wester et al., 1995). While renal disorders are not always accompanied with hearing loss in AS, members with hearing loss in families diagnosed with AS always have accompanied renal disorders (Gubler et al., 1981). Also, there have been no reports of deaf males

without kidney disease transmitting AS to any offspring (Gubler et al., 1981). While the exact mechanism for the loss of hearing in AS patients has not yet been determined, the defect has been isolated to areas of the inner ear (cochlea).

Other common extra renal abnormalities associated with AS include a variety of ocular defects. While ocular defects may be present in less than half of all AS cases (Chance and Stanley, 1977), a condition known as anterior lenticonus is virtually pathognomonic of AS (Nielsen, 1978). Anterior lenticonus is the protrusion of the central portion of the lens into the anterior capsule and is most associated with AS families with deafness and with rapid progression to end stage renal disease. Other ocular defects seen in patients with AS include changes in various basement membranes of the eye including Bruch's membrane, Descemet's membrane and corneal basement membranes (Govan, 1983; Rhys et al., 1997; Teekhasaene et al., 1991).

Other extra renal abnormalities associated with AS include leiomyomatosis (hyperplasia of smooth muscle) (Cochat et al., 1988), and hematological defects such as thrombocytopenia (Epstein et al., 1972). The presence of hematological defects in AS is interesting because unlike the tissues described above, blood cells do not have classic basement membrane architecture.

Since the diagnosis of AS has been based on certain clinical findings, confusion over diagnostic procedures and AS classification is understandable given the heterogeneity described above. Further complications over the proper diagnosis and classification of AS can be attributed to heterogeneity within a particular type of AS. The severity of symptoms varies from person to person and small kindreds contain too

few individuals to meet many of the clinical and histologic criteria for proper classification. Precise diagnosis and classification of AS is important for several reasons including genetic counseling and the establishment of individual prognoses. Furthermore, there are many diseases that can be confused with AS.

Although difficulties in the diagnosis of AS have been reported, a core of traits can be associated with all types of AS that have been described above. These include a hereditary disease with chronic hematuria that is progressive and characterized ultrastructurally by irregular GBM structure. Furthermore, AS can be differentiated from immune related glomerulonephritic diseases. Most importantly, AS can be generally classified as having either X-linked or autosomal inheritance.

Of the many types of AS that have been classified, the molecular basis of disease was first identified for X-linked AS (XLAS). In the initial investigation of the molecular basis of XLAS, it was known that basement membrane ultrastructural aberrations were pathognomonic of AS and that collagen type IV made up the structural framework of the GBM (Churg and Sherman, 1973; Hinglais et al., 1972; Spear and Slusser, 1972). Also, the genetic element responsible for XLAS had been mapped to the X chromosome (Atkin et al., 1988; Szpiro-Tapia et al., 1988). The identification of the minor species of type IV collagen including the COLIVA5 gene (Hostikka et al., 1990; Pihlajaniemi et al., 1990) and the mapping of COLIVA5 to the X-chromosome (Hostikka et al., 1990) identified COLIVA5 as a strong candidate gene. These reports focussed efforts toward the screening of this gene in families with X-linked AS. The first mutations in the COLIVA5 gene were reported in 1990 (Barker et al., 1990). Since that time, hundreds of

different mutations have been found in all parts of the COLIVA5 gene in affected families (Martin et al., 2000). These genetic abnormalities include deletions, insertions, and splice mutations. All of these mutations have been found to alter the COLIVA5 gene producing nonsense mutations that introduce premature stop codons or missense mutations that result in the substitution of amino acids. Frequently these missense mutations result in the substitution of a glycine residue. As discussed previously, glycine is necessary at every third residue for proper collagen folding and function. Major rearrangements in the COLIVA5 gene are very rare (Boye et al., 1991; Renieri et al., 1992). Even though linkage to the COLIVA5 gene has been established, mutations in the COLIVA5 gene can not always be found (Kawai et al., 1996).

Proper folding and subsequent trimer formation is essential for collagen type IV function. The loss of function is visualized in immunohistochemical studies that show the absence of staining of the product in target tissues. The collagen type IV molecules create distinct networks to form the structural framework of the basement membranes. The COLIVA5 product, which is usually present in the GBM, Bowman's capsule and distal tubule basement membrane, is absent in patients with X-linked AS (Nakanishi et al., 1994). COLIVA5 is also normally expressed in the lens capsule and Decemet's membrane of the eye, in the cochlea of the ear and in the skin. In patients with X-linked AS COLIVA5 is also reduced or absent in these tissues. Females who are heterozygous for COLIVA5 mutations frequently show abnormal staining of tissues expressing COLIVA5 (Kleppel et al., 1989a; Yoshioka et al., 1994) (Yoshioka et al., 1994). It is suggested that the variations in histochemical staining patterns in females heterozygous

for COLIVA5 mutations is due to differential lyonization in these tissues of the COLIVA5 locus. It is important to note that the absence of COLIVA5 staining in X-linked AS correlates with the absence of COLIVA3 and COLIVA4 staining in tissues where these genes are also normally expressed including the GBM (Nakanishi et al., 1994).

Mutations in the COLIVA3 and COLIVA4 genes are responsible for autosomal recessive AS. The first study to show aberrations in the COLIVA3 and COLIVA4 genes reported findings from four families (Mochizuki et al., 1994). Two of the families had mutations in the COLIVA3 gene and two had mutations in the COLIVA4 gene. Both of the mutations in the COLIVA3 gene were found in young females diagnosed with juvenile AS and each of these patients had GBM structures characteristic of AS. Both patients also had sensorineural loss of hearing. One of the COLIVA3 mutations found was a five base deletion that resulted in a premature stop codon in the NC-1 domain. The other COLIVA3 mutation was also a nonsense mutation in the NC-1 domain, but resulted from a single C to T transition. Mutations in the COLIVA4 gene were also identified from young female patients with juvenile AS from two other families, but unlike the positioning of the mutations in the NC-1 domain of the COLIVA3 genes above, both of the COLIVA4 mutations were found in areas coding for collagenous sequences. One COLIVA4 mutation was a G-A transition that changed a glycine to a serine and the other, found in a different region, was a single C-A transversion that resulted in a premature stop codon. Several other mutations have been reported in the COLIVA3 and COLIVA4 genes in patients diagnosed with autosomal AS (Boye et al.,

1998; Ding et al., 1995; Heidet et al., 2001; Lemmink et al., 1994; Longo et al., 2002). The above studies show that like the mutations in the COLIVA5 gene in X-linked AS described above, mutations in the COLIVA3 and COLIVA4 have been found in all parts of these genes and are not confined to any one area.

Thin basement membrane disease (TBMD) is also known as benign familial hamaturia and, as the name suggests, is characterized by a thinning of the GBM and is inherited as an autosomal dominant trait in humans (Gauthier et al., 1989; Rogers et al., 1973). TBMD has been linked to the COLIVA3/COLIVA4 locus (Buzza et al., 2001b; Lemmink et al., 1996). Twelve different mutations in the COLIVA4 gene in twelve different families with TBMD have been identified (Buzza et al., 2001a; Lemmink et al., 1996; Ozen et al., 2001)(Buzza 2003). Mutations in the COLIVA3 gene in families with TBMD have also been reported (Longo et al., 2002). Again, the mutations in these genes include nonsense mutations resulting from deletions and insertions, or missense mutations resulting in glycine substitutions that disrupt the triple helical nature of the molecule and the mutations are not localized and have been found throughout these genes. Mutations in either the COLIVA3 and COLIVA4 genes exist as heterozygotes in individuals with TBMD and mutations in both genes existing simultaneously have not been reported. Carriers of X-linked AS may also show thinning of the GBM (Rumpelt et al., 1974). The heterozygous nature of the mutations suggests that individuals diagnosed with TBMD may be carriers of autosomal recessive AS.

An autosomal dominant form of AS has been described in humans (Feingold et al., 1985). These families have a less severe phenotype with a slower rate of progression

as compared to patients with X-linked AS (Pochet et al., 1989). Analysis of a large family with autosomal dominant AS showed strong linkage with the COLIVA3/COLIVA4 locus, exclusion of the COLIVA5 gene and segregation of a COLIVA3 haplotype with the disease (Jefferson et al., 1997). In this family, a splice site mutation resulting in the skipping of exon 21 of the COLIVA3 gene has been reported (van der Loop et al., 2000). This mutation exists as a heterozygote with one normal allele present. The splice mutation of the affected gene does not alter the reading frame, but does result in a COLIVA3 chain with an internal deletion. Since the NC-1 domain is intact, this chain may be incorporated into the GBM and its presence may disrupt the intermolecular framework and interfere with the basement membrane architecture.

Goodpasture's syndrome (GS) is an autoimmune disorder that is characterized by circulating IgG antibodies targeting the GBM (Weber, 1993). Although the disease was first identified in 1919 by Dr. Earnest Goodpasture, it was not until 1987 that the target of the circulating antibodies was identified as a novel chain of type IV collagen (Butkowski et al., 1987). In fact, the $\text{colIV}\alpha 3$ molecule was first identified by co-immunoprecipitation with circulating GS antibodies and subsequent biochemical characterization (Saus et al., 1988). The specific epitope of the anti-GBM antibodies in GS has been identified as a cryptic epitope within the NC-1 region of $\text{ColIV}\alpha 3$ molecules (Netzer et al., 1999a). Although no outright cause of GP has been reported, it is possible that this epitope is revealed after exposure to GBM damaging agents including the inhalation of gasoline or acute infections.

Nail patella syndrome (NPS) is a pleiotropic disorder affecting nails, skeletal system, eyes and kidneys in humans (Beals and Eckhardt, 1969). NPS is also known as hereditary onychodysplasia (HOOD), Fong's disease, Turner-Kiesler syndrome or Osterreicher-Turner syndrome. In NPS, renal pathology involves defects in the GBM and the first sign of renal involvement in NPS is usually proteinuria with or without haematuria and is the main cause of death in these patients. (Gubler et al., 1980). GBM changes in NPS include irregular thickening with electron lucent loci resulting in a "moth eaten" appearance. The cause of NPS in humans is associated with defects in the LMX1B gene (Dreyer et al., 1998). The LMX1B gene has been identified as a transcription factor responsible for increased COLIVA3 and COLIVA4 expression that has binding activity in the first exon of the COLIVA4 gene, and LMX1B knockout mice show a decrease in COLIVA3 and COLIVA4 expression (Morello et al., 2001). Transcriptional induction of genes transcribed by LMX1B is required in podocyte differentiation as indicated by LMX1B knockout mice (Miner et al., 2002).

The above studies show that although the COLIVA3, COLIVA4 and COLIVA5 chains are referred to as the "minor" species of collagen, they have important structural roles in the GBM and that disruption of their function results in a wide range of pathologies.

Canine Hereditary Nephritis

Several models for HN have been described with specific indications of GBM involvement. The models that will be discussed have been reported as progressive kidney disorders with end stage renal disease (ESRD) and have other characteristics similar to human AS.

An example of HN has been described in kindred Samoyed dogs (Jansen et al., 1984). In this model, males spontaneously develop renal failure and their GBM architecture has extensive multilamellar splitting when examined by electron microscopy similar to humans diagnosed with AS (Jansen et al., 1986a; Jansen et al., 1986b). Affected male Samoyed dogs have glomerular changes that are associated with increased proteinuria that increases with the age of the dog and terminates in renal failure by 15 months of age. Carrier female Samoyed dogs were identified on the basis of having given birth to affected males. The carrier females do not regularly develop ESRD (although some cases have been reported after five years of age) and GBM structural analysis by electron microscopy show less extensive aberrations with focal lamellation which does not progress over time.

The rate of progression in the Samoyed model was studied using a selective breeding program (Jansen et al., 1986a). In early development, affected males and carrier females appeared healthy. Proteinuria was not detected until 2-3 months of age in either population, but all affected male dogs died by 15 months of age. The

predominance of affected males indicated an X-linked mode of inheritance which was confirmed with a later study (Jansen et al., 1986b).

An interesting study that would foretell many aspects of GBM physiology was conducted to determine the immunohistochemical nature of Samoyed HN (Thorner et al., 1987). This study was conducted without the knowledge of the existence of the minor species of collagen, but these investigators used Goodpasture syndrome (GS) antibodies, along with antibodies directed against laminin, collagen type IV and fibronectin. The only COLIV molecules known at that time were products of the COLIVA1 and COLIVA2 genes. The above antibodies were used in immunohistochemical studies with electron microscopy evaluation to characterize staining patterns in normal adult dogs, neonatal dogs, female carriers of Samoyed HN and male Samoyeds affected with HN. The results for laminin and COLIV antibodies showed strong staining of these products in the GBM in all of the populations above. Fibronectin staining in the GBM was positive for neonatal dogs and affected male Samoyeds, but was focal in carrier females and negative in normal dogs. This is an important finding in that with our current knowledge of GBM components, fibronectin is absent from normal GBM and agrin is the predominant HSPG in mature GBM. GS antibodies did not stain neonatal tissue, only stained in focal areas of carrier female GBM, was greatly reduced in affected males, but did stain normal dog GBM. We now know that the target of GS antibodies is an NC-1 epitope of the colIV α 3 molecule. Also, it is now recognized that the colIV α 3, colIV α 4 and colIV α 5 chains are negative for staining in patients with AS (Nakanishi et al., 1994). These and other studies indicated

that Samoyed HN was a model for X-linked AS in humans (Baumal et al., 1991; Thorner et al., 1989). After the molecular basis of X-linked AS was revealed to be caused by mutations of the COLIVA5 gene on the X chromosome (Barker et al., 1990), an investigation of this gene was directed against this gene in the Samoyed HN model (Zheng et al., 1994). This study was conducted using the known human COLIVA5 sequence and, using these sequences, primers were developed for the amplification, cloning, and sequencing of the canine COLIVA5 gene. The result was the identification of a single G-T transversion that resulted in a premature stop codon in exon 35 of the gene, thus confirming Samoyed HN as a model for human X-linked AS.

A second form of X-linked HN in a mixed breed of dogs has also been described (Lees et al., 1999). Because this kindred is of a mixed breed and originates from Navasota, Texas, this population has been termed NAV dogs. Individuals of this population were referred to the Texas A&M Veterinary Medical Center for evaluation because of kidney failure. The NAV model of HN has many features similar to the Samoyed model above. Pedigree analysis of the NAV dogs indicated an X-linked form of transmission. Male NAV dogs affected with HN exhibit proteinuria at three months of age and usually die before 15 months of age. Carrier females have proteinuria but do not usually progress to renal failure. Ultrastructurally, GBM aberrations of affected NAV dogs include thickening and multilamellar splitting. Since the NAV dogs were of mixed breed origin, migration of the Samoyed allele responsible for HN into the NAV population could not be excluded. The COLIVA5 locus responsible for HN in the Samoyed model was sequenced in affected males and determined not to be responsible

for HN in the NAV population. Thus, a different mutation responsible for HN in NAV dogs was suggested and established NAV HN as a separate model from Samoyed HN.

A form of HN in bull terriers has also been reported (Hood et al., 1995). Proteinuria is progressive and a high content of albumin in the urine strongly suggests involvement of the GBM. Glomerular changes in Bull terriers affected with HN have been reported. Chronic renal disease in this breed ranges from one to eight years of age (Robinson et al., 1989). Unlike other canine models, bull terrier HN is inherited as an autosomal dominant trait (Hood et al., 1990). Almost equal proportions of affected males and female bull terrier HN have been reported. Although autosomal dominant Alport syndrome in one human family has been determined to be caused by a splice site mutation in the COL4A3 gene, immunohistochemical analysis of the GBM in bull terriers affected with HN show the presence of the COLIV α 3 chain (Hood et al., 1995). A similar model for ADAS has been identified in a naturally occurring kindred in Dalmatians (Hood et al., 2002). Defects in the colIV α 4 chain have been implicated in these models.

A form of NH has been described in English cocker spaniel dogs (ECS) that was initially termed familial nephropathy (Robinson et al., 1985; Steward, 1984). These studies were initiated after presentation of clinical cases were referred to small animal practices and included both affected males and females. They described a rapid progression and proteinuria. Although no EM was attempted to characterize the GBM, the urine protein was determined to be of glomerular origin due to the presence of albumin and globulin (Steward, 1984). A study to characterize HN in ECS reported

findings by EM in affected dogs that indicated the extensive thickening and multilamellar splitting of the GBM characteristic of AS in humans (Lees et al., 1997). Although previous studies reported ultrastructural changes in the GBM associated with canine glomerulonephritis, these studies were not specific to HN in ECS (Macdougall et al., 1986; Murray and Wright, 1974). To study the early features and progression of ECS HN, known carriers were identified and two litters were produced by selective breeding (Lees et al., 1998b). The result of this breeding included one litter with a single male and one litter with one male and two females. Two of the dogs from this breeding (both females) developed HN, and the third dog (a single male) was normal at 30 months of age when this study was completed. After onset of proteinuria, loss of kidney function progressed rapidly and all of the affected dogs demonstrated renal lesions by electron microscopy.

Further investigation indicated ECS HN as a model for autosomal recessive AS (Lees et al., 1998b). This study included normal dogs, obligate carrier ECS dogs and affected ECS dogs. Proteinuria was first detected at five to six months of age in affected ECS dogs, however the rate of progression varied in this population. The development of azotemia in affected dogs varied from two to nine months and two dogs that were identified as affected had proteinuria that was significant, but transient. The pattern of immunohistochemical staining, however, was the same for every affected ECS dog. Laminin staining was universal in normal, ECS carriers and in affected ECS dogs. In affected ECS dogs, the GBM staining of the $\text{colIV}\alpha 1$ and $\text{colIV}\alpha 2$ chain was increased, while the $\text{colIV}\alpha 5$ staining was greatly decreased and the $\text{colIV}\alpha 3$ and $\text{colIV}\alpha 4$ chains

were totally absent. By contrast, staining of normal dog GBM showed a predominance of $\text{colIV}\alpha 3$, $\text{colIV}\alpha 4$ and $\text{colIV}\alpha 5$ chains and lesser staining for the $\text{colIV}\alpha 1$ and $\text{colIV}\alpha 2$ chains. Obligate carriers had a decreased $\text{colIV}\alpha 5$ staining. This is the same pattern of expression reported in the GBM of humans diagnosed with autosomal recessive AS (Gubler et al., 1995). Interestingly, the $\text{colIV}\alpha 6$ chain appeared to be faintly expressed in both normal and affected ECS dogs.

The detection of the $\text{colIV}\alpha 6$ chain in the canine GBM prompted further investigation (Lees et al., 2000). Immunolabeling using antibodies monospecific for the $\text{colIV}\alpha 6$ chain showed positive staining in the GBM of dogs less than 30 months of age, but positive in dogs more than 45 months of age. The population of dogs in this study were individuals from many breeds showing that the $\text{colIV}\alpha 6$ chain is not breed dependent. Human subjects were also included in this investigation, but regardless of age (30 years of age to 100 years of age), the $\text{colIV}\alpha 6$ chain was not detected. In humans an isoform switch that occurs during fetal development has been reported (Kalluri et al., 1997). In fetal GBM, the $\text{colIV}\alpha 1$ and $\text{colIV}\alpha 2$ chains are the only colIV isoforms present, but during development, the $\text{colIV}\alpha 3$, $\text{colIV}\alpha 4$ and $\text{colIV}\alpha 5$ chains are produced and, eventually, become the predominant isoform species in the mature GBM. It is suggested that the identification of the $\text{colIV}\alpha 6$ chain in the GBM of older dogs represents a second isoform switch in this species. Because the $\text{colIV}\alpha 6$ chain was detected in the GBM of ECS dogs affected with HN less than 25 months of age, it is also

suggested that the isoform switch can occur at an earlier age in response to glomerular injury (Lees et al., 1998b).

Summary

Six different isoforms of Type IV collagen (colIV α 1-6) have been identified. The individual isoforms of colIV are termed alpha chains and are translated from six different colIV genes (COLIVA1-A6). Collagen Type IV gene products compose the structural framework of basement membranes. The glomerular basement membrane (GBM) is a specialized basement membrane involved in the ultrafiltration processes of the kidney. The colIV α 1- α 5 chains are expressed in the human GBM while the colIV α 1- α 6 chains are expressed in the canine GBM. Many inherited diseases of the kidney have been reported and mutations in genes regulating kidney function have been identified. Alport syndrome (AS) is the most common form of human hereditary nephritis (HN). AS is defined as an inherited progressive kidney disorder associated with sensorineural deafness and is characterized by extensive thickening and multilaminar splitting of the GBM when examined by electron microscopy. AS has both X-linked (XLAS) and autosomal (ARAS) modes of inheritance. Mutations in the COLIVA5 gene are responsible for XLAS. A form of HN with characteristic splitting of the GBM with X-linked inheritance has been described in Samoyed dogs. A specific mutation in the COLIVA5 gene has been identified in Samoyed dogs affected with HN. Mutations in the

COLIVA3 and COLIVA4 genes are responsible for ARAS. A form of HN has been identified in English cocker spaniel dogs (ECS) that has been described as autosomal in inheritance and includes GBM abnormalities including extensive lamination characteristic of ARAS. The HN in ECS has been hypothesized to be a model for human ARAS. As such, we have isolated and characterized canine COLIVA3 and COLIVA4 sequences from normal dogs and ECS dogs affected with HN and compared the coding regions of these candidate genes.

CHAPTER II

THE ISOLATION AND CHARACTERIZATION OF CANINE COLIVA3

Introduction

Collagens are found in all vertebrate species as well as in *Drosophila*, sea urchins and nematodes (Blumberg et al., 1988). As molecules, the collagens are described as triple helical collagenous domains with the amino acid glycine at every third residue that are expressed extracellularly where they perform structural roles. To date, 19 different types of collagen have been identified (Uitto et al., 1986). Six different isoforms of type IV collagen (colIV α 1- α 6) have been described (Hudson et al., 1993). The individual isoforms of colIV are termed alpha chains and are translated from six different genes (COLIVA1-6).

Collagen type IV gene products compose the structural framework of basement membranes. Other basement membrane components include laminin, entactin/nidogen and heparine sulfate proteoglycan (HSPG). The glomerular basement membrane (GBM) is a specialized structure created predominately by epithelial podocytes in the kidney. The GBM is an important structure for the ultrafiltration processes of the kidney.

Hereditary nephritis (HN) refers to a group of inherited glomerular diseases of collagen type IV that often progress to renal failure. In humans, Alport syndrome (AS) is

the most common form of HN (Grunfeld, 1985). About 85% of AS families show X chromosome-linked inheritance (XLAS); the remainder show autosomal recessive (ARAS) or autosomal dominant inheritance (ADAS) (Reeders, 1992).

Several forms of canine HN have been described. HN in the Samoyed breed of dog has been described as a model of XLAS and a mutation in the COL4A5 gene has been identified as causative for this disease (Zheng et al., 1994). In bull terriers, an autosomal dominant form of HN has been reported, although the molecular basis of disease has yet to be described (Hood et al., 1995).

A form of HN has also been described in English cocker spaniel (ECS) dogs (Lees et al., 1998a). ECS HN is characterized as a progressive kidney disease with proteinuria that often results in chronic renal failure. In ECS HN, the GBM has extensive thickening and splitting that increases with the progression of the disease. Humans with ARAS also have extensive thickening and splitting of the GBM. Mutations in the COL4A3 and COL4A4 genes have been identified in individuals with ARAS.

The similarities of the glomerular changes and the autosomal recessive mode of inheritance in both ECS HN and humans with ARAS suggest genetic similarities. As such, we have hypothesized that COL4A3 and COL4A4 are candidate genes in ECS HN. We have isolated and characterized the coding regions of these candidate genes in normal dogs (without HN) and compared these sequences from ECS dogs diagnosed with HN. This chapter describes the isolation and characterization of the COL4A3 coding regions in these populations.

Materials and Methods

Identification of Affected and Normal Populations. Each affected ECS dog met two criteria for the diagnosis of HN. First, the kidney disease exhibited clinicopathologic features identical to those described previously as familial nephropathy in ECS. Second, glomerular examination with transmission electron microscopy (TEM) that exhibited extensive thickening and multilammellar splitting was determined to be pathognomonic of HN. The form of HN in ECS has been determined to be a breed specific disease (Lees et al., 1998b). Three individual dogs older than two years of age from non-ECS breeds were identified for the normal population for this study. The dogs in the normal population included a mixed breed female, a female bloodhound and a male Labrador retriever. ECS dogs are a rare breed in the United States with a registered population of ~5,000 individuals. Because of the rarity and structured breeding practices of the ECS population, it is unlikely that alleles causing ECS-HN would be present in mixed bred populations.

Tissue Isolation. Affected dogs and two normal dogs (a female bloodhound and mixed breed) were sacrificed and ~1g sections of kidneys containing the renal cortex were snap frozen in liquid nitrogen, transported on dry ice and stored at -80°C . Tissue from one normal dog was from a testicle of a Labrador retriever obtained in the College of Veterinary Medicine Texas A&M University from a standard neutering procedure.

RNA Preparation. Total RNA was isolated from frozen tissue using Trizol (Sigma, St Louis, MO) according to manufacturers protocol. Briefly, ~1g of snap frozen tissue was ground in a metal beaker embedded in dry ice and ~100mg was added to 1ml of Trizol in a pyrex test tube. A plastic coated dounce homogenizer was used to macerate the tissue in the tube. The dounce was manipulated manually for approximately one minute at room temperature. To prevent shearing of the RNA, mechanical methods of dounce manipulation were avoided. The Trizol/tissue mixture was transferred to a 1.8ml microcentrifuge tube, allowed to rest at room temperature for ~5 minutes and centrifuged at 1,000xg for one minute to pellet any unsolubilized material. Occasionally this mixture was stored at -80°C, but immediate use is recommended.

To a new 1.8 ml microcentrifuge tube, .5 ml of the Trizol/tissue mixture and 100ul of chloroform was combined. The tube was briefly vortexed (< 5 seconds) and then centrifuged at 10,000xg for 10 minutes. 400ul of the clear upper liquid was transferred to a new 1.8ml microcentrifuge tube and 200ul of isopropanol was added, mixed briefly and centrifuged at 10,000xg for 10 minutes. The isopropanol was removed and the pellet was washed with 1ml of 75% ethanol. After the removal of ethanol, the pellet was resuspended in 50ul of RNase free water and 3ul of RNase inhibitor (Promega, Madison, WI) was added.

A portion of the RNA was run on a 1% agarose gel in TAE buffer and visualized by staining with ethidium bromide for qualitative analysis. For quantitative purposes, a portion of the RNA was also used for spectroscopic analysis at 280A° and 260A°.

cDNA Synthesis. The synthesis of cDNA was performed using the Superscript II first strand synthesis system (Invitrogen, Carlsbad, California) according to manufacturers suggestions. Briefly, 5 ug of total RNA was incubated with an oligo dT primer and 1uL of 10mM dNTP's at 70°C for 5 minutes, chilled on ice, and then combined with 4 uL 5X first strand buffer, 2 uL .1M DTT and 10 units of Superscript reverse transcriptase at 45°C for one hour. The resulting product was used directly in PCR amplification reactions.

PCR Amplification. To assure the accuracy of the amplification process, the enzyme used for PCR amplification of the cDNA was AccuTaq LA DNA Polymerase (Sigma, St. Louis, MO) which has 3' proofreading ability. According to manufacturers suggestions, the amplification reaction contained 5 uL 10X PCR buffer, 2.5 uL 10mM dNTP mix, 600 nmol of each primer, .1 units of AccuTaq LA DNA Polymerase in a 50 uL reaction. As a template, 2 uL of the cDNA was also included. The amplification conditions for all reactions were an initial denaturation at 98°C for 30 seconds followed by repeated denaturation at 94°C for 30 seconds, annealing at 65°C for 30 seconds and a 5 minute extension at 68°C for 35 cycles. A 7 ul sample of the final reaction was analyzed on a 1% agarose gel using TAE buffer to check for the quality of the reaction.

Amplification Primers. Primers were designed from a human COLIVA3 sequence (Genebank Accession: NM000091). Primers for the targeted areas were analyzed using OLIGO 4.0 software (National Biosciences, Plymouth, MN) for dimerization, the

presence of hairpin loops and internal stability. Three sets of primers were used to amplify the canine COLIVA3 cDNA. The NC-1 region was amplified using COLIVA3 NC-1 Forward 5'CTG GAG AAA AAG GAC GTG ATG3' and COLIVA3 NC-1 Reverse 5' CAG CTT TCA CAG TTG ATG GAA TAG5' primers. The 5' region (to ~ 260 bases of the start codon) was amplified using COLIVA3 Forward 5'CCG ACT TGT CGG TGT ACC AGG ATG3' and COLIVA3 Reverse 5'CTC TCC TTC TTC CCC CTT GGC TCC3' primers. The intervening sequence was amplified using the COLIVA3 Forward 5'GTG ATT GGG ATG ATG GGC TAT C3' and COLIVA3 Reverse 5'TCT CCA ATT GGC CCA GGA GTT C3' primers.

Gel Purification. Amplified products were run on a 1% agarose gel and desired fragments were visualized with low intensity ultra violet light. Desired products were cut out of the gel using a clean razor blade and placed into a 1.8 ml microcentrifuge tube. The PCR product was purified using the QIAquick gel purification system (Qiagen, Valencia, CA) and eluted with 50 uL water.

Cloning. The gel purified amplification products were concentrated in a speed vacuum under centrifugation until the 50 uL of eluted product was reduced to approximately 5uL. The gel purified products were cloned into pT7 cloning vectors using perfectly blunt cloning kits (Novagen, Madison, WI) using T4 ligase. The cloned products were transformed into chemically competent cells and plated on LB media containing 50 ug/ml ampicillin, 20 ug/ml tetracycline and 20 ug/ml IPTG that had been overlayed with 20

uL of 50 ug/ml X-gal. The presence of gene fragments in subsequent plasmids was confirmed by the presence of white colonies and restriction digestion using EcoRI and HindIII.

Sequencing. Sequencing reactions contained 4 uL Big Dye V3.0 (Applied Biosystems, Foster City, CA), 400 ng plasmid DNA and 50 nM primer in a 10 uL total reaction volume. The parameters for the sequencing reactions included incubation at 95°C for 10 seconds, 50°C for 5 seconds and 60°C for 4 minutes in a thermocycler for 44 cycles. The resulting reaction was purified on BioMax SPIN-50 mini-columns (BioMax, Odenton, MD). Briefly, the columns were placed in a microcentrifuge at 1000xg for 3 minutes. The columns were transferred to a new 1.8 ml microcentrifuge tube and the sequencing reaction placed on the column. The BioMax spin columns were again centrifuged at 1000xg for 3 minutes. The columns were discarded and the recovered sample was dried in a speed vacuum for 40 minutes at room temperature under darkened conditions. The sequencing reactions were then analyzed in a commercial sequencing facility.

Sequence Analysis. Chromatograms of the resulting sequence were collected and the sequences were downloaded into MacVector V4.0 software (Eastman Kodak Company, Rochester, NY). The sequences were aligned with the human COLIVA3 sequence (NM000091) to determine its relative position with respect to the human isoform. Canine sequences that were confirmed to be of COLIVA3 origin were transferred into AssemblyLine V1.0 software files (Accelrys, Princeton, NJ) for sequence alignment.

Consensus sequences were then transferred to SDSC Biology Workbench (workbench.sdsc.edu) for subsequent analysis.

Results

Sequences representing the coding regions of the normal canine COL4A3 gene have been isolated by PCR amplification from cDNA derived from total RNA (Appendix 1). A total of 4,643 bases have been characterized by sequence analysis representing 92.6% of the human coding sequence. The complete COL4A3 cDNA from the human is 8,050 bases and has a coding region of 5,010 bases (Genbank accession:NM000091). The mouse COL4A3 cDNA has also been characterized and contains 7,765 bases with a 5,010 base coding region (Genbank accession: NM007734). While the remaining ~275 nucleotides of the 5' end of the canine COL4A3 coding region have yet to be characterized, the 4,643 bases represents a majority of this sequence. Nucleic acid alignment using LALINE sequence analysis (workbench.sdsc.edu) show 83.4% homology between the canine COL4A3 and humanCOL4A3 sequences and 80.0% homology between the canine COL4A3 and mouse COL4A3 sequences. The human COL4A3 and mouse COL4A3 sequences share 81.4% homology (Appendix 2).

The canine COL4A3 sequences that have been isolated code for 1,547 amino acids. The human COL4A3 gene codes for 1,671 amino acids and the mouse COL4A3 gene codes for 1,670 amino acids. Using LALINE sequence analysis

(workbench.sdsc.edu), the amino acid sequence homology between the normal canine COL4A3 sequences and the human COL4A3 sequences is 83.4% and 79.8% homologous to the mouse COL4A3 sequences (Appendix 3). The translated coding regions of the human and mouse COL4A3 gene have 78.8% homology.

Interruptions in the collagenous region are characteristic of type IV collagen molecules. The human has 23 interruptions and the mouse has 21 interruptions in this region. Twenty interruptions have been identified in the canine COL4A3 sequences in the collagenous domain. The presence of glycine at every third residue defines the collagenous sequence. Glycine represents 27.3% of the human COL4A3 coding sequences and 27.6% of the mouse COL4A3 coding sequences. The canine COL4A3 coding region contains 27.2% glycine. Proline is another common amino acid in collagenous sequences. Proline represents 19.2% of the human COL4A3 sequences, 20.6% of the mouse COL4A3 sequences and 20.5% of the canine COL4A3 sequences. The codons for glycine are very rich in guanine (GGG, GGA, GGC, GGT) and the codons for proline are very rich in cytosine (CCC, CCT, CCG, CCA). Thus, collagenous sequences are very GC rich. The canine COL4A3 coding region contains 57.1% GC while the human and mouse COL4A3 sequences each contain 56.3% and 57.2% GC respectively.

COL4A3 sequences have also been isolated from ECS dogs diagnosed with HN from cDNA derived from total RNA isolated from fresh frozen kidney. A 4,645 base COL4A3 consensus sequence has been characterized in this affected population representing 1,547 amino acids. The COL4A3 sequences from this population are almost

identical at the nucleotide level. The specific nature of these sequences will be the focus of a later chapter.

CHAPTER III

THE ISOLATION AND CHARACTERIZATION OF CANINE COLIVA4

Introduction

Collagens are found in all vertebrate species as well as in *Drosophila*, sea urchins and nematodes (Blumberg et al., 1988). As molecules, the collagens are described as triple helical collagenous domains with the amino acid glycine at every third residue that are expressed extracellularly where they perform structural roles. To date, 19 different types of collagen have been identified (Uitto et al., 1986). Six different isoforms of type IV collagen (colIV α 1- α 6) have been described (Hudson et al., 1993). The individual isoforms of colIV are termed alpha chains and are translated from six different genes (COLIVA1-6).

Collagen type IV gene products compose the structural framework of basement membranes. Other basement membrane components include laminin, entactin/nidogen and heparine sulfate proteoglycan (HSPG). The glomerular basement membrane (GBM) is a specialized structure created predominately by epithelial podocytes in the kidney. The GBM is an important structure for the ultrafiltration processes of the kidney.

Hereditary nephritis (HN) refers to a group of inherited glomerular diseases of collagen type IV that often progress to renal failure. In humans, Alport syndrome (AS) is

the most common form of HN (Grunfeld, 1985). About 85% of AS families show X chromosome-linked inheritance (XLAS); the remainder show autosomal recessive (ARAS) or autosomal dominant inheritance (ADAS) (Reeders, 1992).

Several forms of canine HN have been described. HN in the Samoyed breed of dog has been described as a model of XLAS and a mutation in the COL4A5 gene has been identified as causative for this disease (Zheng et al., 1994). In bull terriers, an autosomal dominant form of HN has been reported, although the molecular basis of disease has yet to be described (Hood et al., 1995).

A form of HN has also been described in English cocker spaniel (ECS) dogs (Lees et al., 1998a). ECS HN is characterized as a progressive kidney disease with proteinuria that often results in chronic renal failure. In ECS HN, the GBM has extensive thickening and splitting that increases with the progression of the disease. Humans with ARAS also have extensive thickening and splitting of the GBM. Mutations in the COL4A3 and COL4A4 genes have been identified in individuals with ARAS.

The similarities of the glomerular changes and the autosomal recessive mode of inheritance in both ECS HN and humans with ARAS suggest genetic similarities. As such, we have hypothesized that COL4A3 and COL4A4 are candidate genes in ECS HN. We have isolated and characterized the coding regions of these candidate genes in normal dogs (without HN) and compared these sequences from ECS dogs diagnosed with HN. This chapter describes the isolation and characterization of the COL4A4 coding regions in these populations.

Materials and Methods

Identification of Affected and Normal Populations. Each affected ECS dog met two criteria for the diagnosis of HN. First, the kidney disease exhibited clinicopathologic features identical to those described previously as familial nephropathy in ECS. Second, glomerular examination with transmission electron microscopy (TEM) that exhibited extensive thickening and multilammellar splitting was determined to be pathognomonic of HN. The form of HN in ECS has been determined to be a breed specific disease (Lees et al., 1998b). Three individual dogs older than two years of age from non-ECS breeds were identified for the normal population for this study. The dogs in the normal population included a mixed breed female, a female bloodhound and a male Labrador retriever. ECS dogs are a rare breed in the United States with a registered population of ~5,000 individuals. Because of the rarity and structured breeding practices of the ECS population, it is unlikely that alleles causing ECS-HN would be present in mixed bred populations.

Tissue Isolation. Affected dogs and two normal dogs (a female bloodhound and mixed breed) were sacrificed and ~1g sections of kidneys containing the renal cortex were snap frozen in liquid nitrogen, transported on dry ice and stored at -80°C . Tissue from one normal dog was from a testicle of a Labrador retriever obtained in the College of Veterinary Medicine at Texas A&M University from a standard neutering procedure.

RNA Preparation. Total RNA was isolated from frozen tissue using Trizol (Sigma, St Louis, MO) according to manufacturers protocol. Briefly, ~1g of snap frozen tissue was ground in a metal beaker embedded in dry ice and ~100mg was added to 1ml of Trizol in a pyrex test tube. A plastic coated dounce homogenizer was used to macerate the tissue in the tube. The dounce was manipulated manually for approximately one minute at room temperature. To prevent shearing of the RNA, mechanical methods of dounce manipulation were avoided. The Trizol/tissue mixture was transferred to a 1.8ml microcentrifuge tube, allowed to rest at room temperature for ~5 minutes and centrifuged at 1,000xg for one minute to pellet any unsolubilized material. Occasionally this mixture was stored at -80°C, but immediate use is recommended.

To a new 1.8 ml microcentrifuge tube, .5 ml of the Trizol/tissue mixture and 100ul of chloroform was combined. The tube was briefly vortexed (< 5 seconds) and then centrifuged at 10,000xg for 10 minutes. 400ul of the clear upper liquid was transferred to a new 1.8ml microcentrifuge tube and 200ul of isopropanol was added, mixed briefly and centrifuged at 10,000xg for 10 minutes. The isopropanol was removed and the pellet was washed with 1ml of 75% ethanol. After the removal of ethanol, the pellet was resuspended in 50ul of RNase free water and 3ul of RNase inhibitor (Promega, Madison, WI) was added.

A portion of the RNA was run on a 1% agarose gel in TAE buffer and visualized by staining with ethidium bromide for qualitative analysis. For quantitative purposes, a portion of the RNA was also used for spectroscopic analysis at 280A° and 260A°.

cDNA Synthesis. The synthesis of cDNA was performed using the Superscript II first strand synthesis system (Invitrogen, Carlsbad, California) according to manufacturers suggestions. Briefly, 5 ug of total RNA was incubated with an oligo dT primer and 1uL of 10mM dNTP's at 70°C for 5 minutes, chilled on ice, and then combined with 4 uL 5X first strand buffer, 2 uL .1M DTT and 10 units of Superscript reverse transcriptase at 45°C for one hour. The resulting product was used directly in PCR amplification reactions.

PCR Amplification. To assure the accuracy of the amplification process, the enzyme used for PCR amplification of the cDNA was AccuTaq LA DNA Polymerase (Sigma, St. Louis, MO) which has 3' proofreading ability. According to manufacturers suggestions, the amplification reaction contained 5 uL 10X PCR buffer, 2.5 uL 10mM dNTP mix, 600 nmol of each primer, .1 units of AccuTaq LA DNA Polymerase in a 50 uL reaction. As a template, 2 uL of the cDNA was also included. The amplification conditions for all reactions were an initial denaturation at 98° C for 30 seconds followed by repeated denaturation at 94°C for 30 seconds, annealing at 65°C for 30 seconds and a 5 minute extension at 68°C for 35 cycles. A 7 ul sample of the final reaction was analyzed on a 1% agarose gel using TAE buffer to check for the quality of the reaction.

Amplification Primers. Primers were designed from a human COLIVA4 sequence (Genebank Accession: NM000092). Primers for the targeted areas were analyzed using OLIGO 4.0 software (National Biosciences, Plymouth, MN) for dimerization, the

presence of hairpin loops and internal stability. Three sets of primers were used to amplify the canine COLIVA4 cDNA. The NC-1 region was amplified using COLIVA4 NC-1 Forward 5'TTG GCC CTG GAT ACC TCA GTG3' and COLIVA4 NC-1 Reverse 5'GGC ACC TGC TGA TTT TCT GG3' primers. The 5' region (to -140 bases of the start codon in the human) was amplified using COLIVA4 Forward 5'TGA CCC AGA ACA CAG AAC CTC AC3' and COLIVA4 Reverse 5'CCC TTC TCT CCT TTT GGT CCA ATT C3' primers. The intervening sequence was amplified using the COLIVA4 Forward 5'AGA GGC GCT CCA GGG ATA C3' and COLIVA4 Reverse 5'CCT TCT GGT CCT CTT G3' primers for the 2,155 base region adjacent to the NC-1. The 1,590 base region adjacent to the 5' sequence was amplified with the COL4A4 F 5'GGT CTC CGG GTT CCT ATG3' AND COL4A4 R 5'CTG GCG GCC CAA GAA G3' primers.

Gel Purification. Amplified products were run on a 1% agarose gel and desired fragments were visualized with low intensity ultra violet light. Desired products were cut out of the gel using a clean razor blade and placed into a 1.8 ml microcentrifuge tube. The PCR product was purified using the QIAquick gel purification system (Qiagen, Valencia, CA) and eluted with 50 uL water.

Cloning. The gel purified amplification products were concentrated in a speed vacuum under centrifugation until the 50 uL of eluted product was reduced to approximately 5 uL. The gel purified products were cloned into pT7 cloning vectors using perfectly blunt cloning kits (Novagen, Madison, WI) using T4 ligase. The cloned products were

transformed into chemically competent cells and plated on LB media containing 50 ug/ml ampicillin, 20 ug/ml tetracycline and 20 ug/ml IPTG that had been overlayed with 20 uL of 50 ug/ml X-gal. The presence of gene fragments in subsequent plasmids was confirmed by the presence of white colonies and restriction digestion using EcoRI and HindIII.

Sequencing. Sequencing reactions contained 4 uL Big Dye V3.0 (Applied Biosystems, Foster City, CA), 400 ng plasmid DNA and 50 nM primer in a 10 uL total reaction volume. The parameters for the sequencing reactions included incubation at 95°C for 10 seconds, 50°C for 5 seconds and 60°C for 4 minutes in a thermocycler for 44 cycles. The resulting reaction was purified on BioMax SPIN-50 mini-columns (BioMax, Odenton, MD). Briefly, the columns were placed in a microcentrifuge at 1,000xg for 3 minutes. The columns were transferred to a new 1.8 ml microcentrifuge tube and the sequencing reaction placed on the column. The BioMax spin columns were again centrifuged at 1,000xg for 3 minutes. The columns were discarded and the recovered sample was dried in a speed vacuum for 40 minutes at room temperature under darkened conditions. The sequencing reactions were then analyzed in a commercial sequencing facility.

Sequence Analysis. Chromatograms of the resulting sequence were collected and the sequences were downloaded into MacVector V4.0 software (Eastman Kodak Company, Rochester, NY). The sequences were aligned with the human COLIVA4 sequence (NM000092) to determine its relative position with respect to the human isoform.

Canine sequences that were confirmed to be of COLIVA3 origin were transferred into AssemblyLine V1.0 software files (Accelrys, Princeton, NJ) for sequence alignment. Consensus sequences were then transferred to SDSC Biology Workbench (workbench.sdsc.edu) for subsequent analysis.

Results

Sequences representing the coding regions of the normal canine COLIVA4 gene have been isolated by PCR amplification from cDNA derived from total RNA (Appendix 5). A total of 5,163 bases have been characterized by sequence analysis representing ~98% of the human coding sequence. The complete COL4A4 cDNA from the human is 5,726 bases and has a coding region of 5,073 bases (Genbank accession:NM000092). The mouse COL4A4 cDNA has also been characterized and contains 5,918 bases with a coding region of 5,049 bases (Genbank accession: AF169338). While the remaining ~50 nucleotides of the 3' end of the canine COL4A4 coding region have yet to be characterized, the 5,163 bases represents a majority of this sequence. Nucleic acid alignment using LALINE sequence analysis (workbench.sdsc.edu) show 85.0% homology between the canine COL4A4 and human COL4A4 sequences and 79.0% homology between the canine COL4A4 and mouse COL4A4 sequences. The human COL4A4 and mouse COL4A4 sequences share 80.7% homology (Appendix 6).

The canine COL4A4 sequences that have been isolated code for 1,677 amino acids. The human COL4A4 gene codes for 1,691 amino acids and the mouse COL4A4 gene codes for 1,683 amino acids. Using LALINE sequence analysis (workbench.sdsc.edu), the amino acid sequence homology between the normal canine COL4A4 sequences and the human COL4A4 sequences is 83.2% and 78.3% homologous to the mouse COL4A4 sequences (Appendix 7). The translated coding regions of the human and mouse COL4A4 gene have 78.6% homology.

Interruptions in the collagenous region are characteristic of type IV collagen molecules. The human has 24 interruptions and the mouse has 23 interruptions in this region. In the canine COL4A4 sequences, 23 interruptions have been identified in the collagenous domain. The presence of glycine at every third residue defines the collagenous sequence. Glycine represents 28.0% of the human COL4A4 coding sequences and 27.5% of the mouse COL4A4 coding sequences. The canine COL4A4 coding region contains 27.8% glycine. Proline is another common amino acid in collagenous sequences. Proline represents 20.8% of the human COL4A4 sequences, 20.9% of the mouse COL4A4 sequences and 21.2% of the canine COL4A4 sequences. The codons for glycine are very rich in guanine (GGG, GGA, GGC, GGT) and the codons for proline are very rich in cytosine (CCC, CCT, CCG, CCA). Thus, collagenous sequences are very GC rich. The canine COL4A4 coding region contains 60.5% GC while the human and mouse COL4A4 sequences each contain 59.3% and 59.7% GC respectively.

COL4A4 sequences have also been isolated from ECS dogs diagnosed with HN from cDNA derived from total RNA isolated from fresh frozen kidney. A 5,011 base COL4A4 consensus sequence has been characterized in this affected population representing 1,607 amino acids. The COL4A4 sequences from this population are almost identical at the nucleotide level. The specific nature of these sequences will be the focus of a later chapter.

CHAPTER II

SUMMARY AND CONCLUSIONS

The problematic nature of the identification of mutations in ECS HN can be derived from studies that have attempted to find mutations in the COL4A3 and COL4A4 genes in ARAS. While the detection rate of mutations in the COL4A5 coding region in XLAS has reached ~90%, it has been reported that perhaps 10% of the mutations are in non-coding regions of the CL4A5 gene (Martin et al., 1998). The 90% detection rate of mutations in the COL4A5 gene in XLAS does not represent the current rate of detection of COL4A3 and COL4A4 mutations in ARAS. In the investigation of the COL4A3 or COL4A4 genes in ARAS, the success rate of finding mutations in these genes individually is very low. When both the COL4A3 and COL4A4 genes have been analyzed together in ARAS, the identification of causal mutations is increased, but does not reach 100% detection in these genes. Together, two studies that analyzed both the COL4A3 and COL4A4 genes in ARAS have shown less than a 30% rate of detection of mutations in these genes. This low rate of detection may represent the limitations in the methodology of exon screening which, when applied to the COL4A5 gene in XLAS, has an equally low rate of detection.

In both studies, diagnosis of AS had been made including the splitting of the GBM, and exclusion of the COL4A5 gene had been determined. Although selective

criteria were used to diagnose ARAS in these studies, this methodology represents a more permissive basis of diagnosis as compared to the strict interpretation of ECS-HN. In one study (Mochizuki et al., 1994), mutations in the COL4A3 and COL4A4 genes in 4 of 7 families was reported. These included two mutations in the COL4A3 gene (both nonsense mutations) and two mutations in the COL4A4 gene (one glycine substitution and one nonsense mutation). In another study (Longo et al., 2002), 8 mutations were identified in 30 probands. This study included six probands with ADAS presumed to be of COL4A3 or COL4A4 origin. While Mochizuki et al. reported an equal number of COL4A3 and COL4A4 mutations, Longo et al. reported seven COL4A3 mutations (two nonsense mutations, three microdeletions a glycine substitution, and a missense at the start codon) and only one COL4A4 mutation (one glycine substitution). The combined detection rate in these studies for mutations in either the COL4A3 or COL4A4 genes is 12/37. This low rate of detection suggests that either mutations in the COL4A3 and COL4A4 genes in ARAS are difficult to identify or, possibly, the presence of another genetic locus responsible for this pathology.

Indication of the latter possibility is represented in a study of haplotype segregation in thin basement membrane disease (TBMD) (Buzza et al., 2001b). While it is possible to perform linkage analysis of haplotype segregation of the COL4A3/COL4A4 locus, it is difficult to differentiate segregation patterns to any one of these loci. In both man and mouse, the COL4A3 and COL4A4 genes lie head to head where they share a common promoter and therefore can be considered to be examples of very close linkage. Haplotype segregation of genomic microsatellites of the

COL4A3/COL4A4 locus and the COL4A5 locus were studied in the families of 22 individuals diagnosed with TBMD. In this study, 35% of the families showed linkage to the COL4A3/COL4A4 locus, 18% showed linkage to the COL4A5 locus, but 45% of the families failed to show linkage to either locus. TBMD is often inherited as an autosomal dominant trait and, as the name indicates, is characterized by a thinned GBM (Blumenthal et al., 1988). Heterozygous carriers for causal mutations in ARAS (Lemmink et al., 1996) and female carriers of XLAS (Rumpelt et al., 1974) have clinopathologic symptoms identical to those of TBMD including a thinning of the GBM. It is suggested that TBMD represents a carrier state for ARAS or XLAS (Netzer et al., 1999b). Incomplete penetrance or improper diagnosis may have affected the lack of segregation of either the COL4A3, COL4A4 or COL4A5 genes with TBMD. These results do not preclude, however, the presence of another locus responsible for TBMD: a recognized disease of the GBM. The possibility of another genetic locus responsible for ARAS might be resolved with similar haplotype analysis to determine the frequency of segregation of the COL4A3/COL4A4 locus in these families. Similarly, haplotype analysis could help to determine COL4A3/COL4A4 segregation in ECS HN.

Implication of either the COL4A3 or COL4A4 gene in ECS HN by analysis of haplotype segregation in our current study is very difficult. The experimental design in this study was to determine sequences in alleles not responsible for ECS HN and compare these sequences in ECS-HN populations. Optimally, haplotype analysis would be conducted in individuals of the same breed. This is possible in the analysis of the X-linked HN in the Samoyed breed. In this case, it is possible to positively identify affected

males, normal males and carrier females. Similar haplotype analysis is complicated in our study because of the presumed autosomal nature of ECS HN. It is presumed that affected individuals are homozygous recessive in ECS HN populations. Because the total ECS population in the United States is less than 5,000 individuals, there exists a possibility that any one individual may be a heterozygous carrier of ECS HN alleles. ECS HN has been reported to be breed specific. No other autosomal recessive HN has been reported in any other canine populations. For these reasons, non-ECS populations were selected for the absence of ECS HN alleles. Therefore, haplotype analysis in this study may represent breed specific ECS haplotypes and not represent markers specific for ECS HN. Segregation of haplotypes is often used for the analysis of quantitative trait loci (QTL). The experimental design for these studies involving haplotype segregation is very different from the approach of our study. Haplotype analysis of QTL does not frequently use genetic elements derived from cDNA that has been cloned.

While the vast majority of the COL4A3 and COL4A4 coding regions have been identified, it is possible that mutations exist in other areas yet to be characterized. These include ~250 bases at the 5' end and ~30 bases at the 3' end of the COL4A3 coding region, and ~50 bases at the 5' end and ~50 bases at the 3' end of the COL4A4 coding region. Other regions of interest include promoter elements, 5' and 3' UTRs, exon/intron junctions and intronic sequences.

Several differences have been identified between the normal and ECS-HN COL4A3 sequences. The effects of these individual nucleotide changes with regards to codon changes are summarized in Appendix 9. Several differences have also been

identified between the normal and ECS-HN COL4A4 sequences. The effects of these individual nucleotide changes with regards to codon changes are summarized in Appendix 10.

The most significant differences between normal canine populations and ECS-HN populations in the COL4A3 and COL4A4 sequences are two base changes in the COL4A3 allele. One difference that has been identified is a G-A transition that results in an amino acid change at position ~3,250 (See Appendix 7). While the codon for the normal sequence is AGG and codes for arginine, the G-A transition results in an AAG codon that codes for lysine. The sequence difference has been identified in every affected ECS examined (N=3) from 4 different clones and was present in each of 14 sequence reactions. This transition was identified in two normal non-ECS individuals from 4 clones in 7 sequencing reactions. Another significant difference identified in the COL4A3 sequences is another single base change at position ~3,220 (See Appendix 7). While the normal sequence is AGC and codes for serine, a G-T transversion has been identified that changes the codon in ECS alleles to ATC to code for isoleucine. This transversion was identified in three ECS-HN individuals from 4 different clones and was present in each of 12 sequencing reactions. The normal sequence has been confirmed in two normal individuals from four clones in 6 sequencing reactions. It is difficult to determine if these differences are causal mutations for ECS HN. If identification of heterozygosity in known carriers in these regions could be determined between normal and affected populations, these markers could be very informative. Based on these sequences, primers have been designed from regions flanking areas of interest to amplify

these regions from genomic DNA (gDNA) isolated from circulating lymphocytes for the identification of heterozygous alleles at this locus. The process has been difficult because the primary gDNA sequence of these genes has not been determined. Based on corresponding human sequences, both the COL4A3 and COL4A4 genes should each cover over 150 kb each and contain approximately 50 introns. Because the position and size of introns and exons are unknown for the canine genes, it is difficult to design parameters for amplification reactions. Although this does not preclude the possibility of isolating these sequences from gDNA, the process has proven difficult. An alternative approach to identify heterozygosity in these loci from known carrier populations is to isolate these sequences from the coding regions of these genes. This could include the amplification, cloning and sequencing of these regions from cDNA derived from total RNA. Because of the difficulties involved in the specific amplification of collagen genes, direct sequencing of amplification products with the primers that have been designed has proven difficult. Once the primary sequence of the gDNA has been determined, more specific primers for these areas of interest can be synthesized for direct sequencing of gDNA amplification reactions. Since both alleles may be equally expressed, it is possible to identify the two alleles by sequencing cloned sequences from products amplified from cDNA. Using the binomial equation $([N!/X!(N-X)!](P_x)^x(P_{N-x})^{N-x})$, greater than 95% detection of heterozygous loci can be determined when 5 clones are analyzed from products of individual amplification reactions.

The question as to the origin of the ECS-HN allele is an interesting topic for discussion. Clinically, ECS-HN is not indicated by spontaneous mutation, but rather by

an established genetic aberration in the germ line of this population. ECS-HN has been reported to date to be a breed specific pathology indicating that the ECS-HN allele is not present in other purebred populations and, within the ECS breed, is restricted to “party colored” individuals. Purebred canine populations exist as groups indicating a common origin. Examples of purebred groups include hounds, terriers and retrievers among others. Examples of purebred spaniel populations include the ECS as well as the American cocker spaniel, the Brittany spaniel, and Springer spaniel. No indication of the ECS-HN allele, however, is present in these populations. It is difficult to determine whether the ECS-HN allele was present during the establishment of the ECS population or originated during ECS breed development. If the ECS-HN allele was present during the establishment of the population, then this causal mutation might have also been present in other populations. If the ECS-HN allele was present in other populations, especially other spaniel populations, many factors could have caused its disappearance in non-ECS breeds. Selective forces could have eliminated the allele in non-ECS populations since the ECS-HN allele causes a decreased fitness. However, the perpetual nature of the ECS-HN allele in ECS populations could be due to the recessive nature of the allele that would mask its presence in heterozygous populations making selection difficult. Because these populations are small and because inbreeding tends toward genetic homozygosity, the ECS-HN allele could also have been lost by genetic drift. Alternatively, the ECS-HN allele could have been introduced in the evolution of this population.

Recessive alleles could be perpetuated through breeding practices inherent in the establishment and growth of purebred canine populations. Most purebred canine populations arise from a small number of individuals. Often there is a major sire that may be bred to many females. This practice can lead to a high frequency of alleles from a small portion of the breeding population. In this case, the frequency of the ECS-HN allele may have been amplified in this population by the founder effect. This would suggest that the ECS-HN allele, by whatever means of introduction, would be identical by descent. In pure genetic terms, identity by decent infers loss of all genetic heterogeneity to create identical alleles. This does not necessarily have to be a strict definition. Identity by decent could also infer homozygosity at one locus causing a particular phenotype, but genetic polymorphism at other allelic loci arising from subsequent mutations. The COL4A3 and COL4A4 sequences we have characterized from ECS-HN populations are very homozygous as are the sequences generated from non-ECS populations. Decreased homozygosity is expected in small-inbred populations. The polymorphisms that have been identified in the ECS-HN populations do not preclude, however, homozygosity at the ECS-HN locus. Although the ECS population has been recently established (less than 500 years ago), many hundreds of generations may have occurred thus creating the opportunity for the introduction of polymorphisms. The theory of the molecular clock suggests a constant rate of molecular evolution. Although studies on the molecular clock frequently focus on populations separated genetically over extreme time periods, it is not inconceivable that a causal mutations for ECS-HN may have been introduced into the population's, albeit, brief history.

It is difficult to establish the transitional change that substitutes an arginine for a lysine as the causal mutation in ECS-HN. This change does not result in a nonsense mutation, does not occur at an essential structural element such as glycine and does not drastically effect a proline residue adjacent to a glycine or substitute a non-conserved amino acid. The neutral theory of molecular evolution, first established by Kimura, suggests that most mutations are neutral. The neutrality theory is indicated at two levels. First, that genetic mutations often do not change the amino acid (due to the wobble effect), and, second, when mutations do change the coding sequence they often substitute neutral changes that do not effect protein structure or function. The theory is based on the three types of mutations, namely, deleterious mutations, neutral mutations and advantageous mutations. Deleterious mutations are frequently eliminated from populations, advantageous mutations are exceedingly rare, therefore, most mutations perpetuated in a population are neutral. Kimura's neutral theory of molecular evolution has recently been reformulated with the tremendous amount of sequence data that has been derived from investigation into specific genetic systems and genome projects.

Many supposedly neutral mutations have been reevaluated as functional polymorphisms. Polymorphic analysis working with the "relative clinical observed likelihood" (RCOL) has established the relevance of polymorphisms with regards to clinical relevance (Cooper, 2002). Clinically relevant polymorphisms include a change in a functional amino acid, an evolutionarily conserved amino acid, a biochemically non-conserved amino acid and reversion of phenotype with substitution. Although the arginine-lysine substitution does not meet many of the criteria, it does confer with other

characteristics of clinically relevant phenotypes. It segregates with the clinical phenotype, is present in all individuals tested and is not present in two normal dogs from two different breeds and substitutes a conserved amino acid from both mouse and man. The serine-isoleucine substitution also segregates with the affected phenotype but may be more clinically significant. The substitution changes a hydrophilic amino acid for a hydrophobic amino acid. Although not evolutionarily conserved in the human where the corresponding amino acid is an asparagine (another hydrophilic amino acid), it is conserved in the corresponding position in the mouse.

Folding of the $\alpha 1(\text{IV})$ molecules into a trimer for incorporation into the basement membrane network is essential for $\alpha 1(\text{IV})$ function. $\alpha 1(\text{IV})$ trimer assembly begins at the C-terminus and proceeds to the N-terminus. Any disruption in the winding together of these chains can disrupt the normal molecular assembly.

the spectrum of COL4A5 mutations is represented in an investigation of the natural history of 195 XLAS kindreds (Jais et al., 2000). Of the 195 COL4A5 mutations reported, ~20% were large rearrangements, ~15% were microinsertions or microdeletions, ~27% were nonsense mutations and ~15% were splice mutations. Missense mutations were found in 74/195 kindreds and represented ~38% of all mutations in this study. Although the majority of the missense mutations (59/74) were glycine substitutions, 16/74 of the missense mutations were substitutions that involved amino acids other than glycine.

An earlier investigation described many of these non-glycine substitutions in XLAS (Lemmink et al., 1997). Non-glycine substitutions in the COL4A5 gene in this

study were identified as causal mutations in 11/160 kindreds. Two non-glycine substitutions changed a conserved arginine. Six mutations were identified as hydrophobic to hydrophilic changes and three of these changed an isoleucine to a serine. The serine to isoleucine substitution in the COL4A3 gene identified in ECS-HN individuals is a hydrophilic to hydrophobic change, but this report shows that changes in the hydrophobicity of individual amino acids in the COL4A5 gene have been shown to create the XLAS phenotype. Similar changes have yet to be shown in ARAS, but this may be because of the scarcity of reported mutations in these COL4A3 and COL4A4 genes.

While it is more difficult to establish causality of amino acid substitutions on phenotype, we propose that the aberrations identified in the COL4A3 coding region in ECS-HN individuals inhibits the incorporation of the $\alpha 1(\text{IV})$ chain into mature $\alpha 1(\text{IV})$ trimers. The winding of these molecules could be disrupted by a kink in a collagenous region resulting from a substitution of a hydrophilic amino acid (serine) with a hydrophobic amino acid (isoleucine). The serine-isoleucine substitution lies adjacent to a glycine molecule and it is possible that the change to a hydrophobic amino acid causes a kink in the chain inhibiting the folding of the glycine into the $\alpha 1(\text{IV})$ helix. Each amino acid interacts with other amino acids in corresponding chains. It is now impossible to determine the intramolecular associations between the $\alpha 1(\text{IV})$ chains at this specific locus, however, the substitution of a larger hydrophobic amino acid at this position may inhibit the folding of these chains into a mature functional trimer.

Another factor inhibiting the winding of the $\text{colIV}\alpha 3\alpha 4\alpha 5$ trimer is the substitution of the conserved arginine near the substituted serine. For whatever reason, this colIVA3 arginine is conserved in mouse and man. These two mutations may act synergistically to inhibit the molecular interactions necessary for trimer assembly.

The ultimate goal of molecular medicine is to be able to predict a clinical phenotype associated with a specific genotype. The identification of a specific base change in ECS COL4A3 and COL4A4 alleles, especially with regards to an amino acid change, may provide such a foundation for the molecular basis of ECS-HN.

REFERENCES

- Alport, C.A. 1927. Hereditary Familial Congenital haemorrhagic nephritis. *The British Medical Journal*. 1:504-506.
- Atkin, C.L. 1988. Diseases of the Kidney. Little, Brown, Boston. 617-641 pp.
- Atkin, C.L., S.J. Hasstedt, L. Menlove, L. Cannon, N. Kirschner, C. Schwartz, K. Nguyen, and M. Skolnick. 1988. Mapping of Alport syndrome to the long arm of the X chromosome. *American Journal of Human Genetics*. 42:249-55.
- Aumailley, M., H. Wiedemann, K. Mann, and R. Timpl. 1989. Binding of nidogen and the laminin-nidogen complex to basement membrane collagen type IV. *European Journal of Biochemistry*. 184:241-8.
- Barker, D.F., S.L. Hostikka, J. Zhou, L.T. Chow, A.R. Oliphant, S.C. Gerken, M.C. Gregory, M.H. Skolnick, C.L. Atkin, and K. Tryggvason. 1990. Identification of mutations in the COL4A5 collagen gene in Alport syndrome. *Science*. 248:1224-7.
- Battaglia, C., U. Mayer, M. Aumailley, and R. Timpl. 1992. Basement-membrane heparan sulfate proteoglycan binds to laminin by its heparan sulfate chains and to nidogen by sites in the protein core. *European Journal of Biochemistry*. 208:359-66.
- Baumal, R., P. Thorner, V.E. Valli, R. McInnes, P. Marrano, R. Jacobs, A. Binnington, and A.G. Bloedow. 1991. Renal disease in carrier female dogs with X-linked hereditary nephritis. Implications for female patients with this disease. *American Journal of Pathology*. 139:751-64.
- Beals, R.K., and A.L. Eckhardt. 1969. Hereditary onycho-osteodysplasia (Nail-Patella syndrome). A report of nine kindreds. *Journal of Bone & Joint Surgery - American Volume*. 51:505-16.
- Belkin, A.M., and M.A. Stepp. 2000. Integrins as receptors for laminins. *Microscopy Research & Technique*. 51:280-301.
- Bellamy, G., and P. Bornstein. 1971. Evidence for procollagen, a biosynthetic precursors of collagen. *Proceedings of the National Academy of Sciences of the United States of America*. 68:1138-42.

- Blumberg, B., A.J. MacKrell, and J.H. Fessler. 1988. Drosophila basement membrane procollagen alpha 1(IV). II. Complete cDNA sequence, genomic structure, and general implications for supramolecular assemblies. *Journal of Biological Chemistry*. 263:18328-37.
- Blumenthal, S.S., C. Fritsche, and J. Lemann, Jr. 1988. Establishing the diagnosis of benign familial hematuria. The importance of examining the urine sediment of family members. *JAMA*. 259:2263-6.
- Boute, N., O. Gribouval, S. Roselli, F. Benessy, H. Lee, A. Fuchshuber, K. Dahan, M.C. Gubler, P. Niaudet, and C. Antignac. 2000. NPHS2, encoding the glomerular protein podocin, is mutated in autosomal recessive steroid-resistant nephrotic syndrome.[erratum appears in *Nature Genetics* 2000 May;25(1):125]. *Nature Genetics*. 24:349-54.
- Boye, E., G. Mollet, L. Forestier, L. Cohen-Solal, L. Heidet, P. Cochat, J.P. Grunfeld, J.B. Palcoux, M.C. Gubler, and C. Antignac. 1998. Determination of the genomic structure of the COL4A4 gene and of novel mutations causing autosomal recessive Alport syndrome. *American Journal of Human Genetics*. 63:1329-40.
- Boye, E., D. Vetrie, F. Flinter, B. Buckle, T. Pihlajaniemi, E.R. Hamalainen, J.C. Myers, M. Bobrow, and A. Harris. 1991. Major rearrangements in the alpha 5(IV) collagen gene in three patients with Alport syndrome. *Genomics*. 11:1125-32.
- Brazel, D., I. Oberbaumer, H. Dieringer, W. Babel, R.W. Glanville, R. Deutzmann, and K. Kuhn. 1987. Completion of the amino acid sequence of the alpha 1 chain of human basement membrane collagen (type IV) reveals 21 non-triplet interruptions located within the collagenous domain. *European Journal of Biochemistry*. 168:529-36.
- Brazel, D., R. Pollner, I. Oberbaumer, and K. Kuhn. 1988. Human basement membrane collagen (type IV). The amino acid sequence of the alpha 2(IV) chain and its comparison with the alpha 1(IV) chain reveals deletions in the alpha 1(IV) chain. *European Journal of Biochemistry*. 172:35-42.
- Burgeson, R.E., M. Chiquet, R. Deutzmann, P. Ekblom, J. Engel, H. Kleinman, G.R. Martin, G. Meneguzzi, M. Paulsson, and J. Sanes. 1994. A new nomenclature for the laminins. *Matrix Biology*. 14:209-11.
- Butkowski, R.J., J.P. Langeveld, J. Wieslander, J. Hamilton, and B.G. Hudson. 1987. Localization of the Goodpasture epitope to a novel chain of basement membrane collagen. *Journal of Biological Chemistry*. 262:7874-7.

- Butkowski, R.J., J. Wieslander, M. Kleppel, A.F. Michael, and A.J. Fish. 1989. Basement membrane collagen in the kidney: regional localization of novel chains related to collagen IV. *Kidney International*. 35:195-202.
- Buzza, M., Y.Y. Wang, H. Dagher, J.J. Babon, R.G. Cotton, H. Powell, J. Dowling, and J. Savage. 2001a. COL4A4 mutation in thin basement membrane disease previously described in Alport syndrome.[comment]. *Kidney International*. 60:480-3.
- Buzza, M., D. Wilson, and J. Savage. 2001b. Segregation of hematuria in thin basement membrane disease with haplotypes at the loci for Alport syndrome. *Kidney International*. 59:1670-6.
- Carlin, B., R. Jaffe, B. Bender, and A.E. Chung. 1981. Entactin, a novel basal lamina-associated sulfated glycoprotein. *Journal of Biological Chemistry*. 256:5209-14.
- Caulfield, J.P., and M.G. Farquhar. 1974. The permeability of glomerular capillaries to graded dextrans. Identification of the basement membrane as the primary filtration barrier. *Journal of Cell Biology*. 63:883-903.
- Caulfield, J.P., and M.G. Farquhar. 1978. Loss of anionic sites from the glomerular basement membrane in aminonucleoside nephrosis. *Laboratory Investigation*. 39:505-12.
- Chance, J.K., and J.A. Stanley. 1977. Alport's syndrome: case report and review of ocular manifestations. *Annals of Ophthalmology*. 9:1527-30.
- Chang, R.L., W.M. Deen, C.R. Robertson, and B.M. Brenner. 1975. Permselectivity of the glomerular capillary wall: III. Restricted transport of polyanions. *Kidney International*. 8:212-8.
- Chu, M.L., W. de Wet, M. Bernard, J.F. Ding, M. Morabito, J. Myers, C. Williams, and F. Ramirez. 1984. Human pro alpha 1(I) collagen gene structure reveals evolutionary conservation of a pattern of introns and exons. *Nature*. 310:337-40.
- Churg, J., and R.L. Sherman. 1973. Pathologic characteristics of hereditary nephritis. *Archives of Pathology & Laboratory Medicine*. 95:374-9.
- Cochat, P., P. Guibaud, R. Garcia Torres, B. Roussel, V. Guarner, and F. Larbre. 1988. Diffuse leiomyomatosis in Alport syndrome. *Journal of Pediatrics*. 113:339-43.
- Cooper, D.N. 2002. Human gene mutation in pathology and evolution. *Journal of Inherited Metabolic Disease*. 25:157-82.

- Cotman, S.L., W. Halfter, and G.J. Cole. 1999. Identification of extracellular matrix ligands for the heparan sulfate proteoglycan agrin. *Experimental Cell Research*. 249:54-64.
- Denzer, A.J., R. Brandenberger, M. Gesemann, M. Chiquet, and M.A. Ruegg. 1997. Agrin binds to the nerve-muscle basal lamina via laminin. *Journal of Cell Biology*. 137:671-83.
- Denzer, A.J., M. Gesemann, B. Schumacher, and M.A. Ruegg. 1995. An amino-terminal extension is required for the secretion of chick agrin and its binding to extracellular matrix. *Journal of Cell Biology*. 131:1547-60.
- Denzer, A.J., T. Schulthess, C. Fauser, B. Schumacher, R.A. Kammerer, J. Engel, and M.A. Ruegg. 1998. Electron microscopic structure of agrin and mapping of its binding site in laminin-1. *EMBO Journal*. 17:335-43.
- DeSimone, D.W., M.A. Stepp, R.S. Patel, and R.O. Hynes. 1987. The integrin family of cell surface receptors. *Biochemical Society Transactions*. 15:789-91.
- Ding, J., J. Stitzel, P. Berry, E. Hawkins, and C.E. Kashtan. 1995. Autosomal recessive Alport syndrome: mutation in the COL4A3 gene in a woman with Alport syndrome and posttransplant antglomerular basement membrane nephritis. *Journal of the American Society of Nephrology*. 5:1714-7.
- Dixit, S.N., J.M. Stuart, J.M. Seyer, J. Risteli, R. Timpl, and A.H. Kang. 1981. Type IV collagens' isolation and characterization of 7S collagen from human kidney, liver and lung. *Collagen & Related Research*. 1:549-56.
- Dolz, R., J. Engel, and K. Kuhn. 1988. Folding of collagen IV. *European Journal of Biochemistry*. 178:357-66.
- Dreyer, S.D., G. Zhou, A. Baldini, A. Winterpacht, B. Zabel, W. Cole, R.L. Johnson, and B. Lee. 1998. Mutations in LMX1B cause abnormal skeletal patterning and renal dysplasia in nail patella syndrome. *Nature Genetics*. 19:47-50.
- Durkin, M.E., S. Chakravarti, B.B. Bartos, S.H. Liu, R.L. Friedman, and A.E. Chung. 1988. Amino acid sequence and domain structure of entactin. Homology with epidermal growth factor precursor and low density lipoprotein receptor. *Journal of Cell Biology*. 107:2749-56.
- Dziadek, M., and R. Timpl. 1985. Expression of nidogen and laminin in basement membranes during mouse embryogenesis and in teratocarcinoma cells. *Developmental Biology*. 111:372-82.

- Epstein, C.J., M.A. Sahud, C.F. Piel, J.R. Goodman, M.R. Bernfield, J.H. Kushner, and A.R. Ablin. 1972. Hereditary macrothrombocytopathia, nephritis and deafness. *American Journal of Medicine*. 52:299-310.
- Erickson, A.C., and J.R. Couchman. 2000. Still more complexity in mammalian basement membranes. *Journal of Histochemistry & Cytochemistry*. 48:1291-306.
- Farquhar, M.G., S.L. Wissig, and G.E. Palade. 1961. Glomerular permeability: I. Ferritin transfer across the normal glomerular capillary wall. *Journal of Experimental Medicine*. 113:47-66.
- Feingold, J., E. Bois, A. Chompret, M. Broyer, M.C. Gubler, and J.P. Grunfeld. 1985. Genetic heterogeneity of Alport syndrome. *Kidney International*. 27:672-7.
- Fox, J.W., U. Mayer, R. Nischt, M. Aumailley, D. Reinhardt, H. Wiedemann, K. Mann, R. Timpl, T. Krieg, and J. Engel. 1991. Recombinant nidogen consists of three globular domains and mediates binding of laminin to collagen type IV. *EMBO Journal*. 10:3137-46.
- Fuchshuber, A., G. Jean, O. Gribouval, M.C. Gubler, M. Broyer, J.S. Beckmann, P. Niaudet, and C. Antignac. 1995. Mapping a gene (SRN1) to chromosome 1q25-q31 in idiopathic nephrotic syndrome confirms a distinct entity of autosomal recessive nephrosis. *Human Molecular Genetics*. 4:2155-8.
- Gautam, M., P.G. Noakes, L. Moscoso, F. Rupp, R.H. Scheller, J.P. Merlie, and J.R. Sanes. 1996. Defective neuromuscular synaptogenesis in agrin-deficient mutant mice. *Cell*. 85:525-35.
- Gauthier, B., H. Trachtman, R. Frank, and E. Valderrama. 1989. Familial thin basement membrane nephropathy in children with asymptomatic microhematuria. *Nephron*. 51:502-8.
- Govan, J.A. 1983. Ocular manifestations of Alport's syndrome: a hereditary disorder of basement membranes? *British Journal of Ophthalmology*. 67:493-503.
- Graham, R.C., Jr., and M.J. Karnovsky. 1966. Glomerular permeability. Ultrastructural cytochemical studies using peroxidases as protein tracers. *Journal of Experimental Medicine*. 124:1123-34.
- Grant, D.S., C.P. Leblond, H.K. Kleinman, S. Inoue, and J.R. Hassell. 1989. The incubation of laminin, collagen IV, and heparan sulfate proteoglycan at 35 degrees C yields basement membrane-like structures. *Journal of Cell Biology*. 108:1567-74.

- Griffin, C.A., B.S. Emanuel, J.R. Hansen, W.K. Cavenee, and J.C. Myers. 1987. Human collagen genes encoding basement membrane alpha 1 (IV) and alpha 2 (IV) chains map to the distal long arm of chromosome 13. *Proceedings of the National Academy of Sciences of the United States of America*. 84:512-6.
- Groffen, A.J., F.W. Hop, K. Tryggvason, H. Dijkman, K.J. Assmann, J.H. Veerkamp, L.A. Monnens, and L.P. Van den Heuvel. 1997. Evidence for the existence of multiple heparan sulfate proteoglycans in the human glomerular basement membrane and mesangial matrix. *European Journal of Biochemistry*. 247:175-82.
- Groffen, A.J., M.A. Ruegg, H. Dijkman, T.J. van de Velden, C.A. Buskens, J. van den Born, K.J. Assmann, L.A. Monnens, J.H. Veerkamp, and L.P. van den Heuvel. 1998. Agrin is a major heparan sulfate proteoglycan in the human glomerular basement membrane. *Journal of Histochemistry & Cytochemistry*. 46:19-27.
- Grunfeld, J.P. 1985. The clinical spectrum of hereditary nephritis. *Kidney International*. 27:83-92.
- Gubler, M., M. Levy, M. Broyer, C. Naizot, G. Gonzales, D. Perrin, and R. Habib. 1981. Alport's syndrome. A report of 58 cases and a review of the literature. *American Journal of Medicine*. 70:493-505.
- Gubler, M.C., B. Knebelmann, A. Beziau, M. Broyer, Y. Pirson, F. Haddoum, M.M. Kleppel, and C. Antignac. 1995. Autosomal recessive Alport syndrome: immunohistochemical study of type IV collagen chain distribution. *Kidney International*. 47:1142-7.
- Gubler, M.C., M. Levy, C. Naizot, and R. Habib. 1980. Glomerular basement membrane changes in hereditary glomerular diseases. *Renal Physiology*. 3:405-13.
- Gunwar, S., F. Ballester, M.E. Noelken, Y. Sado, Y. Ninomiya, and B.G. Hudson. 1998. Glomerular basement membrane. Identification of a novel disulfide-cross-linked network of alpha3, alpha4, and alpha5 chains of type IV collagen and its implications for the pathogenesis of Alport syndrome. *Journal of Biological Chemistry*. 273:8767-75.
- Guyton, A.C. 1991. Textbook of Medical Physiology. W.B. Saunders, New York.
- Halfter, W., S. Dong, B. Schurer, and G.J. Cole. 1998. Collagen XVIII is a basement membrane heparan sulfate proteoglycan. *Journal of Biological Chemistry*. 273:25404-12.

- Haralson, M.A., S.J. Federspiel, A. Martinez-Hernandez, R.K. Rhodes, and E.J. Miller. 1985. Synthesis of [pro alpha 1(IV)]3 collagen molecules by cultured embryo-derived parietal yolk sac cells. *Biochemistry*. 24:5792-7.
- Heidet, L., C. Arrondel, L. Forestier, L. Cohen-Solal, G. Mollet, B. Gutierrez, C. Stavrou, M.C. Gubler, and C. Antignac. 2001. Structure of the human type IV collagen gene COL4A3 and mutations in autosomal Alport syndrome. *Journal of the American Society of Nephrology*. 12:97-106.
- Heptinstall, R.H. 1992. Pathology of the Kidney. Little, Brown, Boston.
- Hinglais, N., J.P. Grunfeld, and E. Bois. 1972. Characteristic ultrastructural lesion of the glomerular basement membrane in progressive hereditary nephritis (Alport's syndrome). *Laboratory Investigation*. 27:473-87.
- Hofmann, H., T. Voss, K. Kuhn, and J. Engel. 1984. Localization of flexible sites in thread-like molecules from electron micrographs. Comparison of interstitial, basement membrane and intima collagens. *Journal of Molecular Biology*. 172:325-43.
- Holzman, L.B., P.L. St John, I.A. Kovari, R. Verma, H. Holthofer, and D.R. Abrahamson. 1999. Nephrin localizes to the slit pore of the glomerular epithelial cell. *Kidney International*. 56:1481-91.
- Hood, J.C., C. Huxtable, I. Naito, C. Smith, R. Sinclair, and J. Savige. 2002. A novel model of autosomal dominant Alport syndrome in Dalmatian dogs. *Nephrology Dialysis Transplantation*. 17:2094-8.
- Hood, J.C., W.F. Robinson, C.R. Huxtable, J.S. Bradley, R.J. Sutherland, and M.A. Thomas. 1990. Hereditary nephritis in the bull terrier: evidence for inheritance by an autosomal dominant gene. *Veterinary Record*. 126:456-9.
- Hood, J.C., J. Savige, A. Hendtlass, M.M. Kleppel, C.R. Huxtable, and W.F. Robinson. 1995. Bull terrier hereditary nephritis: a model for autosomal dominant Alport syndrome. *Kidney International*. 47:758-65.
- Hostikka, S.L., R.L. Eddy, M.G. Byers, M. Hoyhtya, T.B. Shows, and K. Tryggvason. 1990. Identification of a distinct type IV collagen alpha chain with restricted kidney distribution and assignment of its gene to the locus of X chromosome-linked Alport syndrome. *Proceedings of the National Academy of Sciences of the United States of America*. 87:1606-10.

- Hostikka, S.L., and K. Tryggvason. 1988. The complete primary structure of the alpha 2 chain of human type IV collagen and comparison with the alpha 1(IV) chain. *Journal of Biological Chemistry*. 263:19488-93.
- Hudson, B.G., S.T. Reeders, and K. Tryggvason. 1993. Type IV collagen: structure, gene organization, and role in human diseases. Molecular basis of Goodpasture and Alport syndromes and diffuse leiomyomatosis. *Journal of Biological Chemistry*. 268:26033-6.
- Humphries, M.J. 2000. Integrin structure. *Biochemical Society Transactions*. 28:311-39.
- Ichikawa, I., and A. Fogo. 1996. Focal segmental glomerulosclerosis.[comment]. *Pediatric Nephrology*. 10:374-91.
- Jais, J.P., B. Knebelmann, I. Giatras, M. De Marchi, G. Rizzoni, A. Renieri, M. Weber, O. Gross, K.O. Netzer, F. Flinter, Y. Pirson, C. Verellen, J. Wieslander, U. Persson, K. Tryggvason, P. Martin, J.M. Hertz, C. Schroder, M. Sanak, S. Krejcova, M.F. Carvalho, J. Saus, C. Antignac, H. Smeets, and M.C. Gubler. 2000. X-linked Alport syndrome: natural history in 195 families and genotype-phenotype correlations in males. *Journal of the American Society of Nephrology*. 11:649-57.
- Jansen, B., P. Thorner, R. Baumas, V. Valli, M.G. Maxie, and A. Singh. 1986a. Samoyed hereditary glomerulopathy (SHG). Evolution of splitting of glomerular capillary basement membranes. *American Journal of Pathology*. 125:536-45.
- Jansen, B., P.S. Thorner, A. Singh, J.M. Patterson, J.H. Lumsden, V.E. Valli, R. Baumas, and R.K. Basrur. 1984. Animal model of human disease: hereditary nephritis in Samoyed dogs. *American Journal of Pathology*. 116:175-8.
- Jansen, B., L. Tryphonas, J. Wong, P. Thorner, M.G. Maxie, V.E. Valli, R. Baumas, and P.K. Basrur. 1986b. Mode of inheritance of Samoyed hereditary glomerulopathy: an animal model for hereditary nephritis in humans. *Journal of Laboratory & Clinical Medicine*. 107:551-5.
- Jefferson, J.A., H.H. Lemmink, A.E. Hughes, C.M. Hill, H.J. Smeets, C.C. Doherty, and A.P. Maxwell. 1997. Autosomal dominant Alport syndrome linked to the type IV collagen alpha 3 and alpha 4 genes (COL4A3 and COL4A4). *Nephrology Dialysis Transplantation*. 12:1595-9.
- Johansson, C., R. Butkowski, and J. Wieslander. 1992. The structural organization of type IV collagen. Identification of three NC1 populations in the glomerular basement membrane. *Journal of Biological Chemistry*. 267:24533-7.

- Kahsai, T.Z., G.C. Enders, S. Gunwar, C. Brunmark, J. Wieslander, R. Kalluri, J. Zhou, M.E. Noelken, and B.G. Hudson. 1997. Seminiferous tubule basement membrane. Composition and organization of type IV collagen chains, and the linkage of alpha3(IV) and alpha5(IV) chains. *Journal of Biological Chemistry*. 272:17023-32.
- Kallunki, P., R.L. Eddy, M.G. Byers, M. Kestila, T.B. Shows, and K. Tryggvason. 1991. Cloning of human heparan sulfate proteoglycan core protein, assignment of the gene (HSPG2) to 1p36.1----p35 and identification of a BamHI restriction fragment length polymorphism. *Genomics*. 11:389-96.
- Kalluri, R., C.F. Shield, P. Todd, B.G. Hudson, and E.G. Neilson. 1997. Isoform switching of type IV collagen is developmentally arrested in X-linked Alport syndrome leading to increased susceptibility of renal basement membranes to endoproteolysis. *Journal of Clinical Investigation*. 99:2470-8.
- Kamagata, Y., M.G. Mattei, and Y. Ninomiya. 1992. Isolation and sequencing of cDNAs and genomic DNAs encoding the alpha 4 chain of basement membrane collagen type IV and assignment of the gene to the distal long arm of human chromosome 2. *Journal of Biological Chemistry*. 267:23753-8.
- Kanwar, Y.S., and M.G. Farquhar. 1979a. Anionic sites in the glomerular basement membrane. In vivo and in vitro localization to the laminae rarae by cationic probes. *Journal of Cell Biology*. 81:137-53.
- Kanwar, Y.S., and M.G. Farquhar. 1979b. Presence of heparan sulfate in the glomerular basement membrane. *Proceedings of the National Academy of Sciences of the United States of America*. 76:1303-7.
- Kao, W.W., D.J. Prockop, and R.A. Berg. 1979. Kinetics for the secretion of nonhelical procollagen by freshly isolated tendon cells. *Journal of Biological Chemistry*. 254:2234-43.
- Kaplan, J.M., S.H. Kim, K.N. North, H. Rennke, L.A. Correia, H.Q. Tong, B.J. Mathis, J.C. Rodriguez-Perez, P.G. Allen, A.H. Beggs, and M.R. Pollak. 2000. Mutations in ACTN4, encoding alpha-actinin-4, cause familial focal segmental glomerulosclerosis. *Nature Genetics*. 24:251-6.
- Kashtan, C.E., M.M. Kleppel, and M.C. Gubler. 1996. Immunohistologic findings in Alport syndrome. *Contributions to Nephrology*. 117:142-53.
- Kawai, S., S. Nomura, T. Harano, K. Harano, T. Fukushima, and G. Osawa. 1996. The COL4A5 gene in Japanese Alport syndrome patients: spectrum of mutations of all exons. The Japanese Alport Network. *Kidney International*. 49:814-22.

- Kefalides, N.A. 1971. Isolation of a collagen from basement membranes containing three identical - chains. *Biochemical & Biophysical Research Communications*. 45:226-34.
- Kestila, M., U. Lenkkeri, M. Mannikko, J. Lamerdin, P. McCready, H. Putaala, V. Ruotsalainen, T. Morita, M. Nissinen, R. Herva, C.E. Kashtan, L. Peltonen, C. Holmberg, A. Olsen, and K. Tryggvason. 1998. Positionally cloned gene for a novel glomerular protein--nephrin--is mutated in congenital nephrotic syndrome. *Molecular Cell*. 1:575-82.
- Kimura, N., T. Toyoshima, T. Kojima, and M. Shimane. 1998. Entactin-2: a new member of basement membrane protein with high homology to entactin/nidogen. *Experimental Cell Research*. 241:36-45.
- Kivirikko, K.I., and R. Myllyla. 1987. Recent developments in posttranslational modification: intracellular processing. *Methods in Enzymology*. 144:96-114.
- Kleinman, H.K., M.L. McGarvey, J.R. Hassell, V.L. Star, F.B. Cannon, G.W. Laurie, and G.R. Martin. 1986. Basement membrane complexes with biological activity. *Biochemistry*. 25:312-8.
- Kleppel, M.M., W.W. Fan, H.I. Cheong, C.E. Kashtan, and A.F. Michael. 1992. Immunochemical studies of the Alport antigen. *Kidney International*. 41:1629-37.
- Kleppel, M.M., C. Kashtan, P.A. Santi, J. Wieslander, and A.F. Michael. 1989a. Distribution of familial nephritis antigen in normal tissue and renal basement membranes of patients with homozygous and heterozygous Alport familial nephritis. Relationship of familial nephritis and Goodpasture antigens to novel collagen chains and type IV collagen. *Laboratory Investigation*. 61:278-89.
- Kleppel, M.M., P.A. Santi, J.D. Cameron, J. Wieslander, and A.F. Michael. 1989b. Human tissue distribution of novel basement membrane collagen. *American Journal of Pathology*. 134:813-25.
- Kohfeldt, E., T. Sasaki, W. Gohring, and R. Timpl. 1998. Nidogen-2: a new basement membrane protein with diverse binding properties. *Journal of Molecular Biology*. 282:99-109.
- Kuhn, K., H. Wiedemann, R. Timpl, J. Risteli, H. Dieringer, T. Voss, and R.W. Glanville. 1981. Macromolecular structure of basement membrane collagens. *FEBS Letters*. 125:123-8.

- Layman, D.L., E.B. McGoodwin, and G.R. Martin. 1971. The nature of the collagen synthesized by cultured human fibroblasts. *Proceedings of the National Academy of Sciences of the United States of America*. 68:454-8.
- Leblond, C.P., and S. Inoue. 1989. Structure, composition, and assembly of basement membrane. *American Journal of Anatomy*. 185:367-90.
- Lees, G.E., R.G. Helman, L.D. Homco, N.J. Millichamp, J.F. Hunter, and M.S. Frey. 1998a. Early diagnosis of familial nephropathy in english cocker spaniels. *Journal of the American Animal Hospital Association*. 34:189-195.
- Lees, G.E., R.G. Helman, C.E. Kashtan, A.F. Michael, L.D. Homco, N.J. Millichamp, Z.T. Camacho, J.W. Templeton, Y. Ninomiya, Y. Sado, I. Naito, and Y. Kim. 1999. New form of X-linked dominant hereditary nephritis in dogs. *American Journal of Veterinary Research*. 60:373-83.
- Lees, G.E., R.G. Helman, C.E. Kashtan, A.F. Michael, L.D. Homco, N.J. Millichamp, Y. Ninomiya, Y. Sado, I. Naito, and Y. Kim. 1998b. A Model of autosomal recessive alport-syndrome in english cocker spaniel dogs. *Kidney International*. 54:706-719.
- Lees, G.E., C.E. Kashtan, A.F. Michael, R.G. Helman, I. Naito, Y. Ninomiya, Y. Sado, and Y. Kim. 2000. Expression of the alpha6 chain of type IV collagen in glomerular basement membranes of healthy adult dogs. *American Journal of Veterinary Research*. 61:38-41.
- Lees, G.E., P.D. Wilson, R.G. Helman, L.D. Homco, and M.S. Frey. 1997. Glomerular ultrastructural findings similar to hereditary nephritis in 4 English cocker spaniels. *Journal of Veterinary Internal Medicine*. 11:80-5.
- Leinonen, A., M. Mariyama, T. Mochizuki, K. Tryggvason, and S.T. Reenders. 1994. Complete primary structure of the human type IV collagen alpha 4(IV) chain. Comparison with structure and expression of the other alpha (IV) chains. *Journal of Biological Chemistry*. 269:26172-7.
- Lemmink, H.H., T. Mochizuki, L.P. van den Heuvel, C.H. Schroder, A. Barrientos, L.A. Monnens, B.A. van Oost, H.G. Brunner, S.T. Reenders, and H.J. Smeets. 1994. Mutations in the type IV collagen alpha 3 (COL4A3) gene in autosomal recessive Alport syndrome. *Human Molecular Genetics*. 3:1269-73.
- Lemmink, H.H., W.N. Nillesen, T. Mochizuki, C.H. Schroder, H.G. Brunner, B.A. van Oost, L.A. Monnens, and H.J. Smeets. 1996. Benign familial hematuria due to mutation of the type IV collagen alpha4 gene. *Journal of Clinical Investigation*. 98:1114-8.

- Lemmink, H.H., C.H. Schroder, L.A. Monnens, and H.J. Smeets. 1997. The clinical spectrum of type IV collagen mutations. *Human Mutation*. 9:477-99.
- Lenkkeri, U., M. Mannikko, P. McCready, J. Lamerdin, O. Gribouval, P.M. Niaudet, C.K. Antignac, C.E. Kashtan, C. Homberg, A. Olsen, M. Kestila, and K. Tryggvason. 1999. Structure of the gene for congenital nephrotic syndrome of the finnish type (NPHS1) and characterization of mutations. *American Journal of Human Genetics*. 64:51-61.
- Longo, I., P. Porcedda, F. Mari, D. Giachino, I. Meloni, C. Deplano, A. Brusco, M. Bosio, L. Massella, G. Lavoratti, D. Roccatello, G. Frasca, G. Mazzucco, A.O. Muda, M. Conti, F. Fasciolo, C. Arrondel, L. Heidet, A. Renieri, and M. De Marchi. 2002. COL4A3/COL4A4 mutations: from familial hematuria to autosomal-dominant or recessive Alport syndrome. *Kidney International*. 61:1947-56.
- Macdougall, D.F., T. Cook, A.P. Steward, and V. Cattell. 1986. Canine chronic renal disease: prevalence and types of glomerulonephritis in the dog. *Kidney International*. 29:1144-51.
- Mann, K., R. Deutzmann, M. Aumailley, R. Timpl, L. Raimondi, Y. Yamada, T.C. Pan, D. Conway, and M.L. Chu. 1989. Amino acid sequence of mouse nidogen, a multidomain basement membrane protein with binding activity for laminin, collagen IV and cells. *EMBO Journal*. 8:65-72.
- Mann, K., R. Deutzmann, and R. Timpl. 1988. Characterization of proteolytic fragments of the laminin-nidogen complex and their activity in ligand-binding assays. *European Journal of Biochemistry*. 178:71-80.
- Mariyama, M., A. Leinonen, T. Mochizuki, K. Tryggvason, and S.T. Reenders. 1994. Complete primary structure of the human alpha 3(IV) collagen chain. Coexpression of the alpha 3(IV) and alpha 4(IV) collagen chains in human tissues. *Journal of Biological Chemistry*. 269:23013-7.
- Mariyama, M., K. Zheng, T.L. Yang-Feng, and S.T. Reenders. 1992. Colocalization of the genes for the alpha 3(IV) and alpha 4(IV) chains of type IV collagen to chromosome 2 bands q35-q37. *Genomics*. 13:809-13.
- Martin, P., N. Heiskari, H. Pajari, C. Gronhagen-Riska, H. Kaariainen, O. Koskimies, and K. Tryggvason. 2000. Spectrum of COL4A5 mutations in Finnish Alport syndrome patients. *Human Mutation*. 15:579.

- Martin, P., N. Heiskari, J. Zhou, A. Leinonen, T. Tumelius, J.M. Hertz, D. Barker, M. Gregory, C. Atkin, U. Styrkarsdottir, H. Neumann, J. Springate, T. Shows, E. Pettersson, and K. Tryggvason. 1998. High mutation detection rate in the COL4A5 collagen gene in suspected Alport syndrome using PCR and direct DNA sequencing. *Journal of the American Society of Nephrology*. 9:2291-301.
- Martin, P.T., and J.R. Sanes. 1997. Integrins mediate adhesion to agrin and modulate agrin signaling. *Development*. 124:3909-17.
- Mayer, U., R. Nischt, E. Poschl, K. Mann, K. Fukuda, M. Gerl, Y. Yamada, and R. Timpl. 1993. A single EGF-like motif of laminin is responsible for high affinity nidogen binding. *EMBO Journal*. 12:1879-85.
- Miller, E.J., E.H. Epstein, Jr., and K.A. Piez. 1971. Identification of three genetically distinct collagens by cyanogen bromide cleavage of insoluble human skin and cartilage collagen. *Biochemical & Biophysical Research Communications*. 42:1024-9.
- Miller, E.J., and V.J. Matukas. 1969. Chick cartilage collagen: a new type of alpha 1 chain not present in bone or skin of the species. *Proceedings of the National Academy of Sciences of the United States of America*. 64:1264-8.
- Miner, J.H., J. Cunningham, and J.R. Sanes. 1998. Roles for laminin in embryogenesis: exencephaly, syndactyly, and placentopathy in mice lacking the laminin alpha5 chain. *Journal of Cell Biology*. 143:1713-23.
- Miner, J.H., and C. Li. 2000. Defective glomerulogenesis in the absence of laminin alpha5 demonstrates a developmental role for the kidney glomerular basement membrane. *Developmental Biology*. 217:278-89.
- Miner, J.H., R. Morello, K.L. Andrews, C. Li, C. Antignac, A.S. Shaw, and B. Lee. 2002. Transcriptional induction of slit diaphragm genes by Lmx1b is required in podocyte differentiation. *Journal of Clinical Investigation*. 109:1065-72.
- Miner, J.H., B.L. Patton, S.I. Lentz, D.J. Gilbert, W.D. Snider, N.A. Jenkins, N.G. Copeland, and J.R. Sanes. 1997. The laminin alpha chains: expression, developmental transitions, and chromosomal locations of alpha1-5, identification of heterotrimeric laminins 8-11, and cloning of a novel alpha3 isoform. *Journal of Cell Biology*. 137:685-701.
- Mochizuki, T., H.H. Lemmink, M. Mariyama, C. Antignac, M.C. Gubler, Y. Pirson, C. Verellen-Dumoulin, B. Chan, C.H. Schroder, and H.J. Smeets. 1994. Identification of mutations in the alpha 3(IV) and alpha 4(IV) collagen genes in autosomal recessive Alport syndrome. *Nature Genetics*. 8:77-81.

- Moeller, M.J., I.A. Kovari, and L.B. Holzman. 2000. Evaluation of a new tool for exploring podocyte biology: mouse *Nphs1* 5' flanking region drives LacZ expression in podocytes. *Journal of the American Society of Nephrology*. 11:2306-14.
- Morello, R., G. Zhou, S.D. Dreyer, S.J. Harvey, Y. Ninomiya, P.S. Thorner, J.H. Miner, W. Cole, A. Winterpacht, B. Zabel, K.C. Oberg, and B. Lee. 2001. Regulation of glomerular basement membrane collagen expression by LMX1B contributes to renal disease in nail patella syndrome. *Nature Genetics*. 27:205-8.
- Morrison, K.E., M. Mariyama, T.L. Yang-Feng, and S.T. Reeders. 1991. Sequence and localization of a partial cDNA encoding the human alpha 3 chain of type IV collagen. *American Journal of Human Genetics*. 49:545-54.
- Muragaki, Y., S. Timmons, C.M. Griffith, S.P. Oh, B. Fadel, T. Quertermous, and B.R. Olsen. 1995. Mouse *Col18a1* is expressed in a tissue-specific manner as three alternative variants and is localized in basement membrane zones. *Proceedings of the National Academy of Sciences of the United States of America*. 92:8763-7.
- Murray, M., and N.C. Wright. 1974. A morphologic study of canine glomerulonephritis. *Laboratory Investigation*. 30:213-21.
- Nagayoshi, T., D. Sanborn, N.J. Hickok, D.R. Olsen, M.J. Fazio, M.L. Chu, R. Knowlton, K. Mann, R. Deutzmann, and R. Timpl. 1989. Human nidogen: complete amino acid sequence and structural domains deduced from cDNAs, and evidence for polymorphism of the gene. *DNA*. 8:581-94.
- Nakanishi, K., N. Yoshikawa, K. Iijima, K. Kitagawa, H. Nakamura, H. Ito, K. Yoshioka, M. Kagawa, and Y. Sado. 1994. Immunohistochemical study of alpha 1-5 chains of type IV collagen in hereditary nephritis. *Kidney International*. 46:1413-21.
- Netzer, K.O., A. Leinonen, A. Boutaud, D.B. Borza, P. Todd, S. Gunwar, J.P. Langeveld, and B.G. Hudson. 1999a. The goodpasture autoantigen. Mapping the major conformational epitope(s) of alpha3(IV) collagen to residues 17-31 and 127-141 of the NC1 domain. *Journal of Biological Chemistry*. 274:11267-74.
- Netzer, K.O., S. Seibold, and M. Weber. 1999b. Thin basement membrane--do we have a window for understanding the molecular pathogenesis? *Nephrology Dialysis Transplantation*. 14:1060-1.
- Nielsen, C.E. 1978. Lenticonus anterior and Alport's syndrome. *Acta Ophthalmologica*. 56:518-30.

- Nitkin, R.M., M.A. Smith, C. Magill, J.R. Fallon, Y.M. Yao, B.G. Wallace, and U.J. McMahan. 1987. Identification of agrin, a synaptic organizing protein from Torpedo electric organ. *Journal of Cell Biology*. 105:2471-8.
- Noakes, P.G., J.H. Miner, M. Gautam, J.M. Cunningham, J.R. Sanes, and J.P. Merlie. 1995. The renal glomerulus of mice lacking s-laminin/laminin beta 2: nephrosis despite molecular compensation by laminin beta 1. *Nature Genetics*. 10:400-6.
- Oh, S.P., Y. Kamagata, Y. Muragaki, S. Timmons, A. Ooshima, and B.R. Olsen. 1994. Isolation and sequencing of cDNAs for proteins with multiple domains of Gly-Xaa-Yaa repeats identify a distinct family of collagenous proteins. *Proceedings of the National Academy of Sciences of the United States of America*. 91:4229-33.
- Olsen, D.R., T. Nagayoshi, M. Fazio, M.G. Mattei, E. Passage, D. Weil, R. Timpl, M.L. Chu, and J. Uitto. 1989. Human nidogen: cDNA cloning, cellular expression, and mapping of the gene to chromosome 1q43. *American Journal of Human Genetics*. 44:876-85.
- Oohashi, T., M. Sugimoto, M.G. Mattei, and Y. Ninomiya. 1994. Identification of a new collagen IV chain, alpha 6(IV), by cDNA isolation and assignment of the gene to chromosome Xq22, which is the same locus for COL4A5. *Journal of Biological Chemistry*. 269:7520-6.
- Ozen, S., D. Ertoyl, L. Heidet, L. Cohen-Solal, H. Ozen, N. Besbas, A. Bakkaoglu, and C. Antignac. 2001. Benign familial hematuria associated with a novel COL4A4 mutation. *Pediatric Nephrology*. 16:874-7.
- Paulsson, M., P.D. Yurchenco, G.C. Ruben, J. Engel, and R. Timpl. 1987. Structure of low density heparan sulfate proteoglycan isolated from a mouse tumor basement membrane. *Journal of Molecular Biology*. 197:297-313.
- Pease, D.C., and R.F. Baker. 1950. Electron microscopy of the kidney. *American Journal of Anatomy*. 87:349-389.
- Peissel, B., L. Geng, R. Kalluri, C. Kashtan, H.G. Rennke, G.R. Gallo, K. Yoshioka, M.J. Sun, B.G. Hudson, and E.G. Neilson. 1995. Comparative distribution of the alpha 1(IV), alpha 5(IV), and alpha 6(IV) collagen chains in normal human adult and fetal tissues and in kidneys from X-linked Alport syndrome patients. *Journal of Clinical Investigation*. 96:1948-57.
- Piez, K.A., Eigner, E. A., and Lewis, M.S. 1963. The chromatographic separation and amino acid composition of several collagens. *biochemistry*. 2:58-66.

- Pihlajaniemi, T., E.R. Pohjola, and J.C. Myers. 1990. Complete primary structure of the triple-helical region and the carboxyl-terminal domain of a new type IV collagen chain, alpha 5(IV). *Journal of Biological Chemistry*. 265:13758-66.
- Pinnell, S.R., S.M. Krane, J.E. Kenzora, and M.J. Glimcher. 1972. A heritable disorder of connective tissue. Hydroxylysine-deficient collagen disease. *New England Journal of Medicine*. 286:1013-20.
- Pinnell, S.R., and G.R. Martin. 1968. The cross-linking of collagen and elastin: enzymatic conversion of lysine in peptide linkage to alpha-aminoadipic-delta-semialdehyde (allysine) by an extract from bone. *Proceedings of the National Academy of Sciences of the United States of America*. 61:708-16.
- Pochet, J.M., G. Bobrie, P. Landais, B. Goldfarb, and J.P. Grunfeld. 1989. Renal prognosis in Alport's and related syndromes: influence of the mode of inheritance.[comment]. *Nephrology Dialysis Transplantation*. 4:1016-21.
- Poschl, E., R. Pollner, and K. Kuhn. 1988. The genes for the alpha 1(IV) and alpha 2(IV) chains of human basement membrane collagen type IV are arranged head-to-head and separated by a bidirectional promoter of unique structure. *EMBO Journal*. 7:2687-95.
- Prockop, D.J., and K.I. Kivirikko. 1995. Collagens: molecular biology, diseases, and potentials for therapy. *Annual Review of Biochemistry*. 64:403-34.
- Putaala, H., K. Sainio, H. Sariola, and K. Tryggvason. 2000. Primary structure of mouse and rat nephrin cDNA and structure and expression of the mouse gene. *Journal of the American Society of Nephrology*. 11:991-1001.
- Putaala, H., R. Soininen, P. Kilpelainen, J. Wartiovaara, and K. Tryggvason. 2001. The murine nephrin gene is specifically expressed in kidney, brain and pancreas: inactivation of the gene leads to massive proteinuria and neonatal death. *Human Molecular Genetics*. 10:1-8.
- Raats, C.J., M.A. Bakker, W. Hoch, W.P. Tamboer, A.J. Groffen, L.P. van den Heuvel, J.H. Berden, and J. van den Born. 1998. Differential expression of agrin in renal basement membranes as revealed by domain-specific antibodies. *Journal of Biological Chemistry*. 273:17832-8.
- Ramachandran, G.N. 1954. Structure of collagen. *Nature*. 174:269-270.
- Ramachandran, G.N. 1955. Structure of collagen. *Nature*. 176:593-595.

- Rambourg, A., and C.P. Leblond. 1967. Staining of basement membranes and associated structures by the periodic acid-Schiff and periodicacid-silver methenamine techniques. *Journal of Ultrastructure Research*. 20:306-9.
- Reeders, S.T. 1992. Molecular genetics of hereditary nephritis. *Kidney International*. 42:783-92.
- Renieri, A., M. Seri, J.C. Myers, T. Pihlajaniemi, A. Sessa, G. Rizzoni, and M. De Marchi. 1992. Alport syndrome caused by a 5' deletion within the COL4A5 gene. *Human Genetics*. 89:120-1.
- Rhys, C., B. Snyers, and Y. Pirson. 1997. Recurrent corneal erosion associated with Alport's syndrome. Rapid communication. *Kidney International*. 52:208-11.
- Rich, A. 1955. The structure of collagen. *Nature*. 176:915-916.
- Risteli, J., G. Wick, and R. Timpl. 1981. Immunological characterization of the 7-S domain of type IV collagens. *Collagen & Related Research*. 1:419-32.
- Robinson, W.F., C.R. Huxtable, and J.P. Gooding. 1985. Familial nephropathy in cocker spaniels. *Australian Veterinary Journal*. 62:109-12.
- Robinson, W.F., S.E. Shaw, B. Stanley, C.R. Huxtable, A.D. Watson, S.E. Friend, and R. Mitten. 1989. Chronic renal disease in bull terriers. *Australian Veterinary Journal*. 66:193-5.
- Rodewald, R., and M.J. Karnovsky. 1974. Porous substructure of the glomerular slit diaphragm in the rat and mouse. *Journal of Cell Biology*. 60:423-33.
- Rogers, P.W., N.A. Kurtzman, S.M. Bunn, Jr., and M.G. White. 1973. Familial benign essential hematuria. *Archives of Internal Medicine*. 131:257-62.
- Rohrbach, D.H., C.W. Wagner, V.L. Star, G.R. Martin, K.S. Brown, and J.W. Yoon. 1983. Reduced synthesis of basement membrane heparan sulfate proteoglycan in streptozotocin-induced diabetic mice. *Journal of Biological Chemistry*. 258:11672-7.
- Rohrbach, D.H.a.T., R. 1991. Molecular and Celular Aspects of Basement Membranes. Academic Press, New York
- Rosenzweig, L.J., and Y.S. Kanwar. 1982. Removal of sulfated (heparan sulfate) or nonsulfated (hyaluronic acid) glycosaminoglycans results in increased permeability of the glomerular basement membrane to 125I-bovine serum albumin. *Laboratory Investigation*. 47:177-84.

- Rumpelt, H.J., K.H. Langer, K. Scharer, E. Straub, and W. Thoenes. 1974. Split and extremely thin glomerular basement membranes in hereditary nephropathy (Alport's syndrome). *Virchows Archiv. A, Pathological Anatomy & Histology*. 364:225-33.
- Ruotsalainen, V., P. Ljungberg, J. Wartiovaara, U. Lenkkeri, M. Kestila, H. Jalanko, C. Holmberg, and K. Tryggvason. 1999. Nephric is specifically located at the slit diaphragm of glomerular podocytes. *Proceedings of the National Academy of Sciences of the United States of America*. 96:7962-7.
- Rupp, F., D.G. Payan, C. Magill-Solc, D.M. Cowan, and R.H. Scheller. 1991. Structure and expression of a rat agrin. *Neuron*. 6:811-23.
- Saleem, M.A., L. Ni, I. Witherden, K. Tryggvason, V. Ruotsalainen, P. Mundel, and P.W. Mathieson. 2002. Co-localization of nephric, podocin, and the actin cytoskeleton: evidence for a role in podocyte foot process formation. *American Journal of Pathology*. 161:1459-66.
- Saus, J., S. Quinones, A. MacKrell, B. Blumberg, G. Muthukumaran, T. Pihlajaniemi, and M. Kurkinen. 1989. The complete primary structure of mouse alpha 2(IV) collagen. Alignment with mouse alpha 1(IV) collagen. *Journal of Biological Chemistry*. 264:6318-24.
- Saus, J., J. Wieslander, J.P. Langeveld, S. Quinones, and B.G. Hudson. 1988. Identification of the Goodpasture antigen as the alpha 3(IV) chain of collagen IV. *Journal of Biological Chemistry*. 263:13374-80.
- Saxena, R., P. Bygren, R. Butkowski, and J. Wieslander. 1990. Entactin: a possible auto-antigen in the pathogenesis of non-Goodpasture anti-GBM nephritis. *Kidney International*. 38:263-72.
- Shih, N.Y., J. Li, R. Cotran, P. Mundel, J.H. Miner, and A.S. Shaw. 2001. CD2AP localizes to the slit diaphragm and binds to nephric via a novel C-terminal domain. *American Journal of Pathology*. 159:2303-8.
- Shih, N.Y., J. Li, V. Karpitskii, A. Nguyen, M.L. Dustin, O. Kanagawa, J.H. Miner, and A.S. Shaw. 1999. Congenital nephrotic syndrome in mice lacking CD2-associated protein.[comment]. *Science*. 286:312-5.
- Siebold, B., R.A. Qian, R.W. Glanville, H. Hofmann, R. Deutzmann, and K. Kuhn. 1987. Construction of a model for the aggregation and cross-linking region (7S domain) of type IV collagen based upon an evaluation of the primary structure of

- the alpha 1 and alpha 2 chains in this region. *European Journal of Biochemistry*. 168:569-75.
- Soininen, R., T. Haka-Risku, D.J. Prockop, and K. Tryggvason. 1987. Complete primary structure of the alpha 1-chain of human basement membrane (type IV) collagen. *FEBS Letters*. 225:188-94.
- Soininen, R., M. Huotari, A. Ganguly, D.J. Prockop, and K. Tryggvason. 1989. Structural organization of the gene for the alpha 1 chain of human type IV collagen. *Journal of Biological Chemistry*. 264:13565-71.
- Soininen, R., M. Huotari, S.L. Hostikka, D.J. Prockop, and K. Tryggvason. 1988. The structural genes for alpha 1 and alpha 2 chains of human type IV collagen are divergently encoded on opposite DNA strands and have an overlapping promoter region. *Journal of Biological Chemistry*. 263:17217-20.
- Spear, G.S., and R.J. Slusser. 1972. Alport's syndrome. Emphasizing electron microscopic studies of the glomerulus. *American Journal of Pathology*. 69:213-24.
- Spiro, R.G. 1967a. The structure of the disaccharide unit of the renal glomerular basement membrane. *Journal of Biological Chemistry*. 242:4813-23.
- Spiro, R.G. 1967b. Studies on the renal glomerular basement membrane. Nature of the carbohydrate units and their attachment to the peptide portion. *Journal of Biological Chemistry*. 242:1923-32.
- Steward, A.P. 1984. Familial nephropathy in the cocker spaniel. *Journal of Small Animal Practice*. 25:15-24.
- Sugimoto, M., T. Oohashi, and Y. Ninomiya. 1994. The genes COL4A5 and COL4A6, coding for basement membrane collagen chains alpha 5(IV) and alpha 6(IV), are located head-to-head in close proximity on human chromosome Xq22 and COL4A6 is transcribed from two alternative promoters. *Proceedings of the National Academy of Sciences of the United States of America*. 91:11679-83.
- Swarm, R.H. 1964. Transplantation of a murine chondrosarcoma in mice of different inbred strains. *J. Natl. Cancer Inst.* 31:973-978.
- Szipiro-Tapia, S., G. Bobrie, M. Guilloud-Bataille, S. Heuertz, C. Julier, J. Frezal, J.P. Grunfeld, and M.C. Hors-Cayla. 1988. Linkage studies in X-linked Alport's syndrome. *Human Genetics*. 81:85-7.

- Teekhasaenee, C., S. Nimmanit, S. Wutthiphan, K. Vareesangthip, T. Laohapand, P. Malasitr, and R. Ritch. 1991. Posterior polymorphous dystrophy and Alport syndrome. *Ophthalmology*. 98:1207-15.
- Thorner, P., R. Bauman, V.E. Valli, D. Mahuran, R. McInnes, and P. Marrano. 1989. Abnormalities in the NC1 domain of collagen type IV in GBM in canine hereditary nephritis. *Kidney International*. 35:843-50.
- Thorner, P., B. Jansen, R. Bauman, V.E. Valli, and A. Goldberger. 1987. Samoyed hereditary glomerulopathy. Immunohistochemical staining of basement membranes of kidney for laminin, collagen type IV, fibronectin, and Goodpasture antigen, and correlation with electron microscopy of glomerular capillary basement membranes. *Laboratory Investigation*. 56:435-43.
- Thorner, P.S., K. Zheng, R. Kalluri, R. Jacobs, and B.G. Hudson. 1996. Coordinate gene expression of the alpha3, alpha4, and alpha5 chains of collagen type IV. Evidence from a canine model of X-linked nephritis with a COL4A5 gene mutation. *Journal of Biological Chemistry*. 271:13821-8.
- Timpl, R., M. Dziadek, S. Fujiwara, H. Nowack, and G. Wick. 1983. Nidogen: a new, self-aggregating basement membrane protein. *European Journal of Biochemistry*. 137:455-65.
- Timpl, R., H. Rohde, P.G. Robey, S.I. Rennard, J.M. Foidart, and G.R. Martin. 1979. Laminin--a glycoprotein from basement membranes. *Journal of Biological Chemistry*. 254:9933-7.
- Timpl, R., H. Wiedemann, V. van Delden, H. Furthmayr, and K. Kuhn. 1981. A network model for the organization of type IV collagen molecules in basement membranes. *European Journal of Biochemistry*. 120:203-11.
- Trueb, B., B. Grobli, M. Spiess, B.F. Odermatt, and K.H. Winterhalter. 1982. Basement membrane (type IV) collagen is a heteropolymer. *Journal of Biological Chemistry*. 257:5239-45.
- Tsen, G., W. Halfter, S. Kroger, and G.J. Cole. 1995. Agrin is a heparan sulfate proteoglycan. *Journal of Biological Chemistry*. 270:3392-9.
- Tsilibary, E.C., and A.S. Charonis. 1986. The role of the main noncollagenous domain (NC1) in type IV collagen self-assembly. *Journal of Cell Biology*. 103:2467-73.
- Uitto, J., L.W. Murray, B. Blumberg, and A. Shamban. 1986. UCLA conference. Biochemistry of collagen in diseases. *Annals of Internal Medicine*. 105:740-56.

- van den Born, J., L.P. van den Heuvel, M.A. Bakker, J.H. Veerkamp, K.J. Assmann, and J.H. Berden. 1992. A monoclonal antibody against GBM heparan sulfate induces an acute selective proteinuria in rats. *Kidney International*. 41:115-23.
- van der Loop, F.T., L. Heidet, E.D. Timmer, B.J. van den Bosch, A. Leinonen, C. Antignac, J.A. Jefferson, A.P. Maxwell, L.A. Monnens, C.H. Schroder, and H.J. Smeets. 2000. Autosomal dominant Alport syndrome caused by a COL4A3 splice site mutation. *Kidney International*. 58:1870-5.
- van der Rest, M., and R. Garrone. 1991. Collagen family of proteins. *FASEB Journal*. 5:2814-23.
- Vuorio, E., and B. de Crombrughe. 1990. The family of collagen genes. *Annual Review of Biochemistry*. 59:837-72.
- Weber, M. 1993. Rapidly progressive glomerulonephritis: recent advances in pathogenesis, diagnosis, and therapy. *Clinical Investigator*. 71:825-9.
- Wester, D.C., C.L. Atkin, and M.C. Gregory. 1995. Alport syndrome: clinical update. *Journal of the American Academy of Audiology*. 6:73-9.
- Wieslander, J., J.F. Barr, R.J. Butkowski, S.J. Edwards, P. Bygren, D. Heinegard, and B.G. Hudson. 1984. Goodpasture antigen of the glomerular basement membrane: localization to noncollagenous regions of type IV collagen. *Proceedings of the National Academy of Sciences of the United States of America*. 81:3838-42.
- Wong, M.A., S. Cui, and S.E. Quaggin. 2000. Identification and characterization of a glomerular-specific promoter from the human nephrin gene. *American Journal of Physiology - Renal Fluid & Electrolyte Physiology*. 279:F1027-32.
- Wu, R.R., and J.R. Couchman. 1997. cDNA cloning of the basement membrane chondroitin sulfate proteoglycan core protein, bamacan: a five domain structure including coiled-coil motifs. *Journal of Cell Biology*. 136:433-44.
- Wu, T.C., Y.J. Wan, A.E. Chung, and I. Damjanov. 1983. Immunohistochemical localization of entactin and laminin in mouse embryos and fetuses. *Developmental Biology*. 100:496-505.
- Yoshioka, K., S. Hino, T. Takemura, S. Maki, J. Wieslander, Y. Takekoshi, H. Makino, M. Kagawa, Y. Sado, and C.E. Kashtan. 1994. Type IV collagen alpha 5 chain. Normal distribution and abnormalities in X-linked Alport syndrome revealed by monoclonal antibody. *American Journal of Pathology*. 144:986-96.

- Yuan, H., E. Takeuchi, G.A. Taylor, M. McLaughlin, D. Brown, and D.J. Salant. 2002. Nephrin dissociates from actin, and its expression is reduced in early experimental membranous nephropathy. *Journal of the American Society of Nephrology*. 13:946-56.
- Yurchenco, P.D., Y.S. Cheng, and H. Colognato. 1992. Laminin forms an independent network in basement membranes.[erratum appears in J Cell Biol 1992 Jun;118(2):493]. *Journal of Cell Biology*. 117:1119-33.
- Yurchenco, P.D., and H. Furthmayr. 1984. Self-assembly of basement membrane collagen. *Biochemistry*. 23:1839-50.
- Yurchenco, P.D., and G.C. Ruben. 1987. Basement membrane structure in situ: evidence for lateral associations in the type IV collagen network. *Journal of Cell Biology*. 105:2559-68.
- Yurchenco, P.D., E.C. Tsilibary, A.S. Charonis, and H. Furthmayr. 1985. Laminin polymerization in vitro. Evidence for a two-step assembly with domain specificity. *Journal of Biological Chemistry*. 260:7636-44.
- Zheng, K., P.S. Thorner, P. Marrano, R. Baupal, and R.R. McInnes. 1994. Canine X chromosome-linked hereditary nephritis: a genetic model for human X-linked hereditary nephritis resulting from a single base mutation in the gene encoding the alpha 5 chain of collagen type IV. *Proceedings of the National Academy of Sciences of the United States of America*. 91:3989-93.
- Zhou, J., M. Ding, Z. Zhao, and S.T. Reeders. 1994a. Complete primary structure of the sixth chain of human basement membrane collagen, alpha 6(IV). Isolation of the cDNAs for alpha 6(IV) and comparison with five other type IV collagen chains. *Journal of Biological Chemistry*. 269:13193-9.
- Zhou, J., J.M. Hertz, A. Leinonen, and K. Tryggvason. 1992. Complete amino acid sequence of the human alpha 5 (IV) collagen chain and identification of a single-base mutation in exon 23 converting glycine 521 in the collagenous domain to cysteine in an Alport syndrome patient. *Journal of Biological Chemistry*. 267:12475-81.
- Zhou, J., A. Leinonen, and K. Tryggvason. 1994b. Structure of the human type IV collagen COL4A5 gene. *Journal of Biological Chemistry*. 269:6608-14.

APPENDIX 1

Canine COL4A3 nucleotide sequence

Sequence Range: 1 to 4643

10	20	30	40	50
AACTT TAAGA AGGAG ATATA CCATG GCGAT GGTGT ACCAG GATGC AATGG				
TTGAA ATTCT TCCTC TATAT GGTAC CGCTA CCACA TGGTC CTACG TTACC				
60	70	80	90	100
TTCTA AGGGT GAGCA AGGGT TTCCA GGCCT TCCAG GGACA CCAGG CTACC				
AAGAT TCCCA CTCGT TCCCA AAGGT CCGGA AGGTC CCTGT GTTCC GATGG				
110	120	130	140	150
CAGGG ATCCC GGGTG CTGTT GGCCT GAAAG GAGAA AAAGG TGCTC CTGCC				
GTCCC TAGGG CCCAC GACAA CCGGA CTTTC CTCTT TTTCC ACGAG GACGG				
160	170	180	190	200
ACAGA AGGTA TAGAA CTTGA TGGAA GAGGT GACCC TGGGT TGCCA GGAGC				
TGTCT TCCAT ATCTT GAACT ACCTT CTCCA CTGGG ACCCA ACGGT CCTCG				
210	220	230	240	250
TCCAG GATTC CAGGG TTTAC CAGGC CTTCC AGGCT TTCCG GGACC TGCTG				
AGGTC CTAAG GTCCC AAATG GTCCG GAAGG TCCGA AAGGC CCTGG ACGAC				
260	270	280	290	300
GTCCA CCTGG CCCTC CGGGA TTCTT AGGCT TTCCA GGAGC CATGG GACCT				
CAGGT GGACC GGGAG GCCCT AAGAA TCCGA AAGGT CCTCG GTACC CTGGA				
310	320	330	340	350
CCAGG ACCTA AGGGT CAAAT GGGCG ATAAA GTGAT AGGAC AAAAA GGAGA				
GGTCC TGGAT TCCCA GTTTA CCCGC TATTT CACTA TCCTG TTTTT CCTCT				
360	370	380	390	400
GCGGG GTGTG AAAGG ATTAA CAGGA CCTCC CGGAC CACCA GGAAC AGTTA				
CGCCC CACAC TTTCC TAATT GTCCT GGAGG GCCTG GTGGT CCTTG TCAAT				
410	420	430	440	450
TTGTG ACGCT AACCG GCCCT GATAA CAGAA CGGAC CTCAA GGGGG AGAAG				
AACAC TGC GA TTGGC CGGGA CTATT GTCTT GCCTG GAGTT CCCCC TCTTC				
460	470	480	490	500
GGAGA CAAGG GAGCC ATGGG GAAGC CTGGA CCTCC TGGGC CCTTG GGACC				
CCTCT GTTCC CTCGG TACCC CTTCC GACCT GGAGG ACCCG GGAAC CCTGG				
510	520	530	540	550
GCCTG GAGAA TCCTA TGGAT CTGAA AAAGG TGCTC CTGGA GAACC TGGCC				
CGGAC CTCTT AGGAT ACCTA GACTT TTTCC ACGAG GACCT CTTGG ACCGG				
560	570	580	590	600
CGCAG GGAAA ACCTG GCAAA GATGG TGCCC CTGGT TTCCC TGGCA CTGAG				
GCGTC CCTTT TGGAC CGTTT CTACC ACGGG GACCA AAGGG ACCGT GACTC				

610	620	630	640	650
GGGCC CAAAG GCAAC AGAGG CTTCC CTGGG TTACG GGGTG AAGAC GGCAT				
CCCGG GTTTC CGTTG TCTCC GAAGG GACCC AATGC CCCAC TTCTG CCGTA				
660	670	680	690	700
TAAGG GGTGG AAAGG GGACA TTGGC CCGCC AGGAT TTCGT GGTCC AACAG				
ATTCC CCACC TTTCC CCTGT AACCG GCGCG TCCTA AAGCA CCAGG TTGTC				
710	720	730	740	750
AATAT TATGA TGCAT ACCAG GAAAA GGGTG ATGAA GGAAT TCCAG GCCCA				
TTATA ATACT ACGTA TGGTC CTTTT CCCAC TACTT CCTTA AGGTC CGGGT				
760	770	780	790	800
CCAGG CCCCA AAGGA GCTCG TGGCC CCCAG GGTCC CAGCG GTCCC CCTGG				
GGTCC GGGGT TTCCT CGAGC ACCGG GGGTC CCAGG GTCGC CAGGG GGACC				
810	820	830	840	850
AGTTC TTGGA AGTGC TGGGT CATCA AAGCC TGGCC TCAGA GGAGC TCCTG				
TCAAG AACCT TCACG ACCCA GTAGT TTCGG ACCGG AGTCT CCTCG AGGAC				
860	870	880	890	900
GATCT CCAGG CATGA AAGGA CGTAA AGGGG AACAA GGGCC CCCGG GAAAG				
CTAGA GGTCC GTACT TTCCT GCATT TCCCC TTGTT CCCGG GGGCC CTTTC				
910	920	930	940	950
AATGC AGTGG GGCCT CCTGG GTCCC CAGGT TGTCC TGGTT CACCA GGCCC				
TTACG TCACC CCGGA GGACC CAGGG GTCCA ACAGG ACCAA GTGGT CCGGG				
960	970	980	990	1000
CATAG GGCTG CCGGG ATATC CAGGA CCACC AGGTG ACATC GTGTT TCGCA				
GTATC CCGAC GGCCC TATAG GTCCT GGTGG TCCAC TGTAG CACAA AGCGT				
1010	1020	1030	1040	1050
AAGGT CCACC TGGGG TTGGT GGACT CCCCC GCCAT GTTGG GTTTC CAGGC				
TTCCA GGTGG ACCCC AACCA CCTGA GGGGC CGGTA CAACC CAAAG GTCCG				
1060	1070	1080	1090	1100
ATCCC AGGAG TCGAG GGGCC CAAAG GGGAA CCAGG CCTCT TGTGC GTACA				
TAGGG TCCTC AGCTC CCCGG GTTTC CCCTT GGTCC GGAGA ACACG CATGT				
1110	1120	1130	1140	1150
GTGTG CTTAT ATCCC AGGGC GTCCA GGTCT CCCAG GACTG CCAGG GTTGG				
CACAC GAATA TAGGG TCCCG CAGGT CCAGA GGGTC CTGAC GTTCC CAACC				
1160	1170	1180	1190	1200
ATGGC ATAAA AGGAA TCCCA GGAGG ACAAG GGGCA GCTGG CATTA AAGGA				
TACCG TATTT TCCTT AGGGT CCTCC TGTTC CCCGT CGACC GTAAT TTCCT				
1210	1220	1230	1240	1250
AGCCC AGGGT CCCCA GGAAG TGCGG GTCTT CCAGG ATTTT CAGGA TTCCC				
TCGGG TCCCA GGGGT CCTTC ACGCC CAGAA GGTCC TAAAG GTCCT AAGGG				
1260	1270	1280	1290	1300
GGGTG CTCCG GGTGC TCCAG GACTT AAAGG AGAAA AAGGA GAAAC ATCTG				
CCCAC GAGGC CCACG AGGTC CTGAA TTTCC TCTTT TTCCT CTTTG TAGAC				

1310	1320	1330	1340	1350
AGCCT GAGGG ACAGG	TGGGT GCTCC AGGGG	ACCCC GGGCT	CAGAG GCCAT	
TCGGA CTCCC TGTCC	ACCCA CGAGG TCCCC	TGGGG CCCGA	GTCTC CGGTA	
1360	1370	1380	1390	1400
CCTGG AAGAA AGGGC	TTGGA TGGAA TTCCT	GGAAC TCCTG	GAATT AAAGG	
GGACC TTCTT TCCCG	AACCT ACCTT AAGGA	CCTTG AGGAC	CTTAA TTTCC	
1410	1420	1430	1440	1450
ACCAC CAGGA CCCAA	AGGTG AACCG GCCCT	GAGTG GTGAG	AAGGG GGACC	
TGGTG GTCCT GGGTT	TCCAC TTGGC CGGGA	CTCAC CACTC	TTCCC CCTGG	
1460	1470	1480	1490	1500
AGGGT CCTCC AGGGG	ATCCT GGGAT CCCAG	GGTCC CCAGG	ACCTG CAGGA	
TCCCA GGAGG TCCCC	TAGGA CCCTA GGGTC	CCAGG GGTCC	TGGAC GTCCT	
1510	1520	1530	1540	1550
CCACC TGGAC CACCA	AACTA TGGAC CACAG	GGAGA GCCTG	GTCCA AAGGG	
GGTGG ACCTG GTGGT	TTGAT ACCTG GTGTC	CCTCT CGGAC	CAGGT TTCCC	
1560	1570	1580	1590	1600
CACCC AAGGA GTTCC	TGGAG CCCCT GGACC	ACCTG GAGAA	GCCGG TCCTA	
GTGGG TTCTT CAAGG	ACCTC GGGGA CCTGG	TGGAC CTCTT	CGGCC AGGAT	
1610	1620	1630	1640	1650
AGGGA GAATT CGGTA	TTTCG ACACC AGTCC	CAGGG CCCCC	AGGAC CTCCA	
TCCCT CTTAA GCCAT	AAAGC TGTGG TCAGG	GTCCC GGGGG	TCCTG GAGGT	
1660	1670	1680	1690	1700
GGGCC CCCTG GTTAT	GCTGG CCCCC GAGGT	CCACC TGGTA	TCCCT GGATC	
CCCGG GGGAC CAATA	CGACC GGGGG CTCCA	GGTGG ACCAT	AGGGA CCTAG	
1710	1720	1730	1740	1750
CATAG GAAAA TGTGG	TGATC CGGGT CTTCC	TGGGC CTGAT	GGTGA ACCAG	
GTATC CTTTT ACACC	ACTAG GCCCA GAAGG	ACCCG GACTA	CCACT TGGTC	
1760	1770	1780	1790	1800
GAATT CCAGG AATTG	GCTTC CCTGG GCCCC	CTGGA CCTAA	GGGAG ACCAA	
CTTAA GTTCC TTAAC	CGAAG GGACC CGGGG	GACCT GGATT	CCCTC TGTTT	
1810	1820	1830	1840	1850
GGTTT TCCAG GAGCA	AAAGG AGCAC CGGGT	TGTCC AGGAG	AAATG GGGAA	
CCAAA AGGTC CTCGT	TTTCC TCGTG GCCCA	ACAGG TCCTC	TTTAC CCCTT	
1860	1870	1880	1890	1900
GCCCG GGTCA CCTGG	AGAAC CAGGT CTCCC	AGGAG CCAAG	GGAGA ACCAG	
CGGGC CCAGT GGACC	TCTTG GTCCA GAGGG	TCCTC GGTTT	CCTCT TGGTC	
1910	1920	1930	1940	1950
GACTA GCCAT ACCTG	GAGAA CCAGG AATAC	CAGGT TTTCC	AGGAG AAAGA	
CTGAT CGGTA TGGAC	CTCTT GGTCC TTATG	GTCCA AAAGG	TCCTC TTTCT	
1960	1970	1980	1990	2000
GGCAA TTCTG GGGAA	AATGG AGAAA TTGGA	CTCCC TGGAC	TTCCA GGTCT	
CCGTT AAGAC CCCTT	TTACC TCTTT AACCT	GAGGG ACCTG	AAGGT CCAGA	

2010	2020	2030	2040	2050
CCCTG GAATT CCAGG AACTG GAGGG CTTGA TGGAC CACCA GGGGA TCCAG				
GGGAC CTTAA GGTCC TTGAC CTCCC GAACT ACCTG GTGGT CCCCT AGGTC				
2060	2070	2080	2090	2100
GGAAG CCTGG ACCAC CTGGA GAAAA AGGAC CCCCA GGAAG TTGCA CAGAG				
CCTTC GGACC TGGTG GACCT CTTTT TCCTG GGGGT CCTTC AACGT GTCTC				
2110	2120	2130	2140	2150
GGTCC CAGAG GAGCC CACGG ACTTC CAGGA TTAAA TGGAT TGGAA GGGCA				
CCAGG GTCTC CTCGG GTGCC TGAAG GTCCT AATTT ACCTA ACCTT CCCGT				
2160	2170	2180	2190	2200
ACCAG GCAGA AGAGG TGAAA CAGGG CCAAA AGGAG ACCCA GGTAT TCCAG				
TGGTC CGTCT TCTCC ACTTT GTCCC GGTTT TCCTC TGGGT CCATA AGGTC				
2210	2220	2230	2240	2250
GCTTG GATAG ATCAG GCTTT CCTGG AGAAC CTGGA CCACC AGGAA TGCCA				
CGAAC CTATC TAGTC CGAAA GGACC TCTTG GACCT GGTGG TCCTT ACGGT				
2260	2270	2280	2290	2300
GGTCA TCGAG GTGAG ATAGG ACCAC CTGGC CAAAA AGGAT ATCCA GGAAA				
CCAGT AGCTC CACTC TATCC TGGTG GACCG GTTTT TCCTA TAGGT CCTTT				
2310	2320	2330	2340	2350
TCCGG GATTT TTAGG ACTAC CAGGT GAAGA TGGAG TGATT GGGAT GATGG				
AGGCC CTAAA AATCC TGATG GTCCA CTTCT ACCTC ACTAA CCCTA CTACC				
2360	2370	2380	2390	2400
GCTAT CCAGG AAACA CTGGC CCTCC TGGGC CTCCC GGGAT CCCAG GCACC				
CGATA GGTCC TTTGT GACCG GGAGG ACCCG GAGGG CCCTA GGGTC CGTGG				
2410	2420	2430	2440	2450
CCAGG ACAGA GGGGT AGCTT TGGAA TTCCA GGTGC AAAGG GTGAG AAAGG				
GGTCC TGTCT CCCCC TCGAA ACCTT AAGGT CCACG TTTCC CACTC TTTCC				
2460	2470	2480	2490	2500
GCCCC CAGGA GCCAA GGGGG AAGAA GGAGA GAAAG GACCT ATGGG GCCTT				
CGGGG GTCCT CGGTT CCCCC TTCTT CCTCT CTTTC CTGGA TACCC CGGAA				
2510	2520	2530	2540	2550
GTCAA ATAAC CAGCT CAGTG GGGGA CAAAG GAGAA CCAGG CCTCA AAGGG				
CAGTT TATTG GTCGA GTCAC CCCCT GTTTC CTCTT GGTCC GGAGT TTCCC				
2560	2570	2580	2590	2600
TTTGC AGGAA AGCCG GGTGA AAAAG GAAAC AGAGG CATTG CAGGG TTACC				
AAACG TCCTT TCGGC CCACT TTTTC CTTTG TCTCC GTAAG GTCCC AATGG				
2610	2620	2630	2640	2650
AGGTT TGAAA GGATT CGAAG GGCCA CCTGG ACCAC CAGGC CCACC AGGCC				
TCCAA ACTTT CCTAA GCTTC CCGGT GGACC TGGTG GTCCG GGTGG TCCGG				
2660	2670	2680	2690	2700
CCCGA GGAGA TCCAG GCAGC ATTGG GAATC CTGGA GAAGC AGGAC CACGT				
GGGCT CCTCT AGGTC CGTCG TAACC CTTAG GACCT CTTCG TCCTG GTGCA				

2710	2720	2730	2740	2750
GGAAG TCCAG GAAAC	ATGGG GAACA TGGGG	ATGCC AGGTC TTAAA	GGACA	
CCTTC AGGTC CTTTG	TACCC CTTGT ACCCC	TACGG TCCAG AATTT	CCTGT	
2760	2770	2780	2790	2800
CAGGG GACCT TTGGG	ACTAC CAGGT GTCAC	TGGAA GACCA GGCCT	CCCAG	
GTCCC CTGGA AACCC	TGATG GTCCA CAGTG	ACCTT CTGGT CCGGA	GGGTC	
2810	2820	2830	2840	2850
GTGTT CACGG TCTCC	AAGGA GACAA GGGAG	AGCCA GGTTA TTCAG	CAGGT	
CACAA GTGCC AGAGG	TTCCCT CTGTT CCCTC	TCGGT CCAAT AAGTC	GTCCA	
2860	2870	2880	2890	2900
ACAAG GCCAG GACCA	CCGGG ACTAA AGGGA	GATCC AGGAT TACCA	GGTGC	
TGTTT CCGTC CTGGT	GGCCC TGATT TCCCT	CTAGG TCCTA ATGGT	CCACG	
2910	2920	2930	2940	2950
CATGG GAAGG AAAGG	AGAAA GAGGG CTACC	TGGCA CCCCT GGCCA	TTCGG	
GTACC CTTCC TTTCC	TCTTT CTCCC GATGG	ACCGT GGGGA CCGGT	AAGCC	
2960	2970	2980	2990	3000
GGCCT GCTGG AACTG	AGGGA GCCCC TGGAC	ATCCC GGAAG TCCTG	GCCAC	
CCGGA CGACC TTGAC	TCCCT CGGGG ACCTG	TAGGG CCTTC AGGAC	CGGTG	
3010	3020	3030	3040	3050
CCAGG AAAGC CAGGC	CCTGA TGGAG ATTTG	GGGTT AAAAG GCATC	AAAGG	
GGTCC TTTCG GTCCG	GGACT ACCTC TAAAC	CCCAA TTTTC CGTAG	TTTCC	
3060	3070	3080	3090	3100
CTTCC CTGGG TCCCC	AGGAG TCAAA GGACC	TCCAG GACCT CCAGG	AATCC	
GAAGG GACCC AGGGG	TCCTC AGTTT CCTGG	AGGTC CTGGA GGTCC	TTAGG	
3110	3120	3130	3140	3150
TAGGA TCTCC TGGAC	CTATG GGCAT GAGAG	GCAAC CAGGG CCGTG	ATGGA	
ATCCT AGAGG ACCTG	GATAC CCGTA CTCTC	CGTTG GTCCC GGCAC	TACCT	
3160	3170	3180	3190	3200
ATCCC TGGTC CCCCC	GGAGA AAAGG GAGAA	ACAGG TTTGC TGGGG	GCACA	
TAGGG ACCAG GGGGT	CCTCT TTTCC CTCTT	TGTCC AAACG ACCCC	CGTGT	
3210	3220	3230	3240	3250
TCCAG GCCCA GGAGG	GAGCC CTGGT CCTCC	AGGAG CCAAA GGAGA	CAGGG	
AGGTC CGGGT CCTCC	CTCGG GACCA GGAGG	TCCTC GGTTT CCTCT	GTCCC	
3260	3270	3280	3290	3300
GCGCC CCAGG CCTAC	CAGGC CCCCC AGGCA	GGAAA GGGGC AGTGG	GAGAT	
CGCGG GTTCC GGATG	GTCCG GGGGG TCCGT	CCTTT CCCC	TCACC	CTCTA
3310	3320	3330	3340	3350
GCCGG GCCAC GGGGA	CCCAC TGGCA TGACG	GGACT CCCAG GGCCA	CCAGG	
CGGCC CGGTG CCCCT	GGGTG ACCGT ACTGC	CCTGA GGGTC CCGGT	GGTCC	
3360	3370	3380	3390	3400
TTTTT CTGGT GCAGT	CATCC CTGGC CAGAA	AGGAA ATCAA GTTCC	ACCAG	
AAAAG GACCA CGTCA	GTAGG GACCG GTCTT	TCCTT TAGTT CCAGG	TGGTC	

3410	3420	3430	3440	3450
GCTTC AGAGG AAACC CAGGT GAGCC TGGTC CTCCA GGCCC TCCAG GGAGC				
CGAAG TCTCC TTTGG GTCCA CTCGG ACCAG GAGGT CCGGG AGGTC CCTCG				
3460	3470	3480	3490	3500
CACGT GAAAG GCATA AAAGG AGACA AGGGA CTCAT GGGTG AGCCT GGCCC				
GTGCA CTTTC CGTAT TTTC TCTGT TCCCT GAGTA CCCAC TCGGA CCGGG				
3510	3520	3530	3540	3550
CAGAG GTCTA CCTGG AACTG TGGGA GACAA GGGGC CACCG GTTCA ACCGG				
GTCTC CAGAT GGACC TTGAC ACCCT CTGTT CCCCC GTGGC CCAGT TGGCC				
3560	3570	3580	3590	3600
GAGCA CCAGG GAGCC CAGGT CTGCC AGGGC TCAGG GGCGA TCCTG GATTC				
CTCGT GGTCC CTCGG GTCCA GACGG TCCCG AGTCC CCGCT AGGAC CTAAG				
3610	3620	3630	3640	3650
TATGG ATTTT CAGGT GTTAA AGGAG AGAAG GGTAA CCCAG GATTT CCGGG				
ATACC TAAAG GTCCA CAATT TCCTC TCTTC CCATT GGGTC CTAAA GGCCC				
3660	3670	3680	3690	3700
ACCAG CTGGA CCTCC AGGGC AAATT GGGCC AAAAG GACCA CCTGG TGTCC				
TGGTC GACCT GGAGG TCCCG TTTAA CCGGG TTTTC CTGGT GGACC ACAGG				
3710	3720	3730	3740	3750
GTGGA GACCC TGGCA CAGTT AAGAT CATCT CCCTT CCAGG AAGCC CAGGG				
CACCT CTGGG ACCGT GTCAA TTCTA GTAGA GGGAA GGTCC TTCGG GTCCC				
3760	3770	3780	3790	3800
CCACC TGGCC GTGCT GGAGG ACCGG GGATG CCAGG AGAAC CAGGG CCACC				
GGTGG ACCGG CACGA CCTCC TGGCC CCTAC GGTCC TCTTG GTCCC GGTGG				
3810	3820	3830	3840	3850
AGGGC CACCA GGAAT CCTAG GACCC TGTGG ACCAA GAGGT AAACC AGGCA				
TCCCG GTGGT CCTTA GGATC CTGGG ACACC TGGTT CTCCA TTTGG TCCGT				
3860	3870	3880	3890	3900
TGGAT GGAAT ACCAG GAACT CCTGG GCCAA TTGGA GAAAA AGGCA ACAA				
ACCTA CCTTA TGGTC CTTGA GGACC CGGTT AACCT CTTTT TCCGT TGTTT				
3910	3920	3930	3940	3950
GGTTG TAAAG GCGAG CAAGG ACCAC CTGGA TTAGA TGGAC TGCCA GGCTT				
CCAAC ATTTT CGCTC GTTCC TGGTG GACCT AATCT ACCTG ACGGT CCGAA				
3960	3970	3980	3990	4000
GAAGG GGAGA CCTGG AGACC CTGGA CCACC TGCAA CTGGG ACCAT GAAGA				
CTTCC CCTCT GGACC TCTGG GACCT GGTGG ACGTT GACCC TGGTA CTTCT				
4010	4020	4030	4040	4050
GGGGC TTTAT CTTCA CCAGG CATAG TCAGA CCACA GTGAT TCCTT CCTGT				
CCCCG AAATA GAAGT GGTCC GTATC AGTCT GGTGT CACTA AGGAA GGACA				
4060	4070	4080	4090	4100
CCAGA AGGGA CAGCG CCACT CTATA GTGGC TTTTC TCTTC TTTTC ATACA				
GGTCT TCCCT GTCGC GGTGA GATAT CACCG AAAAG AGAAG AAAAG TATGT				

4110	4120	4130	4140	4150
AGGAA ATGAA CAAGC CCATG GACAA GACTT GGGAA CTCTC GGCAG CTGCC				
TCCTT TACTT GTTCG GGTAC CTGTT CTGAA CCCTT GAGAG CCGTC GACGG				
4160	4170	4180	4190	4200
TGCAG CGATT TACCA CAATG CCATT CTTGT TCTGT AACAT CAACA ATGTC				
ACGTC GCTAA ATGGT GTTAC GGTA GAACA AGACA TTGTA GTTGT TACAG				
4210	4220	4230	4240	4250
TGTAA TTTTG CATCT CGAAA TGA CT ACTCA TACTG GCTGT CAACA CCAGC				
ACATT AAAAC GTAGA GCTTT ACTGA TGAGT ATGAC CGACA GTTGT GGTGC				
4260	4270	4280	4290	4300
TCTGA TGCCA ATGGA CATGG CTCCA ATTAC TGGCA GGGCC CTGGA GCCTT				
AGACT ACGGT TACCT GTACC GAGGT TAATG ACCGT CCCGG GACCT CGGAA				
4310	4320	4330	4340	4350
ATATT AGCAG ATGCA CTGTC TGTGA AGGTC CTACG ATTGC CATAG CCATT				
TATAA TCGTC TACGT GACAG AACT TCCAG GATGC TAACG GTATC GGTAA				
4360	4370	4380	4390	4400
CACAG CCAAA CCACT GACAT TCCCT CATGT CCCAA TGGCT GGATT TCTCT				
GTGTC GGTTT GGTGA CTGTA AGGGA GTACA GGGTT ACCGA CCTAA AGAGA				
4410	4420	4430	4440	4450
CTGGA AAGGA TTTTC GTTTA TCATG TTCAC AAGTG CAGGT TCTGA GGGTG				
GACCT TTCCT AAAAG CAAAT AGTAC AAGTG TTCAC GTCCA AGACT CCCAC				
4460	4470	4480	4490	4500
CTGGG CAAGC ACTGG CCTCC CCCGG CTCCT GCCTG GAAGA GTTCC GAGCC				
GACCC GTTCG TGACC GGAGG GGGCC GAGGA CGGAC CTTCT CAAGG CTCGG				
4510	4520	4530	4540	4550
AGTCC GTTTA TAGAA TGTCA CGGAA GAGGA ACGTG CAACT ACTAT TCAAA				
TCAGG CAAAT ATCTT ACAGT GCCTT CTCCT TGCAC GTTGA TGATA AGTTT				
4560	4570	4580	4590	4600
TTCCT ACAGT TTCTG GTTGG CTTCTG TTAAA CCCCC AAAGA ATGTT CAGAA				
AAGGA TGTCA AAGAC CAACC GAAGC AATTT GGGGG TTTCT TACAA GTCTT				
4610	4620	4630	4640	
AACCT ATTCC ATCAA CTGTG AAAGC TGAAT CCATA TGA CT AGT				
TTGGA TAAGG TAGTT GACAC TTTCG ACTTA GGTAT ACTGA TCA				

APPENDIX 2

Canine COL4A3 nucleotide alignments

Consensus key (see documentation for details)

- * - single, fully conserved residue
- no consensus

CLUSTAL W (1.81) multiple sequence alignment

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Canine_Normal_5-18      -----
Human_COL4A3           ATGAGCGCCCGGACCGCCCCAGGCCGAGGTGCTCCTGCTGCCGCTCCTGCTGGTGCTC
Mouse_COL4A3           ATGCACTCCAAGACTGCTCCAAGGTTCTTGGTGTCTCCTGCTGCTTACCCTGCTGCTACTC

Canine_Normal_5-18      -----
Human_COL4A3           CTGGCGGCGGCGCCCGCAGCCAGCAAGGGTTGTGTCTGTAAAGACAAAGGCCAGTGCTTC
Mouse_COL4A3           CTGGCTGCCTCTCCTGTGGCTAGCAAGGGCTGTGTCTGCAAAGGCAAAGGACAATGCCTC

Canine_Normal_5-18      -----
Human_COL4A3           TGTGACGGGGCCAAAGGGGAGAAGGGGGAGAAGGGCTTTCCTGGACCCCCCGGTTCTCCT
Mouse_COL4A3           TGCGCGGGGACCAAAGGGGAGAAGGGGGAGAAGGGGTTCTTGTTCCCTGGATTTCCT

Canine_Normal_5-18      -----
Human_COL4A3           GGCCAGAAAGGATTTCACAGGTCTGAAGGCTTGCTTGACCGCAGGGACCCAAGGGCTTT
Mouse_COL4A3           GGCCAGAAAGGATTTCAGGTCTGAAGGCTTGCTTGACCCAGGGACCCAAGGGCTCC

Canine_Normal_5-18      -----
Human_COL4A3           CCAGGACTTCCAGGACTCACGGGTTCCAAAGGTGTAAGGGGAATAAGTGGATTGCCAGGA
Mouse_COL4A3           CCAGGACTTCCGGGACTCACTGGCCCCAAAGGCATCAGGGGAATAACTGGATTACCAGGG

Canine_Normal_5-18      -----AAGTTTAAAGAAG----GAGATATACCATGGCGAT
Human_COL4A3           TTTTCTGGTTCTCCTGGACTTCCAGGCACCCAGGCAATACCGGGCCTTACGGACTTGTC
Mouse_COL4A3           TTTGCAGGTCCTCCTGGACTTCCAGGCCTCCAGGCCACCTGGGCCTCGTGGGCTGGCT
                                *  *                *  *                *

Canine_Normal_5-18      GGTGTACCAGGATGCAATGGTTCTAAGGGTGAGCAAGGGTTTCCAGGCCTTCCAGGGACA
Human_COL4A3           GGTGTACCAGGATGCAGTGGTTCTAAGGGTGAGCAGGGGTTTCCAGGACTCCCAGGGACA
Mouse_COL4A3           GGTTTACCAGGATGCAACGGATCTAAGGGTGAACAAGGATTCCCGGGCTTCCCGGCACA
                                *** ***** ** ***** ** * * * * * * * * * *

Canine_Normal_5-18      CCAGGCTACCCAGGGATCCCGGGTGCTGTTGGCCTGAAAGGAGAAAAAGGTGCTCCTGCC
Human_COL4A3           CCGGGCTACCCAGGGATCCCGGGTGCTGCTGGTTTGAAAGGACAAAAGGGTGCTCCTGCT
Mouse_COL4A3           CCAGGCTATGCAGGGCTCCAGGTCCTGATGGCTTGAAAGGACAAAAGGGTGAGCCTGCT
                                ** ***** ***** ** * * * * * * * * * *

Canine_Normal_5-18      ACAG---AAGGTATAGAACTTGATGGAAGAGGTGACCCTGGGTGCCAGGAGCTCCAGGA
Human_COL4A3           AAAGGAGAAGATATAGAACTTGATGCAAAAGGCGACCCGGGTGCCAGGGGCTCCAGGA
Mouse_COL4A3           CAAGGAGAAGACAGGGGATTCAATGGAAAAGGTGACCCTGGGCCTCCAGGGGTTCCAGGC
                                ** *** * * * * * * * * * * * * * * * *

Canine_Normal_5-18      TTCCAGGGTTTACCAGGCCTTCCAGGCTTTCGGGGACCTGCTGGTCCACCTGGCCCTCCG
Human_COL4A3           CCCAGGGTTTGGCAGGCCCTCCAGGTTTTCCTGGGCCTGTTGGCCACCTGGTCTCTCCG
Mouse_COL4A3           TTCCAGGGTTTTCGGGACTCCAGGTTTTCAGGGCCTGCCGGTCCACAGGACCTCCG
                                ***** ** * * * ***** ** * * * * * * * * *

Canine_Normal_5-18      GGATTCTTAGGCTTTCAGGAGCCATGGGACCTCCAGGACCTAAGGGTCAAATGGGCGAT
Human_COL4A3           GGATTCTTTGGCTTTCAGGAGCCATGGGACCTAGAGGACCTAAGGGTCACATGGGTGAA
Mouse_COL4A3           GGATTTTGTGTTTACCAGGAGCAATGGGACCCAGAGGACCAAGGCCACATGGGCGAT
                                ***** ** * * * ***** ***** ** * * * * * * *

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Canine_Normal_5-18	AAAGTGATAGGACAAAAAGGAGAGCGGGGTGTGAAAGGATTAACAGGACCTCCCGGACCA
Human_COL4A3	AGAGTGATAGGACATAAAGGAGAGCGGGGTGTGAAAGGTTAACAGGACCCCCGGGACCA
Mouse_COL4A3	AGCGTGATAGGACAAAAAGGAGAAAGGGGTATGAAAGGATTAACAGGACCTCCTGGGCCA
	* * * * *
Canine_Normal_5-18	CCAGGAACAGTTATTGTGACGCTAACCGGCCCTGATAACAGAACGGACCTCAAGGGGGAG
Human_COL4A3	CCAGGAACAGTTATTGTGACCTAACTGGCCAGATAACAGAACGGACCTCAAGGGGGAA
Mouse_COL4A3	CCAGGAACAGTGTATTTTACACTCACCAGCCATACAACAAATCGACTTCAAGGGAGAG
	* * * * *
Canine_Normal_5-18	AAGGGAGACAAGGGAGCCATGGGGAAGCCTGGACCTCCTGGGCCCTTGGGACCGCCTGGA
Human_COL4A3	AAGGGAGACAAGGGAGCAATGGGCGAGCCTGGACCTCCTGGACCTCAGGACTGCCTGGA
Mouse_COL4A3	AAAGGAGATGAGGGAGAGAGGGGTGAACCCGGACCTCCTGGACCTCGGGGCCACCTGGA
	* * * * *
Canine_Normal_5-18	GAATCCTATGGATCTGAAAAAGGTGCTCCTGGAGAACCTGGCCCGCAGGGAAAACCTGGC
Human_COL4A3	GAATCATATGGATCTGAAAAGGTGCTCCTGGAGACCTGGCCTGCAGGGAAAACCCGGA
Mouse_COL4A3	GACTCCTATGGATCAGAAAAGGTGCGCCTGGAGAGCCTGGTCTCGGGGCAAACTGGA
	* * * * *
Canine_Normal_5-18	AAAGATGGTGCCCTGGTTCCCTGGCACTGAGGGGCCCAAAGGCAACAGAGGCTTCCCT
Human_COL4A3	AAAGATGGTGTTCCTGGCTTCCCTGGAAGTGAGGGAGTCAAGGGCAACAGGGGTTCCT
Mouse_COL4A3	AAAGATGGTGCCCTGGCTTCCCTGGCACTGAGGGAGCCAAAGGCAACAGGGGCTTTCCT
	* * * * *
Canine_Normal_5-18	GGGTTACGGGTGAAGACGGCATTAAAGGGTGGAAGGGGACATTGGCCCGCCAGGATTT
Human_COL4A3	GGGTTAATGGGTGAAGATGGCATTAAAGGGACAGAAAGGGGACATTGGCCCTCCAGGATTT
Mouse_COL4A3	GGGTTGAGGGGTGAAGCTGGCATTAAAGGGAAGGAAAGGAGACATTGGCCCCCAGGATTT
	* * * * *
Canine_Normal_5-18	CGTGGTCCAACAGAATATTATGATGCATACCAGGAAAAGGGTGATGAAGGAATTCAGGC
Human_COL4A3	CGTGGTCCAACAGAATATTATGACACATACCAGGAAAAGGGAGATGAAGGCACTCCAGGC
Mouse_COL4A3	CCTGGTCCAACAGAATATTATGACGCATACCTGGAAGGGAGAGAGAGGAATGCCAGGC
	* * * * *
Canine_Normal_5-18	CCACCAGGCCCAAAGGAGCTCGTGGCCCCAGGGTCCCAGCGGTCCCCCTGGAGTTCCT
Human_COL4A3	CCACCAGGCCCCAGAGGAGCTCGTGGCCCCACAAGGTCCCAGTGGTCCCCCGAGTTCCT
Mouse_COL4A3	CTTCCTGGGCCCAAAGGAGCCCTGGCCACAGGGTCCAAGTGGTCCCCCTGGAGTCCCT
	* * * * *
Canine_Normal_5-18	GGAAGTGCTGGGTGCATCAAGCCTGGCCTCAGAGGAGCTCCTGGATCTCCAGGCATGAAA
Human_COL4A3	GGAAGTCTGGATGCATCAAGCCTGGCCTCAGAGGAGCCCCCTGGATGGCCAGGCTTGAAA
Mouse_COL4A3	GGAAGTCTGGACTGTCAAGCCTGGCCTCAGAGGACCATTTGGATGGCCAGGCTTGAAA
	* * * * *
Canine_Normal_5-18	GGACGTAAAGGGGAACAAGGGCCCCCGGAAAGAAATGCAGTGGGGCCTCCTGGGTCCCCA
Human_COL4A3	GGAAGTAAAGGGGAACGAGGCCGCCAGGAAAGGATGCCATGGGGACTCCTGGGTCCCCA
Mouse_COL4A3	GGGAGTAAAGGAGAGAGAGGACCCCTGGAAAGACACTGTGGGCCCTCCTGGACCCCTG
	* * * * *
Canine_Normal_5-18	GGTTGTCTGGTTACCAGGCCCCATAGGGCTGCCGGGATATCCAGGACCACCAGGTGAC
Human_COL4A3	GGTTGTCTGGTTACCAGGTCTTCCAGGATCACCGGACCTCCAGGACCGCCAGGTGAC
Mouse_COL4A3	GGATGTCTGGCTCACCAGGTCACACAGGCCCTCCAGGACCTCCAGGATGTCCAGGTGAC
	* * * * *
Canine_Normal_5-18	ATCGTGTTCGCAAAGGTCCACCTGGGGTTGGTGGACTCCCCGGCCATGTTGGGTTTCCA
Human_COL4A3	ATCGTTTTTCGCAAGGTCCACCTGGAGATCACGACTGCCAGGCTATCTAGGGTCTCCA
Mouse_COL4A3	ATTGTTTTT---AAATGTTCCCTGGTGAGCATGGAATGCCAGGTGACACAGGACCTCCA
	* * * * *
Canine_Normal_5-18	GGCATCCCAGGAGTCGAGGGGCCAAAGGGGAACCAGGCCTCTGTGCGTACAGTGTGCT
Human_COL4A3	GGAATCCCAGGAGTTGATGGGCCCAAAGGAGAACAGGCCTCCTGTGTACACAGTGCCCT
Mouse_COL4A3	GGAGTTCAGGACTTGATGGACCAAAGGAGAACAGGCAGTCCATGCACCGAGTGTAC
	* * * * *

Canine_Normal_5-18	TATATCCCAGGGCGTCCAGGTCTCCCAGGACTGCCAGGGTTGGATGGCATAAAAGGAATC
Human_COL4A3	TATATCCCAGGGCCTCCCGGTCTCCCAGGATTGCCAGGGTTACATGGTGTAAGGAATC
Mouse_COL4A3	TGCTTCCCAGGGCCCCCTGGAGTCCCAGGATTCCAGGATTAGATGGTATCAAAGGAATC
	* ***** * * ***** * ***** * ***** * *****
Canine_Normal_5-18	CCAGGAGGACAAGGGGCAGCTGGCATTAAAGGAAGCCAGGGTCCCCAGGAAGTGCGGGT
Human_COL4A3	CCAGGAAGACAAGGCGCAGCTGGCTTGAAAGGAAGCCAGGGTCCCCAGGAAATACAGGT
Mouse_COL4A3	CCAGGAGGACGAGGTGTGCCTGGTCTGAAGGGAACCCAGGGTCCCCAGGAAGTGCAGGT
	***** * * * * * * * * ***** * * *
Canine_Normal_5-18	CTTCCAGGATTTCAGGATTCCCGGGTGTCTCCGGGTGCTCCAGGACTTAAAGGAGAAAAA
Human_COL4A3	CTTCCAGGATTTCAGGTTCCTCCAGGTGCCAGGGTGACCCAGGACTTAAAGGAGAAAAA
Mouse_COL4A3	CTCCAGGATTTCAGGATTCCAGGTGACCAAGGGCATCCAGGACTTAAAGGGGACAAA
	** ***** ***** ***** * ** ***** * ** *
Canine_Normal_5-18	GGAGAAACATCTGAGCCTGAGGGACAGGTGGGTGCTCCAGGGGACCCCGGGCTCAGAGGC
Human_COL4A3	GGTGAACACTTCAGCCTGAGGGGCAAGTGGGTGTCCAGGTGACCCGGGGCTCAGAGGC
Mouse_COL4A3	GGGGATACACCTCTACCTGAGGGGCAAGTGGGTAAATCCAGGTGATCCTGGACTCAGAGGC
	** * * * * * * ***** ***** * * * *****
Canine_Normal_5-18	CATCCTGGAAGAAAGGGCTTGGATGGAATTCCTGGAATCCTGGAATTAAAGGACCACCA
Human_COL4A3	CAACCTGGGAGAAAGGGCTTGGATGGAATTCCTGGAATCCTGGGAGTGAAAGGATTACCA
Mouse_COL4A3	CTGCCCCGGGAGAAAGGGCTTCGATGGAATCCTGGAGGTCCAGGAGCGAAAGGACCACCA
	* * * ***** ***** ***** * * * ***** *****
Canine_Normal_5-18	GGACCCAAAGGTGAACCGGCCCTGAGTGGTGAGAAGGGGGACCAAGGTCTCCAGGGGAT
Human_COL4A3	GGACCTAAAGGCGAATCGGCTCTGAGTGGTGAGAAAGGGGACCAAGGTCTCCAGGGGAT
Mouse_COL4A3	GGACCTCAGGGCGAACCAGGCCCTGAGTGGAAAGGAAAGGGGACCAAGGACCTCCAGGACCT
	***** * * ***** * ***** ***** * ***** *
Canine_Normal_5-18	CCTGGGATCCCAGGGTCCCCAGGACCTGCAGGACCACCTGGACCACCAACTATGGACCA
Human_COL4A3	CCTGGCTCCCCTGGGTCCCCAGGACCTGCAGGACCAGCTGGACCACCTGGCTACGGACCC
Mouse_COL4A3	CCTGGATTCCCTGGACCCCCAGGACCTGCAGGACCAGCTGGACCACCAAGGCTATGGACCT
	***** * * * ***** ***** ***** * * *****
Canine_Normal_5-18	CAGGGAGAGCCTGGTCCAAGGGCACCCAAGGAGTTCCTGGAGCCCTGGACCACCTGGA
Human_COL4A3	CAAGGAGAACCTGGTCTCCAGGGCACGCAAGGAGTTCCTGGGGCCCCCGGACCACCCGGA
Mouse_COL4A3	CAAGGAGAGCCAGGTCCAAGGGAGCCCAAGGAGTCCCGGCGTCTCTGGGACCACCTGGA
	** ***** * ***** * ***** * * * ***** **
Canine_Normal_5-18	GAAGCCGGTCTTAAGGGAGAATTCGGTATTTTCGACACCAGTCCCAGGGCCCCCAGGACCT
Human_COL4A3	GAAGCCGGCCCTAGGGGAGAGCTCAGTGTTCACACACAGTTCAGGCCCTCCAGGACCT
Mouse_COL4A3	GAAGCCGGTCTTAAGGGAGAACCAGTACATCAACTCCAGATCTAGGTCCCCCAGGCCCT
	***** * * ***** * * * * ***** * * * ***** **
Canine_Normal_5-18	CCAGGGCCCCCTGGTTATGTCTGGCCCCGAGGTCCACCTGGTATCCCTGGATCCATAGGA
Human_COL4A3	CCAGGGCCCCCTGGCCATCTGGCCCCCAAGGTCCACCTGGTATCCCTGGATCCCTGGGG
Mouse_COL4A3	CCAGGGCCCCCTGGTCAGGCTGGCCCCAGAGGTCTACCTGGTTGCTGGACCGGTGGGA
	***** ***** * ***** ***** * ***** * **
Canine_Normal_5-18	AAATGTGGTGATCCGGGTCTTCTGGGCCTGATGGTGAACCAGGAATTCAGGAATTGGC
Human_COL4A3	AAATGTGAGATCCTGGTCTTCCAGGGCCTGATGGTGAACCAGGAATTCAGGAATTGGA
Mouse_COL4A3	AAATGTG---ATCCTGGTCTTCTGGACCTGATGGTGAAGCCAGGAATTCAGGAAGCTGGA
	***** ***** ***** * ***** ***** ***** * ***
Canine_Normal_5-18	TTCCCTGGGCCCCCTGGACCTAAGGGAGACCAAGGTTTTCAGGAGCAAAAGGAGCACCG
Human_COL4A3	TTTCTGGGCCACCTGGACCTAAGGGAGACCAAGGTTTTCAGGTACAAAAGGATCAGCTG
Mouse_COL4A3	TGCCCAGGGCCTCCAGGACCTAAGGGAAATCAAGGTTTCCAGGGACAAAAGGATCACCT
	* * ***** * ***** ***** * ***** ***** ***** **
Canine_Normal_5-18	GGTTGTCCAGGAGAAATGGGGAAGCCCGGGTCACTGGAGAACCAGGTCTCCAGGAGCC
Human_COL4A3	GGTTGTCTGGAAAAATGGGAGAGCCTGGGTACCTGGAAAGCCAGGCCTCCAGGAGCC
Mouse_COL4A3	GGTTGTCTGGGGAATGGGAAGCCTGGCCGTCTGGAGAACCAGGCATCCAGGAGCC
	***** * ***** ***** * ***** * ***** *****

Canine_Normal_5-18
Human_COL4A3
Mouse_COL4A3

AAGGGAGAACCAGGACTAGCCATACCTGGAGAACCAGGAATACCAGGTTTCCAGGAGAA
AAGGGAGAACCAGCAGTAGCCATGCCTGGAGGACCAGGAACACCAGGTTTCCAGGAGAA
AAGGGAGAACCATCCGTAGGCAGGCCTGGGAAACCAGGAAACCAGGTTTCCAGGAGAA

Canine_Normal_5-18
Human_COL4A3
Mouse_COL4A3

AGAGGCAATTCTGGGGAAATGGAGAAATTGGACTCCCTGGACTTCCAGGTCTCCCTGGA
AGAGGCAATTCTGGGGAAATGGAGAAATTGGACTCCCTGGACTTCCAGGTCTCCCTGGA
AGAGGCAATGCCGGGGAAATGGAGATATTGGACTCCCTGGGCTTCCAGGCCTCCCTGGA

Canine_Normal_5-18
Human_COL4A3
Mouse_COL4A3

ATTCCAGGAACGGAGGGCTTGATGGACCACCAGGGGATCCAGGGAAGCCTGGACCACCT
ACTCCAGGAAATGAAGGGCTTGATGGACCACGAGGAGATCCAGGGCAGCCTGGACCACCT
ACTCCAGGAAGAGCGGGCTTGACGGGCTCCAGGAGACCAGGACAGCCTGGATCACCT

Canine_Normal_5-18
Human_COL4A3
Mouse_COL4A3

GGAGAAAAAGGACCCCGAGGAAGTTGCACAGAGGGTCCAGAGGAGCCACGGACTTCCA
GGAGAACAAAGGACCCCGAGGAAGGTGCATAGAGGGTCCAGGGGAGCCCAAGGACTTCCA
GGAGCAAAGGATCCCGAGGAAGGTGCATACCAGGACCAGGGGACCCAAGGACTTCCC

Canine_Normal_5-18
Human_COL4A3
Mouse_COL4A3

GGATTAAATGGATTGGAAGGGCAACCAGGCAGAAGAGGTGAAACAGGGCCAAAAGGAGAC
GGCTTAAATGGATTGAAAGGGCAACAAGGCAGAAGAGGTAAACCGGGCCAAAGGGAGAC
GGCTTGAATGGATTGAAAGGGCAACCAGGTGGAAGAGGTGATACCGGGCCAAAGGGAGAC

Canine_Normal_5-18
Human_COL4A3
Mouse_COL4A3

CCAGGTATTCCAGGCTTGGATAGATCAGGCTTCTCTGGAGAACTGGACCACCAGGAATG
CCAGGAATTCCAGGCTTGGATAGATCAGGATTTCTCTGGAGAACTGGATCACCAGGAATT
CCTGGCATCCAGGCATGGACAGGTCTGGGGTCCCGGAGATCCTGGACCACCAGGAACA

Canine_Normal_5-18
Human_COL4A3
Mouse_COL4A3

CCAGGTCATCGAGGTGAGATAGGACCACCTGGCCAAAAAGGATATCCAGGAAATCCGGGA
CCAGGTCATCAAGGTGAAATGGGACCCTGGGTCAAAGAGGATATCCAGGAAATCCGGGA
CCAGGTTGCTTGGTGGATGGGACCACCTGGACAAAAAGGATATCCGGGAGCTCCAGGA

Canine_Normal_5-18
Human_COL4A3
Mouse_COL4A3

TTTTTAGGACTACCAGGTGAAGATGGAGTGATTGGGATGATGGGCTATCCAGGAAACACT
ATTTTAGGGCCACCAGGTGAAGATGGAGTGATTGGGATGATGGGATTTCTGGAGCCATT
TTCCAGGACCACCAGGCAGAGGAAGGAGAGGTTGGAATGATGGGCTATCCTGGAACTACT

Canine_Normal_5-18
Human_COL4A3
Mouse_COL4A3

GGCCCTCCTGGGCTCCCGGATCCAGGCACCCAGGACAGAGGGGTAGCTTTGGAATT
GGCCCTCAGGGCCCCCTGGGAACCCAGGCACACCAGGGCAGAGGGGGAGCCCTGGAATT
GGCCCTCCGGGGCTTCTGGGAAACCAGGCTCACAGGGGCAGCGAGGTAGCCTCGGAATC

Canine_Normal_5-18
Human_COL4A3
Mouse_COL4A3

CCAGGTGCAAAGGTTGAGAAAGGGCCCCAGGAGCCAAGGGGAAGAAGGAGAGAAAGGA
CCAGGAGTAAAGGGCCAGAGAGGAACCCAGGAGCCAAGGGGAACAAGGAGATAAAGGA
CCAGGAATGAAAGGGGAAAAAGGACGCCAGGAGCCAAGGCGAACGAGGAGAGAAAGGA

Canine_Normal_5-18
Human_COL4A3
Mouse_COL4A3

CCTATGGGGCTTGTCAAATAACAGCTCAGTGGGGGACAAAGGAGAACCAGGCCTCAA
AATCCCGGGCTTTCAGAGATATCCACGTAATAGGGGACAAAGGAGAACCAGGTCTCAA
AAACCAGGGCTTCTCAAACAACACTCTTAAAGGGAGACAAAGGAGAGCCTGGACTTAA

Canine_Normal_5-18
Human_COL4A3
Mouse_COL4A3

GGGTTTGCAGGAAAGCCGGGTGAAAAAGGAAACAGAGGCATTCCAGGGTTACCAAGTTT
GGATTTCGAGGAAATCCAGGTGAGAAAGGAAACAGAGGCGTTCCAGGGATGCCAGGTTTA
GGATTTGTGGGAATCCAGGTGAGAAAGGAAACAGAGGCAACCCAGGGTTACCAAGTCCG

Canine_Normal_5-18
Human_COL4A3
Mouse_COL4A3

AAAGGATTCGAAGGGCCACCTGGACCACCAGGCCCACCAGGCCCCGAGGAGATCCAGGC
AAGGGCCTCAAAGGACTACCCGAGCAGCAGGACCACCAGGCCCCAGAGGAGATTGGGC
AAAGGCCTCGAGGGATTGCTGGGCTACCAGGTCTCCAGGCCCTAGAGGAGATACGGGA

Canine_Normal_5-18	AGCCACGTGAAAGGCATAAAAGGAGACAAGGGACTCATGGGTGAGCCTGGCCCCAGAGGT
Human_COL4A3	AGTCATGTAATAGGCATAAAAGGAGACAAGGGTCTATGGGCCACCTGGCCCCAAAGGT
Mouse_COL4A3	CCTATCGGAAAAGGCATAAAAGGTGACAAGGATTTATGGGCCACCTGGCCCCAAAGGC
	* * * * *
Canine_Normal_5-18	CTACCTGGAACGTGGGAGACAAGGGGCCACCGGGTCAACCGGGAGCACCAGGGAGCCCA
Human_COL4A3	CCACCTGGAACGTGAGGAGACATGGGACCACCAGGTCGTCTGGGAGCACCAGTACTCCA
Mouse_COL4A3	CTGCCTGGAACGTAGGGGACATGGGCCACCAGGTTTCCCGGGAGCACCAGGTACCCCC
	* * * * *
Canine_Normal_5-18	GGTCTGCCAGGGCTCAGGGGCGATCCTGGATTCTATGGATTTCAGGTGTTAAAGGAGAG
Human_COL4A3	GGTCTTCCAGGACCCAGAGGTGATCCTGGATTCCAGGGGTTTCCAGGCGTGAAAGGAGAA
Mouse_COL4A3	GGTCTTCCCGGTGTCCAGAGGTGATCCAGGATTCCTTGGATTTCAGGCATAAAAGGAGAA
	* * * * *
Canine_Normal_5-18	AAGGGTAACCCAGGATTTCCGGGACCAGCTGGACCTCCAGGGCAAATTGGGCCAAAAGGA
Human_COL4A3	AAGGGTAATCCTGGATTTCTAGGATCCATTGGACCTCCAGGACCAATTGGGCCAAAAGGA
Mouse_COL4A3	AAGGGTAACCTGGATTTCTTGGGCCAATTGGACATCCAGGACCAGTTGGGCCCAAAGGA
	* * * * *
Canine_Normal_5-18	CCACCTGGTGTCCGTGGAGACCTTGGCACAGTTAAGATCATCTCCCTTCCAGGAAGCCCA
Human_COL4A3	CCACCTGGTGTACGTGGAGACCTTGGCACACTTAAGATTATCTCCCTTCCAGGAAGCCCA
Mouse_COL4A3	CCACAGGTCCACGTGGAACCTTGGCACTCTTAAGGTATCTCTCTTCCGGGAAGCCCA
	* * * * *
Canine_Normal_5-18	GGGCCACCTGGCCGTGTCTGGAGGACCGGGATGCCAGGAGAACCAGGGCCACCAGGGCCA
Human_COL4A3	GGGCCACCTGGCACACCTGGGAGAACAGGGATGCAGGGAGAACCTGGGCCACCAGGGCCA
Mouse_COL4A3	GGGCCACCTGGTGTACCTGGACAGCCAGGAATGAAAGGAGACCCTGGACCCCTGGGACTG
	* * * * *
Canine_Normal_5-18	CCAGGAATCCTTAGGACCTGTGGACCAAGAGGTAAACCAGGCATGGATGGAATACCAGGA
Human_COL4A3	CCTGGAACCTTAGGACCTGTGGGCCAAGAGGTAAGCCAGGCAAGGATGGAAAACCAGGA
Mouse_COL4A3	CCAGGAATCCCAGGACCTGTGGGCCAAGAGGTAAACCAGGCAAGGATGGGAAACCAGGA
	* * * * *
Canine_Normal_5-18	ACTCCTGGGCCAATTGGAGAAAAAGGCAACAAAGGTTGTAAAGGCGAGCAAGGACCACCT
Human_COL4A3	ACTCCTGGACCAGCTGGAGAAAAAGGCAACAAAGGTTCTAAAGGAGAGCCAGGACCAGCT
Mouse_COL4A3	ACTCCAGGACCAGCTGGTACAAAGGGCAACAAGGGCTTGAAAGGACAGCAAGGCCACCT
	* * * * *
Canine_Normal_5-18	GGATTAGATGGACTGCCAGGCTTGAAGGGGAGACCTGGAGACCCTGGACCACCTGCAACT
Human_COL4A3	GGATCAGATGGATTGCCAGGTTTGAAAGGAAAACGTGGAGACAGTGGATCACCTGCAACC
Mouse_COL4A3	GGTCTGGATGGATTGCCAGGCTTAAAGGGAAATCCTGGTGACAGGGGAACGCCGGCCACT
	* * * * *
Canine_Normal_5-18	GGGACCATGAAGAGGGGCTTTATCTTCACCAGGCATAGTCAGACCACAGTGATTCTCTCC
Human_COL4A3	TGGACAAC---GAGAGGCTTTGTCTTACCCGACACAGTCAAACCACAGCAATTCTCTCA
Mouse_COL4A3	GGTACAAGAATGCGAGGCTTCATCTTACCCGACACAGTCAAACCACGCCATTCTCTCA
	* * * * *
Canine_Normal_5-18	TGTCCAGAAGGGACAGCGCCACTCTATAGTGGCTTTTCTCTTTTTCATACAAGGAAAT
Human_COL4A3	TGTCCAGAGGGACAGTGCCACTCTACAGTGGGTTTTCTTTTCTTTTGTACAAGGAAAT
Mouse_COL4A3	TGCCCTGAAGGAACACAGCCACTCTATAGTGGGTTTTCTCTCTTTTGTACAAGGAAAC
	* * * * *
Canine_Normal_5-18	GAACAAGCCCATGGACAAGACTTGGGAACTCTCGGCAGCTGCCTGCAGCGATTACCACA
Human_COL4A3	CAACGAGCCCACGGACAAGACCTTGGAACTCTTGGCAGCTGCCTGCAGCGATTACCACA
Mouse_COL4A3	AAACGTGCACATGGACAAGACCTAGGTACTCTGGGCAGCTGCCTGCAGCGATTACCACA
	* * * * *
Canine_Normal_5-18	ATGCCATTCTTGTCTGTAACATCAACAATGTCTGTAATTTTGCATCTCGAAATGACTAC
Human_COL4A3	ATGCCATTCTTATCTGCAATGTCAATGATGTATGTAATTTTGCATCTCGAAATGATTAT
Mouse_COL4A3	ATGCCGTTCTTATCTGTAACATCAATAATGTATGTAACCTTGCATCAGCAATGATTAT
	* * * * *

Canine_Normal_5-18	TCATACTGGCTGTCAACACCAGCTCTGATGCCAATGGACATGGCTCCAATTACTGGCAGG
Human_COL4A3	TCATACTGGCTGTCAACACCAGCTCTGATGCCAATGAACATGGCTCCCATTACTGGCAGA
Mouse_COL4A3	TCATACTGGCTGTCAACACCAGCTCTGATGCCAATGGACATGGCTCCAATTAGTGGCAGA *****
Canine_Normal_5-18	GCCCTGGAGCCTTATATTAGCAGATGCACTGTCTGTGAAGGTCTACGATTGCCATAGCC
Human_COL4A3	GCCCTTGAGCCTTATATAAGCAGATGCACTGTTGTGAAGGTCTCGATCGCCATAGCC
Mouse_COL4A3	GCTCTCGAACCCTATATTAGCAGATGCACCGTCTGTGAAGGTCCAGCAATGGCCATAGCT ** ** *
Canine_Normal_5-18	ATTCACAGCCAAACCACTGACATTCCTCATGTCCCAATGGCTGGATTCTCTCTGGAAA
Human_COL4A3	GTTACAGCCAAACCACTGACATTCCTCCATGTCTCACGGCTGGATTCTCTCTGGAAA
Mouse_COL4A3	GTTACAGTCAAACCTACTGCTATCCCTCCGTGTCCCAGGACTGGGTTCTCTCTGGAAA *****
Canine_Normal_5-18	GGATTTTCGTTTATCATGTTCACAAGTGCAGGTTCTGAGGGTGCTGGGCAAGCACTGGCC
Human_COL4A3	GGATTTTCATTTCATCATGTTCACAAGTGCAGGTTCTGAGGGCACCGGGCAAGCACTGGCC
Mouse_COL4A3	GGTTTTTCTTTTATTATGTTCACAAGTGCAGGCTCTGAGGGTGCTGGACAAGCACTTGCC ** *****
Canine_Normal_5-18	TCCCCCGCTCCTGCCTGGAAGAGTTCGAGCCAGTCCGTTTATAGAATGTCACGGAAGA
Human_COL4A3	TCCCCTGGCTCCTGCCTGGAAGAATTCCGAGCCAGCCATTTCTAGAATGTCATGGAAGA
Mouse_COL4A3	TCGCCTGGCTCCTGCCTGGAAGAATTCCGAGCCAGTCCATTTATAGAATGCCATGGACGA ** ** *
Canine_Normal_5-18	GGAACGTGCAACTACTATTCAAATTCCTACAGTTTCTGGTTGGCTTCGTTAAACCCCAA
Human_COL4A3	GGAACGTGCAACTACTATTCAAATTCCTACAGTTTCTGGCTGGCTTCATTAAACCCAGAA
Mouse_COL4A3	GGGACATGTAACCTACTACTCAAATTCCTACAGTTTCTGGCTGGCTTCGCTGAACCCAGAA ** ** *
Canine_Normal_5-18	AGAATGTTTCAGAAAACCTATTCCATCAACTGTGAAAGCTG--AATCCATATGACTAGT-
Human_COL4A3	AGAATGTTTCAGAAAACCTATTCCATCAACTGTGAAAGCTGGGGAATTAGAAAAAATAATA
Mouse_COL4A3	AGAATGTTTCAGAAAACCTATTCCATCAACTGTGAAAGCTGGAGACTTAGAGAAAATCATATA *****
Canine_Normal_5-18	-----
Human_COL4A3	AGTCGCTGTCAGGTGTGCATGAAGAAAAGACACTGA
Mouse_COL4A3	AGCCGCTGTCAGGTGTGCATGAAGAAAAGACATTGA

APPENDIX 3

Canine COL4A3 amino acid sequence alignments

Consensus key (see documentation for details)

- * - single, fully conserved residue
- : - conservation of strong groups
- . - conservation of weak groups
- no consensus

CLUSTAL W (1.81) multiple sequence alignment

Canine_A3_Normal_5-18_Transl	-----
Human_COL4A3_Translated_-Fr	MSARTAPRPQVLLPLLLVLLAAAPAASKGCVCKDKGQCFCDGAKGEKGE
Mouse_COL4A3_Translated_-Fr	MHSKTAPRFLVFLLLTLLLLLAASPVASKGCVCKGKQCLCAGTKGEKGE

Canine_A3_Normal_5-18_Transl	-----
Human_COL4A3_Translated_-_Fr	KGFPGPPGSPGQKGFTGPEGLPGPQGPKGFPGLPGLTGSKGVRGISGLPG
Mouse_COL4A3_Translated_-_Fr	KGVPGSPGFPQGKGFPGPEGLPGPQGPKGSPGLPGLTGPKGIRGITGLPG

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Canine_A3_Normal_5-18_Transl      -----NFKKEIYHGDGVPNGCSKGEGQFPGPLPGTPGYPGIPGAV
Human_COL4A3_Translated_-Fr      FSGSPGLPGTPGNTGPYGLVGVPGCSGSKGEGQFPGPLPGTPGYPGIPGAA
Mouse_COL4A3_Translated_-Fr      FAGPPGLPGLPGHPGRGLAGLPGCSKGEGQFPFGPPTGYAGLPGPD
                                   :*** *****:*****:***

```

Canine_A3_Normal_5-18_Transl	GLKGEKGAPATGEGIELDGRGDPGLPGAPGFQGLPGLPGFPGFAGPPGPP
Human_COL4A3_Translated_-Fr	GLKGQKGAPAKGEDIELDAKGDPGLPGAPGFQGLPGPPGFPFVPGPPPP
Mouse_COL4A3_Translated_-Fr	GLKGQKGAPAQGEDRGFNKGKDPGPPGVPGFQGFPLPGFPGFAGPPPPP
	***** ** ** : : : : : * * * * * * * * * * * * * * * *

Canine_A3_Normal_5-18_Transl	GFLGFPGAMGPPGPKGQMGDKVIGQKGKRGVKGLTGPPGPPGTIVITLTG
Human_COL4A3_Translated_-Fr	GFFGFPGAMGRPRGPKGHMGERVIGHKGKRGVKGLTGPPGPPGTIVITLTG
Mouse_COL4A3_Translated_-Fr	GFFGLPGAMGRPRGPKGHMGDSVIGQKGKRGMKGLTGPPGPPGTIVITLTQ
	:** ***:***:*****:*****:*****:***

Canine_A3_Normal_5-18_Transl	PDNRTDLKGEKGDKGAMGKP GPPGLP PPGESYSGSEKGAPGEPGPQ GKPG
Human_COL4A3_Translated_-Fr	PDNRTDLKGEKGDKGAMGEP GPPGPSL PGESYSGSEKGAPGDPLG QGKPG
Mouse_COL4A3_Translated_-Fr	PYNKSDFKGEKGDEGERGEP GP PPSGPPGDSYSGSEKGAPGEPGRPKPG
	* * : * : ***** : * * : ***** : * : ***** : * : *****

Canine_A3_Normal_5-18_Transl	KDGAPGFPGTGEKGNRGFPGLRGEDGIKGWKGDIGPPGFRGPTEYYDAY
Human_COL4A3_Translated_-Fr	KDGVPGFPGSGEVKGNRGFPLMGEDGIKGQKGDIGPPGFRGPTEYYDTY
Mouse_COL4A3_Translated_-Fr	KDGAPGFPGTGEAKGNRGFPGLRGEAGIKGRKGDIGPPGFPGPTEYYDAY
	* * * * *

Canine_A3_Normal_5-18_Transl	QEKGDEGIPGGPPGPKARGPQPSPGPPGVLSAGSSKPLRLGAPGPSPMKM
Human_COL4A3_Translated_-Fr	QEKGDGETPGPPGPRGARGPQGPSPPGVPSGSSRPGLRGAPGWGPLK
Mouse_COL4A3_Translated_-Fr	LEKGERGMPLGP GKARGPQPSPGPPGVPSGPLSRPLRLGPIGWPLKL ***: **::***** ** *:****:

Canine_A3_Normal_5-18_Transl	GRKGEGQPPGKNAVGGPPGSPGCGSPGPIGLPGYPGPPGDIVFRKGPPGV
Human_COL4A3_Translated_-Fr	GSKGERGRPGKDMGTPGSPGCAAGSPGLPGSPGPPGPGDIVFRKGPPGD
Mouse_COL4A3_Translated_-Fr	GSKGERGPPGKDTVGPPGGLGCPGSPGPPGPPGPPGCPGDIVFKCSP-GE
	* * * * * : * * * * * : * * * * * : * * * * * : * *

Canine_A3_Normal_5-18_Transl GGLPGHVGFPGIPGVGEVKPEGLLCVCQAYIPGRGLPLGLDGIKGI
Human_COL4A3_Translated_-Fr HGLPGYLGSPGIPGVDGPKGEPGLLCQCQPIYPGPPGLPLGLHGVKGI
Mouse_COL4A3_Translated_-Fr HGMPGDTGPPGVPGLDGPKGEPGSPCTECHCFPPGPPGVPGFPLDGIKGI

Canine_A3_Normal_5-18_Transl
Human_COL4A3_Translated_-_Fr
Mouse_COL4A3_Translated_-_Fr

PGGQGAAGIKGSPGSPGSAGLPGFPGFPGAPGAPGLKGEKGSETSEPEGQV
PGRQGAAGLKSGSPSPGNTGLPGFPGFPGAQDPGLKGEKGETLQPEGQV
PGRGRVAVPLKGPNPSPGSAGLPGFAGFPGDQGHPLKGDKGDTPLPWQV
* * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * *

Canine_A3_Normal_5-18_Transl
Human_COL4A3_Translated_-_Fr
Mouse_COL4A3_Translated_-_Fr

GAPGDPGLRGHPGRKGLDGIPGTPIKGGPPGPKGEFALSSEKGDQGGPGD
GVPGDPGLRQGPRKGDLGIPTLGVKGLPGRPGEALSSSEKGDQGGPGD
GNPDGPGLRLPLGRKGFDTGTPGGPGAKGPPGPQEPALESSSRKGDQGGPPG

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Canine_A3_Normal_5-18_Transl
Human_COL4A3_Translated_-_Fr
Mouse_COL4A3_Translated_-_Fr

PGIPGSPGPAGPPGPPNYGPGQGEFPGPKGTQGVPGAPGPPGEAGPKGEFGI
PGSPGSPGPAGPAGPPGYGPGQGEFGLQGTQGVPGAPGPPGEAGPRGELSV
PGFPGPPPGAGPAGPPGYGPGQGEFPGPKAQGVPGVGLGPPGEAGLKGEPST

Canine_A3_Normal_5-18_Transl
Human_COL4A3_Translated_-_Fr
Mouse_COL4A3_Translated_-_Fr

STPVPGPPPGPPGPPGYAGPRGPPGIPGSIGKCGDPGLPGPDGEFPIPGIG
STPVPGPPPGPPGPPGHPGPQGPPIPGSLGKCGDPGLPGPDGEFPIPGIG
STPDLGPPGPPGPPGQAGPRGLPLGLPGPVGKC-DPGLPGPDGEFPIPEAG
*** **** * * * * * : * : * : * : * : * : * : * : * : *

Canine_A3_Normal_5-18_Transl
Human_COL4A3_Translated_-_Fr
Mouse_COL4A3_Translated_-_Fr

FPGPPGPKGDQGFPGAAGAPGCPGEMGKPGSPGEPLGAKEPGLAIPG
FPGPPGPKGDQGFPGTKSLGCPGKMGEPLPGKPGLPGAEGEPAVAMPG
CPGPPGPKNQGFPGFTKGSPGCGEMGKPGRPEGPPIGAKEGPSVGRPG
*****:*****:***:****:*:**:*****:***:*

Canine_A3_Normal_5-18_Transl
Human_COL4A3_Translated_-_Fr
Mouse_COL4A3_Translated_-_Fr

EPGIPGFPPGERNSGENGEI GLPGLPGLPGIPGTGGLDGGPPGDPGKPGPP
GPPTPGFPGERNSGEHGEI GLPGLPGLPGTPGNEGLDGGPRDGPQPGPP
KPGKPGFPFGERNAGENGEDIGLPGLPGLPGTPGRGGLDGGPPGDPGQPGSP
* * * * *
* * * * *

Canine_A3_Normal_5-18_Transl
Human_COL4A3_Translated_-Fr
Mouse_COL4A3_Translated_-Fr

GEKGGPPGSCTEGPRGAHGLPLGNLLEGQPGRRGETGPKGDGPIGPLDRSG
GEQGGPPGRCEI GPRGAQGLPLGNLKGQQGRRGKTGPKGDGPIGPLDRSG
GAKGSPGRCIPGPRGTQGLPLGNLKGQPGRRGDTGPKGDGPIGPMDRSG
* : * : * : * : * : * : * : * : * : * : * : * : * : *

Canine_A3_Normal_5-18_Transl
Human_COL4A3_Translated_-Fr
Mouse_COL4A3_Translated_-Fr

FPGEFGPPGMPGHRGEIGPPGQKGYPGNPGFLGLPGEDGVI GMMGYPGNT
 FPGETGSPGIPGHQGEMGPLGQRGYPGNPGILGPPGEDGVI GMMGFPGA
 VPGDPPGPPGTGCPGEMGPPGQKGYPGAPGFPGPPEKEGEVGM GYPGTT
 ** : * * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * * : * * * * *

Canine_A3_Normal_5-18_Transl
Human_COL4A3_Translated_-Fr
Mouse_COL4A3_Translated_-Fr

GPPGPPGPIPGTPGQGRSFGIPGAKGEGKPPGAKGEEGEKGPMPGCQITSS
 GPPGPPGNPGTPGQGRSPGIPGVKGQRGTPGAKGEQGDKGNPSPSEISHV
 GPPGLPGKPGSGQGRSLGIPGMKGKGRPGAKGERGEKGKPGPSQTTLL

Canine_A3_Normal_5-18_Transl
Human_COL4A3_Translated_-Fr
Mouse COL4A3 Translated - Fr

VGDKGEPGLKGFGAGKPGGEKGNRGIPGLPGLKGFEGPPGPPGPPGPRGDPG
IGDKGEPGLKGFGAGNPGEKGNRGVPGMPGLKGLKGLPGPAGPPGPRGDLG
KGDKGEPGLKGFGVGNPGEKGNRGNPGLPGPKGLEGLPGLPGPPGPRGDTG

Canine_A3_Normal_5-18_Transl
Human_COL4A3_Translated_-Fr
Mouse COL4A3 Translated - Fr

SIGNPGEAGPRGSPGNMGMNMGMPGLKGHRGPLGLPGVTRGRPGLPGVHGLQ
STGNPGEPLGRGTPGSMGMNMGMPGSKGKRGTLGFPGRAGRPLPGIHLQ
SRGNPGRPGPHGMPGSMGIMGVPGPKGRKGTSLGPLAGRPGRGTGIHPGQ
* * * * * : * * * * * : * * * * * : * * * * * : * * * * *

Canine_A3_Normal_5-18_Transl
Human_COL4A3_Translated_-Fr
Mouse COL4A3 Translated - Fr

GDKGEPGYSAGTRPGPPGLKGDPGLPGAMGRKGERGLPGTPGHSGPAGTE
GDKGEPGYSEGTRPGPPGPTGDPGLPGDMGKKGEMGQPGPPGHLGPAGPE
GDKGEPGYSEGARPGPPGPKGDPLPGDKGKKGERGVPGPPGQSGPAGPD
* :

Canine_A3_Normal_5-18_Transl
Human_COL4A3_Translated_-Fr
Mouse COL4A3 Translated - Fr

GAPGHPGSPGHPGKPGPDGDLGLKGIGFGPSGVKGGPPGPGILGSPGP
GAPGSPGSPGLPGKPGPHGDLGFGIKGLLGPPGIRGPPGLPGFPGSPGP
GAPGSPGSPGHPGKPGPDGLGLKGQKGFPGPPGGTGGPPGPGPLGPLGP

| | |
|------------------------------|--|
| Canine_A3_Normal_5-18_Transl | MGMRGNQGRDGIPGPPGEKGETGLLGAHPGPGGSPGPPGAKGDRGAPGLP |
| Human_COL4A3_Translated_-_Fr | MGIRGDQGRDGIPGPPAGEKGETGLLRAPPGRGNPGAQGAAGDRGAPGFP |
| Mouse_COL4A3_Translated_-_Fr | MGMRGDQGRDGIPGPPGEKGETGLLGAYPGPKGSPGVPGAKGDRGVPGLS
*:**:*****.***** * ** *.* ** *****.**:. |
| Canine_A3_Normal_5-18_Transl | GPPGRKGAVGDAGPRGPTGMTGLPPPPFGPAVIPGQKGNQGGPFGRGNP |
| Human_COL4A3_Translated_-_Fr | GLPGRKGAMGDAGPRGPTGIEGFPGPPGLPGAIIPGQTGNRGGPPSRGSP |
| Mouse_COL4A3_Translated_-_Fr | GLPGRKGMVDVGPQGPPGTAGLPGLPGAIIPGPKGDRGLPGLRGNP
* *****.:**.*:***.* *:*****:***:***. **::* ** **.* |
| Canine_A3_Normal_5-18_Transl | GEPGPPGPPGSHVKGIKGDKGLMGEPGRGLPGTVGDKGPPGQPGAPGSP |
| Human_COL4A3_Translated_-_Fr | GAPGPPGPPGSHVIGIKGDKGSMDHGPKGPPGTAGDMGPPGRLGAPGTP |
| Mouse_COL4A3_Translated_-_Fr | GEPGPPGPPGPIKGIKGDKGMGPPGPKGLPGTVGDMGPPGFPGAPGTP
* *****. ***** ** ***: ** **.* ** ** ** ** *****:* |
| Canine_A3_Normal_5-18_Transl | GLPGLRGDPGFYGFPGVKGEKGNPFGPPGAGPPQIGPKGPPGVRGDPGT |
| Human_COL4A3_Translated_-_Fr | GLPGPRGDPGFQGFPGVKGEKGNPGLSGISPPGPIGPKGPPGVRGDPGT |
| Mouse_COL4A3_Translated_-_Fr | GLPGVVRGDPGFPGFPGIKGEKGNPGLPPIGHPGPVGPKGPPGPRGKPGT
**** ***** *****:***** * . * ** :***** ** .*** |
| Canine_A3_Normal_5-18_Transl | VKIISLPGSPGPPGRAGGPGMPGEPGPPGPPGILGPCGPRGKPGMDGIPG |
| Human_COL4A3_Translated_-_Fr | LKIIISLPGSPGPPGTPGEPGMQGEPPGPPGNLGPCGPRGKPGKDGP |
| Mouse_COL4A3_Translated_-_Fr | LKVISLPGSPGPPGVPGQPGMKGDGPGPLGLPGIPGPCGPRGKPGKDGP
*:*****. * ** *:*** * ** ***** ** ** |
| Canine_A3_Normal_5-18_Transl | TPGPIGEKGKNGCKGEQGGPGLDGLPGLKGRPGDPGPATGTMRGFIPT |
| Human_COL4A3_Translated_-_Fr | TPGPAGEKGKNGSKGEPGAGSDGLPGLKGRGDSGSPATWTTR-GFVFT |
| Mouse_COL4A3_Translated_-_Fr | TPGPAGTKGNKGLKGQQGPPGLDGLPGLKGNPGDRGTPATGTMRGFIPT
**** * ***** **: *.* *****. ** *.* ** ***:** |
| Canine_A3_Normal_5-18_Transl | RHSQTTVIPSCPEGTAPLYSGFSLFLFIQNEQAHGQDLGTLGSLQRFTT |
| Human_COL4A3_Translated_-_Fr | RHSQTTAIPSCPEGTVPVLYSGFSFLFVQGNQRAHGQDLGTLGSLQRFTT |
| Mouse_COL4A3_Translated_-_Fr | RHSQTTAIPSCPEGTQPLYSGFSLFLVQGNKRAHGQDLGTLGSLQRFTT
*****.***** *****:***:***:*****:***** |
| Canine_A3_Normal_5-18_Transl | MPFLFCNINNVCNFASTRNDYSYWLSTPALMPMDMAPITGRALEPYISRCT |
| Human_COL4A3_Translated_-_Fr | MPFLFCNVNDVCNFASTRNDYSYWLSTPALMPMNMAPITGRALEPYISRCT |
| Mouse_COL4A3_Translated_-_Fr | MPFLFCNINNVCNFASTRNDYSYWLSTPALMPMDMAPISGRALEPYISRCT
*****:***:*****:*****:*****:*****:***** |
| Canine_A3_Normal_5-18_Transl | VCEGPTIAIAIHSQTTDIPSCPNGWISLWKGSFIMFTSAGSEGAGQALA |
| Human_COL4A3_Translated_-_Fr | VCEGPAIAIAVHSQTTDIPPCPHGWISLWKGSFIMFTSAGSEGAGQALA |
| Mouse_COL4A3_Translated_-_Fr | VCEGPAMAIHAVHSQTTAIPPCPDWVSLWKGSFIMFTSAGSEGAGQALA
*****:***:***** **.*:.*:*****:*****:***** |
| Canine_A3_Normal_5-18_Transl | SPGSCLEEFRASPFIECHGRGTCNYYSNSYSFWLASLNPQRMFRKPIPT |
| Human_COL4A3_Translated_-_Fr | SPGSCLEEFRASPFIECHGRGTCNYYSNSYSFWLASLNPQRMFRKPIPT |
| Mouse_COL4A3_Translated_-_Fr | SPGSCLEEFRASPFIECHGRGTCNYYSNSYSFWLASLNPQRMFRKPIPT
*****:*****:*****:*****:*****:***** |
| Canine_A3_Normal_5-18_Transl | VKA---ESIL----- |
| Human_COL4A3_Translated_-_Fr | VKAGELEKIIISRCQVCMKKRH |
| Mouse_COL4A3_Translated_-_Fr | VKAGDLEKIIISRCQVCMKKRH
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APPENDIX 4

Canine COL4A4 nucleotide sequence

Sequence Range: 1 to 5163

| | | | | |
|-------|-------|-------|-------|-------|
| 10 | 20 | 30 | 40 | 50 |
| CTAGA | GGATC | TACTA | GTCAT | ATGGA |
| GATCT | CCTAG | ATGAT | CAGTA | TACCT |
| 60 | 70 | 80 | 90 | 100 |
| GAAAC | CTGAC | AATCT | TTTTT | AAGTC |
| CTTTG | GACTG | TTAGA | AAAAA | TTCAG |
| 110 | 120 | 130 | 140 | 150 |
| ATGCA | CATGG | CATCA | GTAAA | GTGTT |
| TACGT | GTACC | GTAGT | CATTT | CACAA |
| 160 | 170 | 180 | 190 | 200 |
| CAGAG | ATCCC | TGGTC | ACTTA | TATTT |
| GTCTC | TAGGG | ACCAG | TGAAT | ATAAA |
| 210 | 220 | 230 | 240 | 250 |
| ATGGG | AGTGG | AAAGA | AGTTT | GTCGG |
| TACCC | TCACC | TTTCT | TCAAA | CAGCC |
| 260 | 270 | 280 | 290 | 300 |
| GTGTG | CCATT | GCTTT | CCTGA | AAAAG |
| CACAC | GGTAA | CGAAA | GGACT | TTTTT |
| 310 | 320 | 330 | 340 | 350 |
| GGGGC | CACAG | GGTCC | CATTG | GACCC |
| CCCCG | GTGTC | CCAGG | GTAAC | CTGGG |
| 360 | 370 | 380 | 390 | 400 |
| TTCCA | GGAGA | GAAAG | GGATG | AGAGG |
| AAGGT | CCTCT | CTTTC | CCTAC | TCTCC |
| 410 | 420 | 430 | 440 | 450 |
| GGTGA | CAAAG | GTGAC | AAGGG | TCCAA |
| CCACT | GTTTC | CACTG | TTCCC | AGGTT |
| 460 | 470 | 480 | 490 | 500 |
| GGATG | GCATA | CCTGG | CCACC | CGGGG |
| CCTAC | CGTAT | GGACC | GGTGG | GCCCC |
| 510 | 520 | 530 | 540 | 550 |
| GCATG | CATGG | CTACA | ATGGT | TCACG |
| CGTAC | GTACC | GATGT | TACCA | AGTGC |
| 560 | 570 | 580 | 590 | 600 |
| AGAGG | AGTTC | CTGGC | CCAGG | AGGCC |
| TCTCC | TCAAG | GACCG | GGTCC | TCCGG |
| 610 | 620 | 630 | 640 | 650 |
| AGAAA | AAGGA | AATTC | AGTGT | TCATT |
| TCTTT | TTCTT | TTAAG | TCACA | AGTAA |

| | | | | |
|---|------|------|------|------|
| 660 | 670 | 680 | 690 | 700 |
| GAGAC AGGGG GGACC CAGAC CTGCC TGGCT TGCCG GGACC GAAGG GGGCA | | | | |
| CTCTG TCCCC CCTGG GTCTG GACGG ACCGA ACGGC CCTGG CTTCC CCCGT | | | | |
| 710 | 720 | 730 | 740 | 750 |
| GGAGG ACCGG CAGGT CCCAA GGGAC AGCCA GGAGA GCCTG GGTTA ACAGG | | | | |
| CCTCC TGGCC GTCCA GGGTT CCCTG TCGGT CCTCT CGGAC CCAAT TGTCC | | | | |
| 760 | 770 | 780 | 790 | 800 |
| TGCTC CGGGC CATCC TGGGA GACCA GGCTT GAAGG GTAAT CCTGG TGTGG | | | | |
| ACGAG GCCCG GTAGG ACCCT CTGGT CCGAA CTTCC CATTG GGACC ACACC | | | | |
| 810 | 820 | 830 | 840 | 850 |
| GAGTA AAGGG GCAAA TGGGA GACCC GGGTG AGGTT GGCCA GCAGG GTTCT | | | | |
| CTCAT TTCCC CGTTT ACCCT CTGGG CCCAC TCCAA CCGGT CGTCC CAAGA | | | | |
| 860 | 870 | 880 | 890 | 900 |
| CCTGG ACCCA CCTTA TTGGT ACAGC CACCT GATTG CTGTC TGTAT AAAGG | | | | |
| GGACC TGGGT GGAAT AACCA TGTCG GTGGA CTAAG GACAG ACATA TTTCC | | | | |
| 910 | 920 | 930 | 940 | 950 |
| AGAAA AGGGC ATAAA AGGAA TGCCT GGTAT GATTG GACCT CCAGG ACTAC | | | | |
| TCTTT TCCCG TATTT TCCTT ACGGA CCATA CTAAC CTGGA GTGCC TGATG | | | | |
| 960 | 970 | 980 | 990 | 1000 |
| CGGGA CCCAA GGGAG AACCT GGAAT TGGAC CAAAA GGAGA GAAGG GTATT | | | | |
| GCCCT GGGTT CCCTC TTGGA CCTTA ACCTG GTTTT CCTCT CTTCC CATAA | | | | |
| 1010 | 1020 | 1030 | 1040 | 1050 |
| CCTGG GTTCT CAGGA CCTCG GGGTG ATCCG GGTTT CTATG GCTCT CCAGG | | | | |
| GGACC CAAGA GTCCT GGAGC CCCAC TAGGC CCAAG GATAC CGAGA GGTCC | | | | |
| 1060 | 1070 | 1080 | 1090 | 1100 |
| TTTTT CAGGA TTAAA GGGGA AACCA GGAAT GTTTG GAGAT CCTGG ATCAT | | | | |
| AAAAG GTCCT AATTT CCCCT TTGGT CCTGA CAAAC CTCTA GGACC TAGTA | | | | |
| 1110 | 1120 | 1130 | 1140 | 1150 |
| TTGGA TTTGT TGGCC CAAAG GGGGA TCCTG GAGAC CGTGG GTACC CAGGA | | | | |
| AACCT AAACA ACCGG GTTTC CCCCT AGGAC CTCTG GCACC CATGG GTCCT | | | | |
| 1160 | 1170 | 1180 | 1190 | 1200 |
| CCACC GGGTG TTCTG GTAAC TCCAT CTGTT CCACT CAAAG GCCCT CCAGG | | | | |
| GGTGG CCCAC AAGAC CATTG AGGTA GACAA GGTGA GTTTC CGGGA GGTCC | | | | |
| 1210 | 1220 | 1230 | 1240 | 1250 |
| GGATC CGGGG CGCCC TGGCC ACTAT GGAGA AATGG GGTCC GTTGG ACCAC | | | | |
| CCTAG GCCCC GCGGG ACCGG TGATA CCTCT TTACC CCAGG CAACC TGGTG | | | | |
| 1260 | 1270 | 1280 | 1290 | 1300 |
| CTGGT CCCCC TGGTC CCCCA GGTCC ACCAG GGGAA GACTG TGCAG GCATG | | | | |
| GACCA GGGGG ACCAG GGGGT CCAGG TGGTC CCCTT CTGAC ACGTC CGTAC | | | | |
| 1310 | 1320 | 1330 | 1340 | 1350 |
| ATGGG ACCCC CTGGA CCAAG AGGAT TTCCT GGTCA TCCAG GATTT CCAGG | | | | |
| TACCC TGGGG GACCT GGTTC TCCTA AAGGA CCAGT AGGTC CTAAA GGTCC | | | | |

| | | | | |
|-------------------|-------------------|-------------------|-------------------|-------------------|
| 1360 | 1370 | 1380 | 1390 | 1400 |
| GGCAG CTGGT ATCCC | AGGGA GAGCG GATTT | AAGTC CAGGA AAACC | AGGGA CCGTC GACCA | TAGGG TCCCT CTCGC |
| CTAAA TTCAG | GTCCT TTTGG | TCCCT | | |
| 1410 | 1420 | 1430 | 1440 | 1450 |
| AACCG GGACC TCCTG | GGTTG CCCGG AGCAC | CAGGG CTGCA AGGAC | CCCCA TTGGC | CCTGG AGGAC |
| CCAAC GGGCC | TCGTG GTCCC | GACGT TCCTG | GGGGT | |
| 1460 | 1470 | 1480 | 1490 | 1500 |
| GGATC AGATG TTATC | TATTG TAGTG TTGGG | CATCC TGGAC CACAA | GGGAT CCTAG | TCTAC |
| AATAG ATAAC | ATCAC AACCC | GTAGG ACCTG | GTGTT | CCCTA |
| 1510 | 1520 | 1530 | 1540 | 1550 |
| AAAAG GCAAA ATGGG | TCCTC CAGGA AGAAG | AGGCT CGAAA | GGAGA AAAAG | TTTTC |
| CGTTT TACCC | AGGAG GTCCT | TCTTC TCCGA | GCTTT CCTCT | TTTTC |
| 1560 | 1570 | 1580 | 1590 | 1600 |
| GAAAT GCGGG GCTCT | GTGCC TGTGA GCCTG | GTCCC ATGGG | CCCAC CAGGC | CTTTA |
| CGCCC CGAGA | CACGG ACACT | CGGAC CAGGG | TACCC GGGTG | GTCCC |
| 1610 | 1620 | 1630 | 1640 | 1650 |
| CCTCC AGGAC TTCCT | GGAAG GCAGG GTAGT | AAGGG AGACT | TGGGG CTCCC | GGAGG |
| TCCTG AAGGA | CCTTC CGTCC | CATCA TTCCC | TCTGA ACCCC | GAGGG |
| 1660 | 1670 | 1680 | 1690 | 1700 |
| TGGGT GGATT GGAGA | GAAAG GTCAC CCAGG | CCCTC CTGGT | GCTGA AGGAT | ACCCA |
| CCTAA CCTCT | CTTTC CAGTG | GGTCC GGGAG | GACCA CGACT | TCCTA |
| 1710 | 1720 | 1730 | 1740 | 1750 |
| CTCCA GGATC ACCAG | GAAAA CATGG CGCCT | CAGGA CCACC | TGGCA GCAAA | GAGGT |
| CCTAG TGGTC | CTTTT GTACC | GCGGA GTCCT | GGTGG ACCGT | CGTTT |
| 1760 | 1770 | 1780 | 1790 | 1800 |
| GGAGA AAAGG GCGAC | AGGGT TGTAT CAAGA | GTGAA AGGGA | AAAAA GGAGA | CCTCT |
| TTTCC CGCTG | TCCCA ACATA | GTTCT CACTT | TCCCT TTTTT | CCTCT |
| 1810 | 1820 | 1830 | 1840 | 1850 |
| AAGAG GTCCT GATGG | GCTCC CAGGA TTTCC | AGGAC AACAG | GGACA AAATG | TTCTC |
| CAGGA CTACC | CGAGG GTCCT | AAAGG TCCTG | TTGTC CCTGT | TTTAC |
| 1860 | 1870 | 1880 | 1890 | 1900 |
| GTCGG GATGG ACTTC | CTGGA AAAAA | GGGGG ATCCA | GGCCC CCCGG | GGGGT |
| CAGCC CTACC | TGAAG GACCT | TTTTT CCCCC | TAGGT CCGGG | GGGCC |
| 1910 | 1920 | 1930 | 1940 | 1950 |
| CATGA GGATG CAGCC | CCAGG TGCTA AAGGG | TCTCC TGGAC | CACCG GGGCC | GGTAC |
| GTACT CCTAC | GTCGG GGTCC | ACGAT TTCCC | AGAGG ACCTG | GTGGC |
| 1960 | 1970 | 1980 | 1990 | 2000 |
| CCCAG GCAGA ACAGG | ACCCA GGGGA | CAACC AGGTC | TGGGA TTTCC | TGGTC |
| GGGTC CGTCT | TGTCC TGGGT | CCCCT GTTGG | TCCAG ACCCT | AAAGG |
| 2010 | 2020 | 2030 | 2040 | 2050 |
| CACCA GGAGA GAGAG | GACAA CCAGG | AGCTC CAGGC | CGCCC TGGCG | AGAGG |
| GTGGT CCTCT | CTCTC CTGTT | GGTCC TCGAG | GTCCG GCGGG | ACCGC |
| | | | | TCTCC |

| | | | | |
|---|------|------|------|------|
| 2060 | 2070 | 2080 | 2090 | 2100 |
| GGCTT CGGGG GCTTG AAGGG CCAGA AAGGT GATAC AATTT CTTGT AATGT | | | | |
| CCGAA GCCCC CGAAC TTCCC GTCTT TTCCA CTATG TTAAA GAACA TTACA | | | | |
| 2110 | 2120 | 2130 | 2140 | 2150 |
| CACCT ACCCT GGGAG GCCAG GACCC CCAGG TTTTG ATGGA CCTCC AGGAC | | | | |
| GTGGA TGGGA CCCTC CGGTC CTGGG GGTCC AAAAC TACCT GGAGG TCCTG | | | | |
| 2160 | 2170 | 2180 | 2190 | 2200 |
| CAAAG GGATT TCCAG GTCCT CAAGG CGCTC CAGGG TTGAG GTGCT TGGAT | | | | |
| GTTTC CCTAA AGGTC CAGGA GTTCC GCGAG GTCCC AACTC CACGA ACCTA | | | | |
| 2210 | 2220 | 2230 | 2240 | 2250 |
| GGGCC AAAAG GTCAG CGTGG CAAAC CAGGA ATGTC AGAGA TACCC GGTCC | | | | |
| CCCGG TTTTC CAGTC GCACC GTTTG GTCCT TACAG TCTCT ATGGG CCAGG | | | | |
| 2260 | 2270 | 2280 | 2290 | 2300 |
| ACCTG GTTTT CGTGG TGACA TGGGA GATCC GGGTT TTGAA GGTGA AAAGG | | | | |
| TGGAC CAAAA GCACC ACTGT ACCCT CTAGG CCCAA AACTT CCACT TTTCC | | | | |
| 2310 | 2320 | 2330 | 2340 | 2350 |
| GGTCC TCCCT TCTTG GGCCG CCAGG CTTTC CTGGT TCTCG TGGAG CAAAT | | | | |
| CCAGG AGGGA AGAAC CCGGC GGTCC GAAAG GACCA AGAGC ACCTC GTTTA | | | | |
| 2360 | 2370 | 2380 | 2390 | 2400 |
| GGTCA GAAAG GAATG ATGGG AGACA TTGCC TATGG CCTCC CAGGC CCCCC | | | | |
| CCAGT CTTTC CTTAC TACCC TCTGT AACGG ATACC GGAGG GTCCG GGGGG | | | | |
| 2410 | 2420 | 2430 | 2440 | 2450 |
| AGGAA AGAGA GGTCC TTCCG GAGTG CCAGG GTCCA AAGGA CACAG AGGTG | | | | |
| TCCTT TCTCT CCAGG AAGGC CTCAC GGTCC CAGGT TTCCT GTGTC TCCAC | | | | |
| 2460 | 2470 | 2480 | 2490 | 2500 |
| ATTCA GGACG TCCAG GGTTC GCAGG GCCAG CCGGC AAGCC TGGAT CCCCC | | | | |
| TAAGT CCTGC AGGTC CCAAA CGTCC CGGTC GGCCG TTCGG ACCTA GGGGT | | | | |
| 2510 | 2520 | 2530 | 2540 | 2550 |
| GGTCT CAAAG GTCCC AGAGG CAGAG AGGGA AGTGC TGGGT TTCAA GGTAT | | | | |
| CCAGA GTTTC CAGGG TCTCC GTCTC TCCCT TCACG ACCCA AAGTT CCATA | | | | |
| 2560 | 2570 | 2580 | 2590 | 2600 |
| CCCGG GTCCA CCTGG CCATT CCTGC AAAAG AGGCG CTCCA GGGAT ACCAG | | | | |
| GGGCC CAGGT GGACC GTTAA GGACG TTTTC TCCGC GAGGT CCCTA TGGTC | | | | |
| 2610 | 2620 | 2630 | 2640 | 2650 |
| GGCAA CCGGG GCTCC CTGGG GCCCC GGGGA GTCCA GGTGC CCCAG GTTGG | | | | |
| CCGTT GGCCC CGAGG GACCC CGGGG CCCCT CAGGT CCACG GGGTC CAACC | | | | |
| 2660 | 2670 | 2680 | 2690 | 2700 |
| AAAGG ACAGC GAGGG GATGT GGGGC CTCCG GGTCC AGCTG GAATG AAAGG | | | | |
| TTTCC TGTCG CTCCC CTACA CCCCC GAGGC CCAGG TCGAC CTTAC TTTCC | | | | |
| 2710 | 2720 | 2730 | 2740 | 2750 |
| CCTCC CTGGA GTCCC TGGAC GGCCA GGGGC AGAGG GACCC CTAGG ATTCC | | | | |
| GGAGG GACCT CAGGG ACCTG CCGGT CCCCC TCTCC CTGGG GATCC TAAGG | | | | |

| | | | | |
|--|------|------|------|------|
| 2760 | 2770 | 2780 | 2790 | 2800 |
| CAGGA GTCCC AGGCC CCTTA GGGGA TGATG GGCTG CCTGG TCTTC CAGGC | | | | |
| GTCCCT CAGGG TCCGG GGAAT CCCCT ACTAC CCGAC GGACC AGAAG GTCCG | | | | |
| 2810 | 2820 | 2830 | 2840 | 2850 |
| CCGAA GGGAC CCCAG GGGCT GCCTG GCTTC CCCGG TTTTC CAGGG GAGAG | | | | |
| GGCTT CCCTG GGGTC CCCGA CGGAC CGAAG GGGCC AAAAG GTCCC CTCTC | | | | |
| 2860 | 2870 | 2880 | 2890 | 2900 |
| AGGAA AGCCT GGCCC AGAGG GACGC CCTGG CAGCA AGGGG GAACC AGGAG | | | | |
| TCCTT TCGGA CCGGG TCTCC CTGCG GGACC GTCGT TCCCC CTTGG TCCTC | | | | |
| 2910 | 2920 | 2930 | 2940 | 2950 |
| AGGAT GGTCG GCCTG GCTTC TTCGG AGACC AAGGG GTGAA AGGTG CCAAA | | | | |
| TCCTA CCAGC CGGAC CGAAG AAGCC TCTGG TTCCC CACTT TCCAC GGTTC | | | | |
| 2960 | 2970 | 2980 | 2990 | 3000 |
| GGAGC GAGAG GACCC CCAGG AGATG AAGGA GAGAT GGCGG TCTTT TCCCA | | | | |
| CCTCG CTCTC CTGGG GTTCC TCTAC TTCCT CTCTA CCGCC AGAAA AGGGT | | | | |
| 3010 | 3020 | 3030 | 3040 | 3050 |
| AAAAG GGAAA ACCGG GGAAC CTGGA CCTCC AGGAG ATGAT GGATT CCCAG | | | | |
| TTTTC CCTTT TGGCC CCTTG GACCT GGAGG TCCTC TACTA CCTAA GGGTC | | | | |
| 3060 | 3070 | 3080 | 3090 | 3100 |
| GAGAA GAAGG TGATA AAGGC AATCC AGGGA TGCCG GGGAG GAGAG GAGAG | | | | |
| CTCTT CTTCC ACTAT TTCCG TTAGG TCCCT ACGGC CCCTC CTCTC CTCTC | | | | |
| 3110 | 3120 | 3130 | 3140 | 3150 |
| CCTGG AAGAT TCGGA GCACC TGGAT TTCAC AGAGG GGAGC CCGGG AGAAC | | | | |
| GGACC TTCTA AGCCT CGTGG ACCTA AAGTG TCTCC CCTCG GGCCC TCTTG | | | | |
| 3160 | 3170 | 3180 | 3190 | 3200 |
| CGGGC AGCCA GGGCT TCCTG GCCCC CCAGG CCCCC CAGGC TCACC TGGGC | | | | |
| GCCCC TCGGT CCCGA AGGAC CGGGG GGTCC GGGGG GTCCG AGTGG ACCCG | | | | |
| 3210 | 3220 | 3230 | 3240 | 3250 |
| TGAGA GGGAT TATTG GTTTT CCGGG ATTTT CAGGT GACCA GGGTG AGCCA | | | | |
| ACTCT CCCTA ATAAC CAAAA GGCCC TAAAG GTCCA CTGGT CCCAC TCGGT | | | | |
| 3260 | 3270 | 3280 | 3290 | 3300 |
| GGTTC TCCAG GGTCT CCTGG ACTTT CAGGA ATTGA TGGAA TGAGA GGACC | | | | |
| CCAAG AGGTC CCAGA GGACC TGAAA GTCCT TAACT ACCTT ACTCT CCTGG | | | | |
| 3310 | 3320 | 3330 | 3340 | 3350 |
| TAAAG GAAGC AAAGG TGACC CTGCA AGTCA GTTCG GCCCA CCTGG TCCAA | | | | |
| ATTTT CTTCG TTTCC ACTGG GACGT TCAGT CAAGC CGGGT GGACC AGGTT | | | | |
| 3360 | 3370 | 3380 | 3390 | 3400 |
| AGGGT GAACC AGGTA GCCCT GGATG TCCAG GACAA GTTGG AGCAC CTGGA | | | | |
| TCCCA CTTGG TCCAT CGGGA CCTAC AGGTC CTGTT CAACC TCGTG GACCT | | | | |
| 3410 | 3420 | 3430 | 3440 | 3450 |
| GAGCA GGGCT TGCCT GGTGT TCAAG GGCTC AGAGG ACCAC CTGGA AGGCC | | | | |
| CTCGT CCCGA ACGGA CCACA AGTTC CCGAG TCTCC TGGTG GACCT TCCGG | | | | |

| | | | | |
|--|------|------|------|------|
| 3460 | 3470 | 3480 | 3490 | 3500 |
| AGGAT TACCT GGCTC CTCCG GACCA CCAGG GTGTC CAGGT AATCA AGGGG | | | | |
| TCCTA ATGGA CCGAG GAGGC CTGGT GGTCC CACAG GTCCA TTAGT TCCCC | | | | |
| 3510 | 3520 | 3530 | 3540 | 3550 |
| TGCCT GGGCT GACAG GGCCT CCAGG AGAAA CAGGG GATCC TGGGC CAAGA | | | | |
| ACGGA CCCGA CTGTC CCGGA GGTCC TCTTT GTCCC CTAGG ACCCG GTTCT | | | | |
| 3560 | 3570 | 3580 | 3590 | 3600 |
| GGCAT GATGG GAGAT CCAGG GCCAC CAGGT CTTCC AGGAA TAAAA GGTCC | | | | |
| CCGTA CTACC CTCTA GGTCC CGGTG GTCCA GAAGG TCCTT ATTTT CCAGG | | | | |
| 3610 | 3620 | 3630 | 3640 | 3650 |
| CTCCG GGTCT CCGGG TCTGA ATGGC TTGCA TGGTT TAAAG GTTCA GAAAG | | | | |
| GAGGC CCAGC GGCCC AGACT TACCG AACGT ACCAA ATTTT CCAGT CTTTC | | | | |
| 3660 | 3670 | 3680 | 3690 | 3700 |
| GAGCC AAAGG TGCTT CAGGT TTGCA CGAAG TGGGC CCACC CGGTC CAGTG | | | | |
| CTCGG TTTCC ACGAA GTCCA AACGT GCTTC ACCCG GGTGG GCCAG GTCAC | | | | |
| 3710 | 3720 | 3730 | 3740 | 3750 |
| GGCGT ACCTG GGCTG AAAGG GGAGC CGGGA GACCC TGGGA GCCCA GGAAT | | | | |
| CCGCA TGGAC CCGAC TTTCC CCTCG GCCCT CTGGG ACCCT CGGGT CCTTA | | | | |
| 3760 | 3770 | 3780 | 3790 | 3800 |
| TTCTC CCCCA GGCCT TTCTG GAGAA AGAGG CCCCC CCGGC CCCCC AGGGA | | | | |
| AAGAG GGGGT CCGGA AAGAC CTCTT TCTCC GGGGG GGCCG GGGGG TCCCT | | | | |
| 3810 | 3820 | 3830 | 3840 | 3850 |
| GACCT GGAGC CCCTG GTCCT GCAGG TGCCA CAGGA AGAGC TGCTG AAGGT | | | | |
| CTGGA CCTCG GGGAC CAGGA CGTCC ACGGT GTCCT TCTCG ACGAC TTCCA | | | | |
| 3860 | 3870 | 3880 | 3890 | 3900 |
| CACGT TCCTG ACCCA GGTCC ACCCG GAGAT GTGGG TCCTC CTGGC CCCGA | | | | |
| GTGCA AGGAC TGGGT CCAGG TGGGC CTCTA CACCC AGGAG GACCG GGGCT | | | | |
| 3910 | 3920 | 3930 | 3940 | 3950 |
| TGGTC CGAGA GGAGC GCCCG GGCCC CCAGG CCCCC CTGGG AGCGT TGACC | | | | |
| ACCAG GCTCT CCTCG CGGGC CCGGG GGTCC GGGGG GACCC TCGCA ACTGG | | | | |
| 3960 | 3970 | 3980 | 3990 | 4000 |
| TTCTG AGAGG GGAAC CAGGA GACTG TGGGC CGCCG GGGCC TCCAG GTCCC | | | | |
| AAGAC TCTCC CCTTG GTCCT CTGAC ACCCG GCGGC CCCGG AGGTC CAGGG | | | | |
| 4010 | 4020 | 4030 | 4040 | 4050 |
| CCGGG CCCAC CCGGC CCTCC AGGAC ACCAA GGCTT CCCAG GATGC GATGG | | | | |
| GGCCC GGGTG GGCCG GGAGG TCCTG TGGTT CCGAA GGGTC CTACG CTACC | | | | |
| 4060 | 4070 | 4080 | 4090 | 4100 |
| AAAAG ACGGC CAGAA AGGAC CAATA GGATT CCCGG GGCTG CAGGG GCCTC | | | | |
| TTTTC TGCCG GTCTT TCCTG GTTAT CCTAA GGGCC CCGAC GTCCC CGGAG | | | | |
| 4110 | 4120 | 4130 | 4140 | 4150 |
| AGGGA CTTCC TGGGC TCCCT GGGGA GAAGG GTCTG CTGGG CATTC CAGGC | | | | |
| TCCCT GAAGG ACCCG AGGGA CCCCT CTTCC CAGAC GACCC GTAAAG GTCCC | | | | |

| | | | | |
|-------------------|--------------------|-------------------|--------|------|
| 4160 | 4170 | 4180 | 4190 | 4200 |
| CGGCG AGGGC ACCCC | GGTCC TCCAG GTTCC | AGAGG TGAAC CAGGG | CCTCC | |
| GCCGC TCCCG TGGGG | CCAGG AGGTC CAAGG | TCTCC ACTTG GTCCC | GGAGG | |
| 4210 | 4220 | 4230 | 4240 | 4250 |
| TGCAG ATGTG GATTG | GTGCC CCCGC ATCCC | CGGGC TTCCT GGGGT | ACCAG | |
| ACGTC TACAC CTAAG | CACGG GGGCG TAGGG | GCCCC AAGGA CCCCA | TGGTC | |
| 4260 | 4270 | 4280 | 4290 | 4300 |
| GCCCA AGAGG ACCAG | AAGGA GCCAT GGGGG | TCCCT GGAAG GAGAG | GGCCC | |
| CGGGT TCTCC TGGTC | TTCCCT CGGTA CCCCC | AGGGA CCTTC CTCTC | CCGGG | |
| 4310 | 4320 | 4330 | 4340 | 4350 |
| CCAGG ACCAG GGTGC | AAAGG AGAGC CGGGA | CTGGA GGGCA GGAGG | GGCGA | |
| GGTCC TGGTC CCACG | TTTCC TCTCG GCCCT | GACCT CCCGT CCTCC | CCGCT | |
| 4360 | 4370 | 4380 | 4390 | 4400 |
| GGCCG GCCTC CCAGG | ACCTC CGGGG CCTCC | CGGAA GCACA GGGGA | CGCAG | |
| CCGGC CGGAG GGTCC | TGGAG GCCCC GGAGG | GCCTT CGTGT CCCCT | GCGTC | |
| 4410 | 4420 | 4430 | 4440 | 4450 |
| GAGAA GCCGG TTGCC | CCGGA GCACC AGGGC | CCCCC GGGCC CATTG | GGGAC | |
| CTCTT CGGCC AACGG | GGCCT CGTGG TCCCG | GGGGG CCCGG GTAAC | CCCTG | |
| 4460 | 4470 | 4480 | 4490 | 4500 |
| CCCCG GCCCA GAGGG | TTTGG GCCTG GATAC | CTCAG TGGCT TCCTC | CTGGT | |
| GGGCC CGGGT CTCCC | AAACC CGGAC CTATG | GAGTC ACCGA AGGAG | GACCA | |
| 4510 | 4520 | 4530 | 4540 | 4550 |
| TCTCC ACAGT CAGAC | GGATG GGGAG CCCAC | CTGCC CCGCG GGCAT | GCCCCA | |
| AGAGG TGTCA GTCTG | CCTAC CCCTC GGGTG | GACGG GGCGC CCGTA | CGGGT | |
| 4560 | 4570 | 4580 | 4590 | 4600 |
| GGCCC TGGAC GGGCT | ACAGT CTGTT ATACC | TGGAA GGACA GGAGA | AGGCC | |
| CCGGG ACCTG CCCGA | TGTCA GACAA TATGG | ACCTT CCTGT CCTCT | TCCGG | |
| 4610 | 4620 | 4630 | 4640 | 4650 |
| CACAA TCAGG ACCTT | GGTCT GGCAG GGTCT | TGCCT TCCCA TGTTC | AGCAC | |
| GTGTT AGTCC TGGAA | CCAGA CCGTC CCAGA | ACGGA AGGGT ACAAG | TCGTG | |
| 4660 | 4670 | 4680 | 4690 | 4700 |
| CCTGC CCTTT GCCTA | CTGCA ACATC CACCA | AGTGT GCCAC TACGC | CCGGA | |
| GGACG GGAAA CGGAT | GACGT TGTAG GTGGT | TCACA CGGTG ATGCG | GGCCT | |
| 4710 | 4720 | 4730 | 4740 | 4750 |
| GAAAC GACCG GTCCT | ACTGG CTGGC CAGTG | CTGCG CCCCT GCCCA | TGATG | |
| CTTTG CTGGC CAGGA | TGACC GACCG GTCAC | GACGC GGGGA CGGGT | ACTAC | |
| 4760 | 4770 | 4780 | 4790 | 4800 |
| CCGCT CTCAG AGGAG | GAGAT CCGCC CCTAC | ATCAG CCGCT GTGCC | GTGTG | |
| GGCGA GAGTC TCCTC | CTCTA GGCGG GGATG | TAGTC GGCGA CACGG | CACAC | |
| 4810 | 4820 | 4830 | 4840 | 4850 |
| CGAGG CCCCC GCGGG | CGTGG TGGCG CTGCA | CAGCC AGGAC CGGTC | CATCC | |
| GCTCC GGGGT CGGGG | GCACC ACCGC GACGT | GTCGG TCCTG GCCAG | GTAGG | |

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      4860      4870      4880      4890      4900
CGCCG TGTCC ACGGA GCTGG AGGAG CCTCT GGATC GGGTA CTCGT TCCTG
GCGGC ACAGG TGCCT CGACC TCCTC GGAGA CCTAG CCCAT GAGCA AGGAC

      4910      4920      4930      4940      4950
ATGCA CACAG GGGCT GGAGA CCAAG GAGGA GGGCA GGCCC TCATG TCCCC
TACGT GTGTC CCCGA CCTCT GGTTC CTCCT CCCGT CCGGG AGTAC AGGGG

      4960      4970      4980      4990      5000
CGGCA GCTGT CTGGA AGATT TCCGA GCCGC ACCGT TCCTC GAATG CCAAG
GCCGT CGACA GACCT TCTAA AGGCT CGGCG TGGCA AGGAG CTTAC GGTTC

      5010      5020      5030      5040      5050
GCCGA CAGGG AACTT GCCAC TTTT TGC AA ATGAG TATAG CTTCT GGCTG
CGGCT GTCCC TTGAA CGGTG AAAAA ACGTT TACTC ATATC GAAGA CCGAC

      5060      5070      5080      5090      5100
ACGAC GGTGA GGCCT GACTT GCAGT TTTCC TCGGC ACCAT CACCA GACAC
TGCTG CCACT CCGGA CTGAA CGTCA AAAGG AGCCG TGGTA GTGGT CTGTG

      5110      5120      5130      5140      5150
CTTGA AAGAA AGCCA GGCCC AGCGC CAGAA AATCA GCAGG TGCCA GGTAT
GAACT TTCTT TCGGT CCGGG TCGCG GTCTT TTAGT CGTCC ACGGT CCATA

      5160
GCATA TGA CT AGT
CGTAT ACTGA TCA

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APPENDIX 5

Canine COL4A4 nucleotide alignment

Consensus key (see documentation for details)

- * - single, fully conserved residue
- no consensus

CLUSTAL W (1.81) multiple sequence alignment

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Canine_A4_Normal_5-21      CTAGAGGATCTACTAGTCATATGGATGTGAGAGTCTTCTGGAAGAGAAGA
Human_COL4A4               -----
Mouse_COL4A4               -----

Canine_A4_Normal_5-21      GAAACCTGACAATCTTTTTTAAGTCCCAAGACTTGACTGTGAGGTAATCC
Human_COL4A4               -----ATGTGGTCT
Mouse_COL4A4               -----

Canine_A4_Normal_5-21      ATGCACATGGCATCAGTAAAGTGTTCATTTCAGGTGGATCAAGCCCTTGGC
Human_COL4A4               CTGCACATAGTACTAATGAGGTGCTCCTTCAGATTGACCAAGTCCTTGGC
Mouse_COL4A4               -----ATGAGGTGCTTTTTCAGATGGACCAAGTCCTTGT
                        * * * * *

Canine_A4_Normal_5-21      CAGAGATCCCTGGTCACTTATATTTATCCTCTTTTCTGTACAACATGCAT
Human_COL4A4               CACAGGTCCCTGGTCACTTATACTCATTCTCTTTTCTGTACAATATGTAT
Mouse_COL4A4               CACAGCCCCCTGGTCACTAATATTTATCCTCTTTTACTATACAATATGAAT
                        ** * * * * *

Canine_A4_Normal_5-21      ATGGGAGTGGAAAGAAGTTTGTTCGGCCCCCTGTGGAGGAAGAGATTGCTCG
Human_COL4A4               ATGGGAGTGGAAAGAATAACATTGGTCCTTGTGGAGGAAGAGATTGCTCT
Mouse_COL4A4               ATGGGAGTGGCAAGAAGTATGGCGGCCCTGCGGAGGAAGAAACTGCTCT
                        *****

Canine_A4_Normal_5-21      GTGTGCCATTGCTTTCTGAAAAAGGGTCTCGGGGTCAACCAGGACCGCC
Human_COL4A4               GTTTGCCACTGTGTTCCTGAAAAGGGGTCTCGGGGTCCACCAGGACCC
Mouse_COL4A4               GTCTGCCAGTGTTCCTGAGAAGGGATCTCGGGGTCAACCAGGACCACT
                        ** * * * * *

Canine_A4_Normal_5-21      GGGGCCACAGGGTCCCATTGGACCCCTGGGACTGCCAGGACCCACAGGAA
Human_COL4A4               AGGGCCACAGGGTCCAATTGGACCCCTGGGAGCCCAGGACCCATTGGGC
Mouse_COL4A4               GGGACCACAGGGACCAATAGGGCCCTTGGGACCCCTGGGACCTATTGGGA
                        ** * * * * *

Canine_A4_Normal_5-21      TTCCAGGAGAGAAAAGGGATGAGAGGTGACAGTGGCCCTCCTGGAGCAGCA
Human_COL4A4               TTTCAGGAGAGAAAAGGAATGAGAGGGGACCGCGGCCCTCCTGGAGCAGCA
Mouse_COL4A4               TTCCAGGAGAGAAAAGGAGAGAGAGAGACAGTGGCTCACCTGGACCACCA
                        ** * * * * *

Canine_A4_Normal_5-21      GGTGACAAAGGTGACAAGGGTCCAACCTGGTGTTCCTGGATTTCAGGGTT
Human_COL4A4               GGGGACAAAGGAGATAAGGGTCCAACCTGGTGTTCCTGGATTTCAGGGTT
Mouse_COL4A4               GGAGAGAAAGGAGACAAGGGTCCAACCTGGTGTTCCTGGATTTCAGGTGT
                        ** * * * * *

Canine_A4_Normal_5-21      GGATGGCATACCTGGCCACCCGGGGCCTCCTGGATCCAGAGGCAAGCCTG
Human_COL4A4               AGATGGCATACCTGGGCACCCAGGGCCTCCTGGACCCAGAGGCAACCTG
Mouse_COL4A4               GGATGGTGTACCTGGTCACCCAGGGCCTCCTGGACCCAGAGGCAACCTG
                        *****

Canine_A4_Normal_5-21      GCATGCATGGCTACAATGGTTCACGAGGTGATCCAGGGTTTCAGGAAAA
Human_COL4A4               GTATGAGTGGCCACAATGGCTCAAGAGGTGACCCAGGGTTTCAGGAGGA
Mouse_COL4A4               GTGTGGACGCTACAATGGCTCACGTGGTATCCGGGCTATCCAGGAGAA
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| | |
|-----------------------|--|
| Canine_A4_Normal_5-21 | AGAGGAGTTCCTGGCCCAGGAGGCCCCCAGGCCTTCCTGGGGAAGGGG |
| Human_COL4A4 | AGAGGAGCTCTTGGCCCAGGAGGCCCCCTAGGCCATCCTGGGGAAGGGG |
| Mouse_COL4A4 | AGAGGAGCTCCAGGCCAGGCGGCCCCCTGGTCAGCCTGGGGAATATG |
| | ***** ** ***** ***** ** * ***** ** |
| Canine_A4_Normal_5-21 | AGAAAAAGGAAATTCAGTGTTCATTTTAGGTGCCATTAAAGGTATTCAGG |
| Human_COL4A4 | AGAAAAAGGAAATTCAGTGTTCATTTTAGGTGCCGTTAAAGGTATTCAGG |
| Mouse_COL4A4 | AGAAAAAGGAAGATCTGTGTACATTACTGGTGGCGTTAAAGGTATTCAGG |
| | ***** ** * ** * ** * ** * ** * ***** |
| Canine_A4_Normal_5-21 | GAGACAGGGGGACCCAGACCTGCCTGGCTTGCCGGGACCGAAGGGGGCA |
| Human_COL4A4 | GAGACAGAGGGGACCCAGGACTGCCTGGCTTACCAGGATCTTGGGGTGCA |
| Mouse_COL4A4 | GAGACCGTGGGGACCCAGGACCCCGGCTTACCAGGATCTAGGGGTGCA |
| | ***** * ***** * ** * ** * ** * ** * ** * ** * |
| Canine_A4_Normal_5-21 | GGAGGACCGGCAGGTCCCAGGGACAGCCAGGAGAGCCTGGGTAAACAGG |
| Human_COL4A4 | GGAGGACCGGCAGGTCCCACAGGATATCCTGGAGAGCCAGGGTTAGTGGG |
| Mouse_COL4A4 | CAAGGGTCACCAGGGCCCATGGGGCATGCAGGCGCACCAGGGCTAGCAGG |
| | *** * ***** ** * * * * * ** * ** * ** * |
| Canine_A4_Normal_5-21 | TGCTCCGGGCCATCCTGGGAGACCAGGCTTGAAGGGTAATCCTGGTGTGG |
| Human_COL4A4 | ACCTCCGGGCCAACCCAGGGCGTCCAGGTTTGAAGGGAATCCCGGTGTGG |
| Mouse_COL4A4 | ACCTATAGGTCATCCTGGAAGCCCTGGGTGAAGGGAATCCTGCCACTG |
| | ** * * * * ** * * * * * ** * * * * * ** * * |
| Canine_A4_Normal_5-21 | GAGTAAAGGGGCAATGGGAGACCCGGGTGAGGTTGGCCAGCAGGGTTCT |
| Human_COL4A4 | GAGTAAAGGGGCAATGGGAGACCCGGGTGAGGTTGGTCAGCAAGGTTCT |
| Mouse_COL4A4 | GACTGAAGGACAAGGGGAGAGCCGGGTGAAGTTGGCCAGCGTGGTCTCT |
| | ** * ***** ** * * * * * ** * * * * * ** * ** |
| Canine_A4_Normal_5-21 | CCTGGACCCACCTTATTGGTACAGCCACCTGATTCCTGTCTGTATAAAGG |
| Human_COL4A4 | CCTGGACCCACCTTGTGGTAGAGCCACCTGACTTTTGTCTCTATAAAGG |
| Mouse_COL4A4 | CCTGGGCCACGCTGTGGTACAGCCGCTGATTTGAGTATCTATAAAGG |
| | ***** ** * * * * * ** * * * * * ** * * * * * ** * |
| Canine_A4_Normal_5-21 | AGAAAAGGGCATAAAAGGAATGCCTGGTATGATTGGACCTCCAGGACTAC |
| Human_COL4A4 | AGAAAAGGGTATAAAAGGAATTCCTGGAATGGTTGGACTGCCAGGACCAC |
| Mouse_COL4A4 | AGAAAAGGGTGTAAAAGGAATGCCTGGAATGATTGGTCCCCCAGGACCTC |
| | ***** ***** ***** ** * * * * * ** * * * * * |
| Canine_A4_Normal_5-21 | CGGGACCCAAGGGAGAACCTGGAATTGGACCAAAGGAGAGAAAGGGTATT |
| Human_COL4A4 | CAGGACGCAAGGGAGAAATCTGGTATGGGGCAAAAGGAGAAAAAGGTATT |
| Mouse_COL4A4 | CAGGTGCAAGGGAGCGCCTGGTGTGGGATCAAGGAGAGAAAGGTATC |
| | * * * * * ***** ** * * * * * ***** ** * * * * * |
| Canine_A4_Normal_5-21 | CCTGGGTTCTCAGGACCTCGGGGTGATCCGGGTTCTATGGCTCTCCAGG |
| Human_COL4A4 | CCTGGATTTCAGGGCCTCGGGGGATCCTGGTTCTATGGATCTCCAGG |
| Mouse_COL4A4 | CCTGGGTTCCAGGACCCCGGGGTGAGCCTGGTTCGCATGGGCCTCCAGG |
| | ***** ** * * * * * ** * * * * * ** * * * * * ***** |
| Canine_A4_Normal_5-21 | TTTTCCAGGATTAAAGGGGAAACCAGGACTGTTTGGAGATCCTGGATCAT |
| Human_COL4A4 | TTTTCCAGGATTAAAGGGAGAACTAGGACTGGTTGGAGATCCTGGGCTAT |
| Mouse_COL4A4 | TTTTCCAGGATTCAGGGGATACAGGAGCAGCTGGAGAGCCTGGGCTGT |
| | ***** ***** ** * * * * * ***** ***** * |
| Canine_A4_Normal_5-21 | TTGGATTTGTGGCCCAAAGGGGGATCCTGGAGACCGTGGGTACCCAGGA |
| Human_COL4A4 | TTGGATTAATTGGCCCAAAGGGGGATCCTGGAATCGAGGGCACCAGGA |
| Mouse_COL4A4 | TCGGATTTCTGGTCCAAAGGGGGATCTGGGAGATCGGGGTACCCGGGA |
| | * * * * * ***** ***** ** * * * * * ** * * * * * |
| Canine_A4_Normal_5-21 | CCACCGGGTGTCTGGTAACTCCATCTGTTCCTCAAAAGGCCCTCCAGG |
| Human_COL4A4 | CCACCGAGTGTCTTGGTGACTCCACCTCTTCCACTTAAAGGCCACCAGG |
| Mouse_COL4A4 | CCACCGGATTTTGTGACTCCAGTCCACCTCAAAAGGTGTTCCAGG |
| | ***** ** * * * * * ***** ** * * * * * ***** |

| | |
|-----------------------|--|
| Canine_A4_Normal_5-21 | GGATCCGGGGCGCCCTGGCCACTATGGAGAAATGGGGTCCGTGGACCAC |
| Human_COL4A4 | GGACCCAGGGTTCCTTGCCCGCTATGGAGAAACAGGGGATGTTGGACCAC |
| Mouse_COL4A4 | GGACCCAGGACCCCTGGCTACTATGGGGAGATTGGAGATGTTGGATTAC |
| | *** ** * ***** ** * * * ***** ** |
| Canine_A4_Normal_5-21 | CTGGTCCCCCTGGTCCCCAGGTCCACCAGGGGAAGACTGTGCAGGCATG |
| Human_COL4A4 | CTGGTCCCCAGGTCTCTTGGGCAGACCAGGGGAAGCCTGTGCAGGCATG |
| Mouse_COL4A4 | CAGGTCTCCAGGGCCCCCAGGCAGACCAGGGGAACTTGTCCAGGCATG |
| | * ***** ** * * * ** ***** ** ***** |
| Canine_A4_Normal_5-21 | ATGGGACCCCTGGACCAAGAGGATTTCTTGGTTCATCCAGGATTTCCAGG |
| Human_COL4A4 | ATAGGACCCCTGGGCCACAAGGATTTCTTGGTCTTCTGGGCTTCCAGG |
| Mouse_COL4A4 | ATGGGACCTCCTGGACCACCAGGGGTTCTTGGTCTCCAGGATTTCCAGG |
| | ** ***** ** * * * ** ***** ** * ***** |
| Canine_A4_Normal_5-21 | GGCAGCTGGTATCCAGGGAGAGCGGATTAAAGTCCAGGAAAACAGGGA |
| Human_COL4A4 | AGAAGCTGGTATTCTTGGGAGACCTGATTCTGCTCCAGGAAAACAGGGA |
| Mouse_COL4A4 | AGAAGCTGGGGTTCTTGAAGACTCGACTGTGCCCCAGGAAAACCTGGGA |
| | * ***** * ** * * ** ** * ***** ** * |
| Canine_A4_Normal_5-21 | AACCGGGACCTCCTGGGTGCCCCGAGCACCAGGGCTGCAAGGACCCCCA |
| Human_COL4A4 | AGCCAGGATCACCTGGCTTGCCCTGGAGCACCAGGCCTGCAGGGCCTCCCA |
| Mouse_COL4A4 | AGCCAGGGCTACCCGGCTGCCTGGCGCACCAGGACCAGGGGGCCCCCT |
| | * ** * * ** * * * * ** * * * * * * * * * * * * * * * |
| Canine_A4_Normal_5-21 | GGATCAGATGTTATCTATTGTAGTGTGGGCATCCTGGACCACAAGGGAT |
| Human_COL4A4 | GGATCAAGTGTGATATACTGTAGTGTGGGAACCCCGACCACAAGGAAT |
| Mouse_COL4A4 | GGATCAGATGTGATATACTGTGCGCCTGGGTGCCCTGGGCCAATGGGTGA |
| | ***** ** * * * * * * * * * * * * * * * * * |
| Canine_A4_Normal_5-21 | AAAAGGCAAAATGGGTCTCCAGGAAGAAGAGGCTCGAAAGGAGAAAAAG |
| Human_COL4A4 | AAAAGGCAAAAGTTGGTCCCCCAGGAGGAAGAGGCCAAAAGGAGAAAAAG |
| Mouse_COL4A4 | AAAGGAAAAAGTGGGTCTCCAGGAAGAAGAGGAGCCAAAGGAGCAAAAG |
| | *** ** * * * * * * * * * * * * * * * * * |
| Canine_A4_Normal_5-21 | GAAATGCGGGGCTCTGTGCCTGTGAGCCTGGTCCCATGGGCCCACCAGGC |
| Human_COL4A4 | GAAATGAAGGACTCTGTGCCTGTGAGCCTGGACCCATGGGCCCCCTGGC |
| Mouse_COL4A4 | GAAACAAGGACTCTGTACCTGTCCACCTGGTCCCATGGGACCCCTGGT |
| | **** ** ***** ***** ***** ***** ** * * |
| Canine_A4_Normal_5-21 | CCTCCAGGACTTCTTGAAGGCAGGGTAGTAAGGGAGACTTGGGGCTCCC |
| Human_COL4A4 | CCTCAGGACTTCTTGGGAGGCAGGGGAGTAAGGGAGACTTGGGGCTCCC |
| Mouse_COL4A4 | CCCCCAGGACCTCTTGAAGACAAGGAAGTAAAGGAGACTTAGGGCTCCC |
| | ** ***** ***** * * * * * ***** ***** ***** |
| Canine_A4_Normal_5-21 | TGGGTGGATGGAGAGAAAGGTCACCCAGGCCCTCCTGGTGTCTGAAGGAT |
| Human_COL4A4 | TGGCTGGCTTGGAAACAAAAGGTGACCCAGGACCTCCTGGTGTCTGAAGGAC |
| Mouse_COL4A4 | TGGTTGGCATGGAGAAAAGGGCGATCCAGGTCAGCCTGGTGTCTGAAGGAC |
| | *** ** * * * * * * * * * * * * * * * * * |
| Canine_A4_Normal_5-21 | CTCCAGGATCACCAGGAAAACATGGCGCCTCAGGACCACCTGGCAGCAAA |
| Human_COL4A4 | CTCCAGGGCTACCAGGAAAGCATGGTGCCTCTGGACCACCTGGCAACAAA |
| Mouse_COL4A4 | CGCCAGGGCCACCAGGAAGACCTGGTGCCATGGGGCCCCCTGGTCACAAA |
| | * ***** ***** * * * * * ** * * * * * |
| Canine_A4_Normal_5-21 | GGAGAAAAGGGCGACAGGGTTGTATCAAGAGTGAAAGGGAAAAAGGAGA |
| Human_COL4A4 | GGGGCGAAGGGTGACATGGTTGTATCAAGAGTTAAAGGGCACAAAGGAGA |
| Mouse_COL4A4 | GGGAAAAGGGTGACATGGTCATATCAAGAGTGAAAGGGCAAAAGGAGA |
| | ** * ***** ** * * * ***** ***** * ***** |
| Canine_A4_Normal_5-21 | AAGAGGTCCTGATGGGCTCCAGGATTTCCAGGACAACAGGGACAAAATG |
| Human_COL4A4 | AAGAGGTCCTGATGGGCCCCAGGATTTCCAGGGCAGCCAGGATCACATG |
| Mouse_COL4A4 | AAGAGGACTGGATGGGCCACCAGGATTTCCGGGGCCACATGGACAAGATG |
| | ***** * ***** ***** ***** ** * * * * * |

| | |
|-----------------------|---|
| Canine_A4_Normal_5-21 | GGTCTCAAAGGTCCCAGAGGCAGAGAGGGAAGTGCTGGGTTTCAAGGTAT |
| Human_COL4A4 | GGTCTCAAAGGTCCCAGAGGCAGAGAGGACATGCTGGGTTTCCAGGTGT |
| Mouse_COL4A4 | GGTCTCAAAGGTCCCAGAGGTAGAGAGGGAAGTCGTGGCTTTCCAGGGAT |
| | ***** * * * * * |
| Canine_A4_Normal_5-21 | CCCGGGTCCACCTGGCCATTCTGTGAAAAGAGGCGCTCCAGGGATACCAG |
| Human_COL4A4 | CCCAGGTCCACCTGGCCATTCTGTGAAAAGAGGTGCTCCAGGGATACCAG |
| Mouse_COL4A4 | CCCAGGATCCCCTGGCCATTCTGTGAAAAGAGGTGCTCCCGGGATCCCAG |
| | *** * * * * * |
| Canine_A4_Normal_5-21 | GGCAACCGGGGCTCCCTGGGGCCCCGGGAGTCCAGGTGCCCCAGGTTGG |
| Human_COL4A4 | GGCAACCGGGACTCCCTGGGTATCCAGGTAGCCCAGGTGCTCCAGGTGGG |
| Mouse_COL4A4 | GACAACCGGGACTTCTTGGAATCCAGGTGATCCAGGGGCCCCAGGTTGG |
| | * * * * * * |
| Canine_A4_Normal_5-21 | AAAGGACAGCGAGGGGATGTGGGGCCTCCGGGTCCAGCTGGAATGAAAGG |
| Human_COL4A4 | AAAGGACAGCCGGGAGATGTGGGGCCTCCCGGCCAGCTGGAATGAAAGG |
| Mouse_COL4A4 | AAAGGCCAGCCAGGAGACATGGGTCCCTCTGGACCAGCTGGGATGAAGGG |
| | ***** * * * * * |
| Canine_A4_Normal_5-21 | CCTCCCTGGAGTCCCTGGACGGCCAGGGGCAGAGGACCCCTAGGATTCC |
| Human_COL4A4 | CCTCCCCGGACTCCCAGGACGGCCTGGGGGCACATGGTCCCCCAGGCCTCC |
| Mouse_COL4A4 | CCTCCCTGGTCTCCAGGCCTACCAGGAGCTGATGGTCTCCGAGGGCCTC |
| | ***** * * * * * |
| Canine_A4_Normal_5-21 | CAGGAGTCCCAGGCCCTTAGGGGATGATGGGCTGCCTGGTCTTCCAGGC |
| Human_COL4A4 | CAGGAATCCCAGGTCCCTTTGGAGATGATGGGCTACCTGGTCTTCCAGGT |
| Mouse_COL4A4 | CTGGGATTCCAGGCCCAATGGGGAAGATGGTCTACCTGGTCTTCCAGGC |
| | * * * * * * |
| Canine_A4_Normal_5-21 | CCGAAGGGACCCAGGGGCTGCCTGGCTTCCCCGTTTTTCCAGGGGAGAG |
| Human_COL4A4 | CCAAAGGGACCCCGGGGGCTGCCCTGGTTTTCCAGGTTTTCCCGAGAAAAG |
| Mouse_COL4A4 | CTGAAGGGACTCCCAGGGCTGCCTGGCTTCCCTGGTTTTTCCAGGAGAGAG |
| | * * * * * * |
| Canine_A4_Normal_5-21 | AGGAAAGCCTGGCCCAGAGGGACGCCCTGGCAGCAAGGGGAACCAAGGAG |
| Human_COL4A4 | AGGAAAGCCTGGTGCAGAGGGATGTCCTGGCGCAAAGGGAGAACCTGGAG |
| Mouse_COL4A4 | AGGAAAGCCAGGGCCAGATGGAGAACCTGGAAGAAAGGGAGAGGTTGGAG |
| | ***** * * * * * |
| Canine_A4_Normal_5-21 | AGGATGGTCGGCCTGGCTTCTTCGGAGACCAAGGGGTGAAAGGTGCCAAA |
| Human_COL4A4 | AGAAGGGCATGTCTGCCCTTCTTGAGACCGGGGACTGAGAGGGGCCAAA |
| Mouse_COL4A4 | AGAAGGGCTGGCTTGGCTTGAAGGGAGACCTGGGAGAGCGAGGCGCCAAA |
| | ** * * * * * |
| Canine_A4_Normal_5-21 | GGAGCGAGAGGACCCCAAGGAGATGAAGGAGAGATGGCGGTCTTTTCCCA |
| Human_COL4A4 | GGAGCCATAGGACCTCCCGGAGATGAAGGAGAAATGGCTATCATTTTCCCA |
| Mouse_COL4A4 | GGAGACAGAGGACTTCTTGGTGATGCAGGAGAA--GCAGTCACTTCCAG |
| | **** * * * * * |
| Canine_A4_Normal_5-21 | AAAAGGGAAAACCGGGGAACCTGGACCTCCAGGAGATGATGGATTCCCAG |
| Human_COL4A4 | AAAGGGAACACCTGGGGAACTGGACCTCTGGAGATGATGGATTCCCAG |
| Mouse_COL4A4 | AAAGGGTGAACCTGGGGATGCTGGGCCTCCAGGAGATGGTGGGTTCTCAG |
| | *** * * * * * |
| Canine_A4_Normal_5-21 | GAGAAGAAGGTGATAAAGGCAATCCAGGGATGCCGGGAGGAGAGGAGAG |
| Human_COL4A4 | GAGAAAGAGGTGATAAAGGAACTCCCGGGATGCAAGGGAGAAGAGGAGAG |
| Mouse_COL4A4 | GAGAGAGAGGTGATAAAGGAAGCTCAGGGATGCGAGGGGAAGAGGGGAT |
| | **** * * * * * |
| Canine_A4_Normal_5-21 | CCTGGAAGATTGGGACACCTGGATTTCACAGAGGGGAGCCGGGAGAAC |
| Human_COL4A4 | CTGGGAAGATACGGACCCTGGATTTCACAGAGGGGAACCTGGTGAGAA |
| Mouse_COL4A4 | CCCGGAAGAGATGGACTACCGGGACTCCACAGAGGGCAGCCTGGGATAGA |
| | * * * * * * |

| | |
|-----------------------|---|
| Canine_A4_Normal_5-21 | CGGGCAGCCAGGGCTTCCTGGCCCCCAGGCCCCCAGGCTCACCTGGGC |
| Human_COL4A4 | AGGTGAGCCAGGGCTCCTGGACCCCCAGGCCCCCAGGCTCAACTGGTC |
| Mouse_COL4A4 | TGGGCTCCTGGGCTCCTGGGCCCCCTGGCCCTCAGGATCACCTGGAC |
| | *** ** |
| Canine_A4_Normal_5-21 | TGAGAGGGATTATTGGTTTTCCGGGATTTCAGGTGACCAGGGTGAGCCA |
| Human_COL4A4 | TAAGAGGGTTCATTGGTTTTCCAGGACTTCAGGTGACCAGGGTGAGCCA |
| Mouse_COL4A4 | TGAGAGGAGTCATTGGTTTTCCAGGATTTCAGGTGACCAGGGTGATCCA |
| | * **** |
| Canine_A4_Normal_5-21 | GGTTCTCCAGGTCTCCTGGACTTTCAGGAATTGATGGAATGAGAGGACC |
| Human_COL4A4 | GGTTCTCCAGGTCCCCCTGGATTTCAGGAATTGATGGAGCAAGAGGACC |
| Mouse_COL4A4 | GGTTCTCCAGGACCCCTGGATTCCAGGAGATGATGGAGCAAGAGGACC |
| | ***** * **** * **** |
| Canine_A4_Normal_5-21 | TAAAGGAAGCAAAGGTGACCCTGCAAGTCAGTTCGGCCCCACCTGGTCCAA |
| Human_COL4A4 | TAAAGGAAACAAAGGTGACCCTGCCAGTCACCTTTGGTCCACCTGGTCCAA |
| Mouse_COL4A4 | TAAAGGATACAAAGGTGACCCTGCCAGTCAGTGTGGTCCACCTGGTCCAA |
| | ***** **** * ** |
| Canine_A4_Normal_5-21 | AGGGTGAACCAGGTAGCCCTGGATGTCCAGGACAAGTTGGAGCACCTGGA |
| Human_COL4A4 | AGGGTGAGCCAGGTAGCCCTGGATGTCCAGGGCATTTTGGAGCATCCGGA |
| Mouse_COL4A4 | AGGGTGAGCCAGGTAGTCTCTGGATACCAAGGACGTACTGGAGTCCCCGGA |
| | ***** **** * **** |
| Canine_A4_Normal_5-21 | GAGCAGGGCTTGCCTGGTGTCAAGGGCTCAGAGGACCACCTGGAAGGCC |
| Human_COL4A4 | GAGCAGGGCTTGCCTGGTATTCAAGGGCCAGAGGATCACCTGGAAGGCC |
| Mouse_COL4A4 | GAGAAAGGCTTTCCTGGAGATGAAGGACCCCGAGGACCACAGGAGACC |
| | *** * **** * **** |
| Canine_A4_Normal_5-21 | AGGATTACCTGGCTCCTCCGGACCACCAGGGTGTCCAGGTAATCAAGGGG |
| Human_COL4A4 | AGGGCCACCTGGCTCCTCTGGACCACCAGGGTGCCCAGGTGATCACGGGA |
| Mouse_COL4A4 | TGGACAGCCTGGATCTTTTGGACCACCAGGGTGTCCAGGTGACCCAGGGA |
| | ** **** * **** |
| Canine_A4_Normal_5-21 | TGCCTGGGCTGACAGGGCCTCCAGGAGAAACAGGGGATCCTGGGCCAAGA |
| Human_COL4A4 | TGCCTGGGCTGAGGGGACAGCCAGGAGAAATGGGAGACCTGGGCCAAGA |
| Mouse_COL4A4 | TGCCCGGACTCAAGGGACATCCAGGAGAAAGTGGGAGACCTGGGCCAAGA |
| | **** ** * **** |
| Canine_A4_Normal_5-21 | GGCATGATGGGAGATCCAGGGCCACCAGGTCTTCCAGGAATAAAAGGTCC |
| Human_COL4A4 | GGCCTCCAGGGGATCCAGGGATACCAGGTCTTCCGGGAATAAAAGGTCC |
| Mouse_COL4A4 | GGTGATGCGGGAGATTTCCGGGAGGCCAGGTCTGCTGGAGTGAAGGGCC |
| | ** *** |
| Canine_A4_Normal_5-21 | CTCCGGGTCGCCGGTCTGAATGGCTTGCATGGTTTAAAGGGTCAGAAAG |
| Human_COL4A4 | CTCCGGATCACCTGGCCTGAACGGCTTGCATGGATTGAAAGGTGAGAAAG |
| Mouse_COL4A4 | TCTCGGGTCTCCTGGTCTGAATGGCTTACATGGTCTGAAGGGTGAGAAAG |
| | *** ** |
| Canine_A4_Normal_5-21 | GAGCCAAAGGTGCTTCAGGTTTGACAGAAAGTGGGCCACCCGGTCCAGTG |
| Human_COL4A4 | GAACTAAAGGTGCTTCAGGTTTGATGATGTGGGGCCACCTGGTCCAGTG |
| Mouse_COL4A4 | GAACAAAAGGCGCTCAGGTTTGCTCGAAATGGGTCCACTGGGCCAATG |
| | ** * **** |
| Canine_A4_Normal_5-21 | GGCGTACCTGGGCTGAAAGGGAGCCGGGAGACCTGGGAGCCCAGGAAT |
| Human_COL4A4 | GGAATACCTGGGCTAAAAGGGGAGAGAGGAGACCTGGGAGCCCAGGAAT |
| Mouse_COL4A4 | GGGATGCCGAGGCAAAAAGGAGAGAAAGGAGACCTGGGAGTCCAGGGAT |
| | ** * **** |
| Canine_A4_Normal_5-21 | TTCTCCCCAGGCCTTCTGGAGAAAGAGGCCCCCGGCCCCCAGGGA |
| Human_COL4A4 | CTCTCCTCCAGGTCTCGTGGAAAGAAAGGTCCCCAGGACCCCCAGGGA |
| Mouse_COL4A4 | TTCTCCTCCAGGTCTTCTGGAGAAAGGGCTTCCAGGACCCCGAGGGA |
| | ***** |

| | |
|---|---|
| Canine_A4_Normal_5-21
Human_COL4A4
Mouse_COL4A4 | GACCTGGAGCCCCTGGTCTGCAGGTGCCACAGGAAGAGCTGCTGAAGGT
GTTCAGGACCACCTGGTCTGCAGGTGCCACAGGAAGAGCTCCTAAGG--
GACCAGGACCACCTGGTCCCGCAGGTGCCACAGGAAGAGCTGCTAAAGGT
* * * * * |
| Canine_A4_Normal_5-21
Human_COL4A4
Mouse_COL4A4 | CACGTTCCTGACCCAGGTCCACCCGAGATGTGGGTCTCCTGGCCCCGA
-ACATTCCTGACCCGGGTCCACCTGGAGATCAGGGACCTCCTGGTCTGA
GACATTCCTGATCCAGGTCCACCTGGAGACCGGGGACCTCCTGGCCCCGA
* * * * * |
| Canine_A4_Normal_5-21
Human_COL4A4
Mouse_COL4A4 | TGGTCCGAGAGGAGCGCCCGGGCCCCAGGCCCTGGGAGCGTTGACC
TGGCCCAAGAGGAGCACCTGGGCCTCCAGGCCTCCTGGGAGTGTGACC
TGGCCCAAGAGGAGTACCTGGGCCTCCGGTTCCTGGGAATGTGACC
* * * * * |
| Canine_A4_Normal_5-21
Human_COL4A4
Mouse_COL4A4 | TTCTGAGAGGGGAACAGGAGACTGTGGGCCCGGGGCCTCCAGGTCCC
TTCTGAGAGGGGAGCCAGGTGACTGTGGTCTACCAGGGCCACAGGTCCC
TTCTGAAAGGGGACCCAGGTGACTGTGGTTGCCAGGACCACCTGGCTCC
* * * * * |
| Canine_A4_Normal_5-21
Human_COL4A4
Mouse_COL4A4 | CCGGGCCCCACCCGGCCTCCAGGACACCAAGGCTTCCAGGATGCGATGG
CCTGGCCCCACAGGCCTCCAGGATACAAGGCTTCCAGGATGTGATGG
CGAGGCCACAGGCCTCCAGGGTGTGAGGGTCCCCAGGATGTGATGG
* * * * * |
| Canine_A4_Normal_5-21
Human_COL4A4
Mouse_COL4A4 | AAAAGACGGCCAGAAAGGACCAATAGGATTCCTGGGGCTGCAGGGGCCTC
AAAAGATGGCCAGAAAGGACCAATGGGATTCCTGGGACCGCAGGGACCAC
CAAAGATGGCCAGAAAGGACCAATGGGACTTCCAGGGTTGCCAGGGCCAC
* * * * * |
| Canine_A4_Normal_5-21
Human_COL4A4
Mouse_COL4A4 | AGGGACTTCCTGGGCTCCCTGGGGAGAAGGGTCTGCTGGGCATTCCAGGC
ATGGATTTCCTGGGCCACCTGGAGAGAAGGGTTTACCTGGACCTCCAGGG
CTGGTCTTCCTGGGGCACCTGGCGAGAAGGGATTACCTGGCCCTCCAGGC
* * * * * |
| Canine_A4_Normal_5-21
Human_COL4A4
Mouse_COL4A4 | CGGCGAGGGCACCCCGTCTCCAGGTTCAGAGGTGAACAGGGCCTCC
AGAAAAGGGCCCACTGGTCTTCCAGGTCCAGAGGTGAACCGGGGCCACC
AGAAAAGGGCCAGTAGGTCTCCAGGCTGCAGAGGTGAACCTGGGCCTCC
* * * * * |
| Canine_A4_Normal_5-21
Human_COL4A4
Mouse_COL4A4 | TGCAGATGTGGATTCTGTGCCCCGCATCCCCGGGCTTCTGGGGTACCAG
TGCAGATGTGGATGACTGTCCCGAATCCAGGCCTTCTGGGGCGCCAG
TGCAGACGTGGATTCTGTCTCGAATCCAGGACTTCTGGAGTGCCAG
* * * * * |
| Canine_A4_Normal_5-21
Human_COL4A4
Mouse_COL4A4 | GCCCAAGAGGACCAGAAGGAGCCATGGGGGTCCCTGGAAGGAGAGGGCCC
GCATGAGAGGACCAGAAGGAGCCATGGGGCTCCCTGGAATGAGAGGCCCC
GTCCCAAGGAGACCAGAAGGAGCTATGGGGAACTGGACGGAGAGGCCTG
* * * * * |
| Canine_A4_Normal_5-21
Human_COL4A4
Mouse_COL4A4 | CCAGGACCAGGGTGCAAAGGAGAGCCGGGACTGGAGGGCAGGAGGGCGA
CCAGGACCAGGGTGCAAAGGAGAGCCTGGGCTGGATGGCAGGAGGGGTGT
CCTGGACCAGGGTGCAAAGGAGAGCCTGGACCGGATGGCAGACGGGGCCA
* * * * * |
| Canine_A4_Normal_5-21
Human_COL4A4
Mouse_COL4A4 | GGCCGGCCTCCAGGACCTCCGGGGCTCCCGGAAGCACAGGGGACGCGAG
GGATGGCGTCCCTGGGTCTCTGGGCCTCCCGGACGTAAGGTGACACAG
GGATGGCATCCCTGGATCTCCCGGGCTCCTGGACGCAAAGGTGACACTG
* * * * * |
| Canine_A4_Normal_5-21
Human_COL4A4
Mouse_COL4A4 | GAGAAGCCGGTTGCCCGGAGCACCAGGGCCCCCGGGCCATTGGGGAC
GAGAAGACGGCTACCCTGGAGGACCAGGGCCTCCTGGTCCATTGGGGAT
GAGAGGCAGGGTGTCTGGAGCACCAGGCCCTCCTGGTCCAAGGTGTGAT
* * * * * |

| | |
|-----------------------|--|
| Canine_A4_Normal_5-21 | CCCGGGCCCAAGGGGTTTGGGCCTGGATACCTCAGTGGCTTCCTCCTGGT |
| Human_COL4A4 | CCTGGGCCCCAAGGGGTTTGGCCCTGGATACCTCGGTGGCTTCCTCCTGGT |
| Mouse_COL4A4 | CCTGGGCCCCAAGGGGTTTGGCCCTGGATCGCTCAGTGGCTTCCTCCTGGT |
| | *** ***** |
| Canine_A4_Normal_5-21 | TCTCCACAGTCAGACGGATGGGGAGCCACCTGCCCCGCGGGCATGCCCA |
| Human_COL4A4 | TCTCCACAGTCAGACGGACAGGAGCCACCTGCCCCCTGGGCATGCCCA |
| Mouse_COL4A4 | TCTCCACAGTCAGACTGACCAGGAACCGGCCTGCCCCGTGGGCATGCCTC |
| | ***** ** |
| Canine_A4_Normal_5-21 | GGCCCTGGACGGGTACAGTCTGTTATACCTGGAAGGACAGGAGAAGGCC |
| Human_COL4A4 | GGCTCTGGACTGGGTATAGTCTGTTATACCTGGAAGGGCAAGAGAAAGCT |
| Mouse_COL4A4 | GGCTTTGGACAGGTACAGCTTGTTGTACATGGAAGGACAGGAGAAGGCG |
| | *** ***** |
| Canine_A4_Normal_5-21 | CACAATCAGGACCTTGGTCTGGCAGGGTCTTGCCTTCCCATGTTTACGAC |
| Human_COL4A4 | CACAATCAAGACCTTGGTCTGGCAGGGTCTTGCCTTCCCGTATTTAGCAC |
| Mouse_COL4A4 | CACAATCAAGATCTTGGTTTGGCAGGCTCTGTCTTCTGTGTTTAGCAC |
| | ***** ** |
| Canine_A4_Normal_5-21 | CCTGCCCTTTGCCTACTGCAACATCCACCAAGTGTGCCACTACGCCCGGA |
| Human_COL4A4 | GCTGCCCTTTGCCTACTGCAACATCCACCAGGTGTGCCACTATGCCCAGA |
| Mouse_COL4A4 | TCTGCCCTTTGCCTACTGCAACATCCACCAAGTGTGCCACTACGCCCAGA |
| | ***** |
| Canine_A4_Normal_5-21 | GAAACGACCGGTCTCTACTGGCTGGCCAGTGTGCGCCCTGCCCATGATG |
| Human_COL4A4 | GAAACGACAGATCTCTACTGGCTGGCCAGCGCTGCGCCCTTCCCATGATG |
| Mouse_COL4A4 | GAAATGACAGGTCTCTACTGGCTGTCCAGTGTGCGCTCTCTCTTATGATG |
| | **** * * |
| Canine_A4_Normal_5-21 | CCGCTCTCAGAGGAGAGATCCGCCCTTACATCAGCCGCTGTGCCGTGTG |
| Human_COL4A4 | CCACTCTCTGAAGAGGCGATCCGCCCTATGTCTAGCCGCTGTGCCGTATG |
| Mouse_COL4A4 | CCACTCTCAGAGGAAGAGATCCGCTCTTACATCAGCCGCTGTGCCGTGTG |
| | ** ***** |
| Canine_A4_Normal_5-21 | CGAGGCCCCAGCCCCCGTGGTGGCGCTGCACAGCCAGGACCGGTCCATCC |
| Human_COL4A4 | CGAGGCCCCGCCCCAGGCGGTGGCGGTGCACAGCCAGGACCGGTCCATCC |
| Mouse_COL4A4 | TGAGGCCCCAGCACAGGCCGTGGCGGTCCACAGTCAGGACCAATCCATCC |
| | ***** ** * |
| Canine_A4_Normal_5-21 | CGCCGTGTCCACGGAGCTGGAGGAGCCTCTGGATCGGGTACTCGTTCCTG |
| Human_COL4A4 | CCCCATGTCCGCAGACCTGGAGGAGCCTCTGGATCGGGTATTCATTCCTG |
| Mouse_COL4A4 | CACCGTGTCCACGGACATGGAGGAGTCTCTGGATTGGGTACTCATTTCTG |
| | * ** ***** |
| Canine_A4_Normal_5-21 | ATGCACACAGGGGCTGGAGACCAAGGAGGAGGGCAGGCCCTCATGTCCCC |
| Human_COL4A4 | ATGCACACAGGAGCTGGGGACCAAGGAGGAGGGCAGGCCCTTATGTACC |
| Mouse_COL4A4 | ATGCACACTGGAGCTGGGGACCAAGGAGGTGGGCAGGCACTGATGTACC |
| | ***** ** |
| Canine_A4_Normal_5-21 | CGGCAGCTGTCTGGAAGATTTCGAGCCGCACCGTTTCCTCGAATGCCAAG |
| Human_COL4A4 | TGGCAGCTGCCTGGAAGATTTCAGAGCAGCACCATTCTTGAATGCCAGG |
| Mouse_COL4A4 | TGGCAGCTGCCTAGAGGACTTCAGAGCTGCACCGTTTCGTTGAATGCCAGG |
| | ***** ** * |
| Canine_A4_Normal_5-21 | GCCGACAGGGAACCTTGCCACTTTTTTGCAAATAGTATAGCTTCTGGCTG |
| Human_COL4A4 | GCCGGCAGGGAACCTTGCCACTTTTTTCGCAAATAAGTATAGCTTCTGGCTC |
| Mouse_COL4A4 | GCCGACAGGGAACCTGTCACTTTTTTGAAACGAGTACAGCTTCTGGCTG |
| | **** ***** |
| Canine_A4_Normal_5-21 | ACGACGGTGAGGCCTGACTTGCAGTTTTCTCGGCACCATCACCAGACAC |
| Human_COL4A4 | ACAACGGTGAAAGCAGACTTGCAGTTTTCTCTGCTCCAGCACCAGACAC |
| Mouse_COL4A4 | ACCACCGTGAACCCAGACCTGCAGTTTGCTCTGGCCCCGTCACCAGACAC |
| | ** * * |

| | |
|-----------------------|---|
| Canine_A4_Normal_5-21 | CTTGAAAGAAAGCCAGGCCAGCGCCAGAAAATCAGCAGGTGCCAGGTAT |
| Human_COL4A4 | CTTAAAAGAAAGCCAGGCCAACGCCAGAAAATCAGCCGGTGCCAGGTCT |
| Mouse_COL4A4 | CTTGAAAGAAGTTCAGGCCAGCGCGGAAAATCAGCAGGTGCCAGGTTT |
| | *** ***** ***** ***** ***** ***** ***** * |
| Canine_A4_Normal_5-21 | GCAT-----ATGACTAGT |
| Human_COL4A4 | GCGTGAAGTATAGCTAG- |
| Mouse_COL4A4 | GCATGAAGCACAGCTAA- |
| | ** * * * |

APPENDIX 6

Canine COL4A4 amino acid sequence alignments

Consensus key (see documentation for details)

- * - single, fully conserved residue
- : - conservation of strong groups
- . - conservation of weak groups
- no consensus

CLUSTAL W (1.81) multiple sequence alignment

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Canine_A4_Normal_5-21_Transl      RIYSYGCESLLEEKRNLTIFFKSQDLTVSRMHMASVKCSFRWIKPLARDP
Human_COL4A4_Translated_-_Fr      -----MWSLHIVLMRCSFRLTKSLATGP
Mouse_COL4A4_Translated_-_Fr      -----MRCFFRWTKSFVTAP
                                   ::* ** *.:. *

Canine_A4_Normal_5-21_Transl      WSLIFILFSVQHAYGSGKKFVGPCGGRDCSVCHCFPEKGSRGQPGPPGPQ
Human_COL4A4_Translated_-_Fr      WSLILILFSVQYVYGSGKKYIGPCGGRDCSVCHCVPEKGSRGPPGPPGPQ
Mouse_COL4A4_Translated_-_Fr      WSLIFILFTIQYEGSGKKYGGPCGGRNCSVCQCPEKGSRGHPGPLGPQ
                                   *****:***:***: *****: *****:*****:* ***** ** **

Canine_A4_Normal_5-21_Transl      GPIGPLGLPGPTGIPGEKGMRGDSGPPGAAGDKGDKGPTGVPGFPGLDGI
Human_COL4A4_Translated_-_Fr      GPIGPLGAPGPIGLSGEKGMRGDRGPPGAAGDKGDKGPTGVPGFPGLDGI
Mouse_COL4A4_Translated_-_Fr      GPIGPLGLPGPIGIPGEKGERGDSGSPGPPGEKGDKGPTGVPGFPGVDGV
                                   ***** ** *: ***** ** *.*. *:*****:***:

Canine_A4_Normal_5-21_Transl      PGHPGPPGSRGKPGMHGYNGSRGDPGFPGRGVPVPGGPPGLPGERGEKG
Human_COL4A4_Translated_-_Fr      PGHPGPPGPRGKPGMSGHNGSRGDPGFPGGRGALPGGGLGHPGEKGEKG
Mouse_COL4A4_Translated_-_Fr      PGHPGPPGPRGKPGVDGYNGSRGDPGYPGERGAPGPGGPPGQPGENGEKG
                                   *****.******: *:*****:* ** . ***** * **.*****

Canine_A4_Normal_5-21_Transl      NSVFILGAIKGIQDRGDPDLPLGLPGPKGAGGPAGPKQPGEPGLTGAPG
Human_COL4A4_Translated_-_Fr      NSVFILGAVKGIQDRGDPGLPLPGSWGAGGPAGPTGYPGEPGLVGPFG
Mouse_COL4A4_Translated_-_Fr      RSVYITGGVKGIQDRGDPGPPGLPGSRGAQGSPPGMGHAGAPGLAGPIG
                                   .**:* *:*****. *****. ** *.*. * * **.*. *

Canine_A4_Normal_5-21_Transl      HPGRPGLKGNPGVGKQMGDPGEVQQGSPGPTLLVQPPDSCLYKGEKG
Human_COL4A4_Translated_-_Fr      QPGRPGLKGNPGVGKQMGDPGEVQQGSPGPTLLVEPPDFCLYKGEKG
Mouse_COL4A4_Translated_-_Fr      HPGSPGLKGNPATGLKGQGEPEVQGRGPPGPTLLVQPPDLISYKGEKG
                                   :* *****. *:*** *:*****:* *****:*** .:*****

Canine_A4_Normal_5-21_Transl      IKGMPGMIGPPGLPGPKGEPGIGPKGEKGIPGFSGPRGDPGSYSGSPGFP
Human_COL4A4_Translated_-_Fr      IKGIPGMVGLPGPPGRKGESGIGAKGEKGIPGFPGRGDPGSYSGSPGFP
Mouse_COL4A4_Translated_-_Fr      VKGMPGMIGPPGPPGRKGAPGVGIKGEKGIPGFPGRGEPGSHGPPGFP
                                   :**:*:* * ** * * .*: *****. *****:***.* *****

Canine_A4_Normal_5-21_Transl      LKKGKPLFGDPGSFVGVPKGDGDRGYPPGVLVTPSVPLKGPPGDPG
Human_COL4A4_Translated_-_Fr      LKGLGLVGDPLFLGLIGPKGDGPNRGHPGPPGVLTPLPLKGPPGDPG
Mouse_COL4A4_Translated_-_Fr      FKGIQGAAGEPLFLGLGPKGDLGDRGYPPGPIILLTPAPPLKGVPDPG
                                   :* * * *:*** ***:***** *:***:*****:***. *****

Canine_A4_Normal_5-21_Transl      RPHYGEVGSVGGPPGPPGPPGEDCAGMMGPPGPRGFPGHGPFPGAAG
Human_COL4A4_Translated_-_Fr      FPGRYGETGDVGGPPGPPGLLGRPEACAGMIGPPGPQGFPLPLGEAG
Mouse_COL4A4_Translated_-_Fr      PPGYIGEIGDVLPGPPGPPGRGETCPGMMGPPGPPGVGPPGPFGEAG
                                   ** ** *.* ** ***** * ** *.*:***** *.* **:* **

Canine_A4_Normal_5-21_Transl      IPGRADLSPGKPGKPGPPGLPGAPGLQPPGSDVIYCSVGHGPGQGIKKG
Human_COL4A4_Translated_-_Fr      IPGRPDSAPGKPGKPGSPGLPGAPGLQLPGSSVIYCSVGNPGPQGIKKG
Mouse_COL4A4_Translated_-_Fr      VPGRLDCAPGKPGKPLGLPGAPGPEGPPGSDVIYCRPGCPGPMGEKKG
                                   :*** * :***** ***** :* **.* ***** * ** * **

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| | |
|------------------------------|--|
| Canine_A4_Normal_5-21_Transl | MGPPGRRGSKGEKGNAGLCACEPGPMGPPGPPGLPGRQGSKGLDGLPGWI |
| Human_COL4A4_Translated_-_Fr | VGPPGRRGPKGEKNEGLCACEPGPMGPPGPPGLPGRQGSKGLDGLPGWL |
| Mouse_COL4A4_Translated_-_Fr | VGPPGRRGAKGKNKGLCTCPGPMGPPGPPGPPGRQGSKGLDGLPGWH
:**** *.** ** ** *:***** ***** |
| Canine_A4_Normal_5-21_Transl | GEKGHPGPPGAEGSPGSPGKHGASGPPGSKGEKGDVVSRVKGKKGERGP |
| Human_COL4A4_Translated_-_Fr | GTKGDPGPPGAEGPPGLPGKHGASGPPGNKGAKGDMVVSrvKHKKGERGP |
| Mouse_COL4A4_Translated_-_Fr | GEKGDPGQPGAEGPPGPPGRPGAMGPPGHKGEKGDVSRVKQKGERGL
* **.** *****. ** *: * ** ** ** *:*****:***** |
| Canine_A4_Normal_5-21_Transl | DGLPGFPQGQNGRDGLPGKKGDPGPPGGHEDAAPGAKGSPGPPGPPGR |
| Human_COL4A4_Translated_-_Fr | DGPPGFPQPGSHGRDGHAGEKGDPPGPDHEDATPGKGFPGLPGPPGK |
| Mouse_COL4A4_Translated_-_Fr | DGPPGFPGPHGQDGGDGRPERGDPGPRGDHKDAPGERGLPLPGPPGR
** ***** *.** ** .*:***** .*:***** :* ** *****: |
| Canine_A4_Normal_5-21_Transl | TGPRGQPLGFPGPPGERGQPGAPGRPGERGFGLKGQKGDITSCNVTYP |
| Human_COL4A4_Translated_-_Fr | AGPVGPPGLGFPGPPGERGHPGVPHGVRGPDGLKGQKGDITSCNVTYP |
| Mouse_COL4A4_Translated_-_Fr | TGPEGPPGLGFPGPPGQRLPGEPGRPGTRGFDGTKGQKGDILCNVSY
: ** * *****: ** ** *: ** ** * *****: ** ** ** |
| Canine_A4_Normal_5-21_Transl | GRPGPPGFDGPPGPKGFPGPQGAFLRCLDGPKGQRGKPGMSEIPGPPGF |
| Human_COL4A4_Translated_-_Fr | GRHGPPGFDGPPGPKGFPGPQGAFLSGSDGHKGRPGTPTAETPGPPGF |
| Mouse_COL4A4_Translated_-_Fr | GKPLGLDGPPLGKGFPGPPGAPGMRCPDGQKQGRGKPGMSGIPGPPGF
*: * ** :***** ***** *****: ** **: *.** : ***** |
| Canine_A4_Normal_5-21_Transl | RGMGDPGFGEKEGSSSLGPPGFPGRGANGQKGMMDIAYGLPGPPGKR |
| Human_COL4A4_Translated_-_Fr | RGMGDPGFGEKEGSSSPVGPFPSPGSPVNGQKIPGDPAFGLGPPGKR |
| Mouse_COL4A4_Translated_-_Fr | RGMGDPGPIKGEKGTSPGPPGPPGSPGKDGQKIPGDPAFGDPGPPGER
*****: *****: ***** ** * :*****: ** *: * *****: |
| Canine_A4_Normal_5-21_Transl | GPSGVPGSKGHRGDSGRPGFAGPAGKPGSPGLKGRGREGSAGFQGI |
| Human_COL4A4_Translated_-_Fr | GLSGVPGIKGRGDPGCPGAEGPAGIPGFLGLKGPKGREGHAGFPGVPGP |
| Mouse_COL4A4_Translated_-_Fr | GLPGAPGMKGQKHGPGCPGAGPPGIPGSPGLKGPKGREGSRGFPPIPS
* *.** ** *: *.** ** *.** *****:***** ** **: |
| Canine_A4_Normal_5-21_Transl | PGHSCRGAPGIPGQPLPGAPGSPGAPGWKGQRGDVGPAGMKGLPG |
| Human_COL4A4_Translated_-_Fr | PGHSCRGAPGIPGQPLPGYPGSPGAPGGKQPGDVGPPGAGMKGLPG |
| Mouse_COL4A4_Translated_-_Fr | PGHSCRGAPGIPGQPLPGTPGDPGAPGWKGQPDGMPGSGPAGMKGLPG
*****:***** ***** *.***** ** *:***** |
| Canine_A4_Normal_5-21_Transl | VGPRPGAEGPLGFPVPGPLGDDGLPLPGPKGPQGLPGFPFPGGERGK |
| Human_COL4A4_Translated_-_Fr | LPGRPGAHGPPGLPGIPGPFDDGLPGPPGPKGPRGLPGFPFPGGERGK |
| Mouse_COL4A4_Translated_-_Fr | LPGLPGADGLRPPGIPGPNEDGLPLPLGLKGLPLPGFPFPGGERGK
: ** ** *. * ** :*** *:***** ** * ***** |
| Canine_A4_Normal_5-21_Transl | GPEGRPGSKGEPGEDGRPGFFGDQGVKGAKGARGPPGDEGEMAVFSQK |
| Human_COL4A4_Translated_-_Fr | GAEGCPGAKGEPGEKGMGLPGDRGLRGAKGAIGPPGDEGEMAIISQK |
| Mouse_COL4A4_Translated_-_Fr | GPDGEPGRKGEVGEKGWPLKGLDGERGAKGDRGLPGDAG-EAVTSRKGE
.: ** ** ** *.** *: ** * :***** * ** * *: ** * |
| Canine_A4_Normal_5-21_Transl | TGEPGPPGDDGFPGEEDKGNPGMPGRRGEPGRFGAPGFHRGEPGRGQ |
| Human_COL4A4_Translated_-_Fr | PGEFPGPDGDFPGERGDKGTGPMQGRRGELGRYGPFGFHRGEPGEKGQ |
| Mouse_COL4A4_Translated_-_Fr | PGDAGPPGDDGFSGERGDKSSGMRRGGRDPGRDGLPLHRGQPGIDGPP
*.:.*****.***.***.***.*** ** ** *: ** * ** :***** ** * |
| Canine_A4_Normal_5-21_Transl | GLPGPPGPPGSPGLRGIIGFPGFPGDQGEPSGSPGSLSGIDGMRGPKGS |
| Human_COL4A4_Translated_-_Fr | GPPGPPGPPGSTGLRGFIFGPGPLPGDQGEPSGPPGFSGIDGARGPKGN |
| Mouse_COL4A4_Translated_-_Fr | GPPGPPGPPGSPGLRGVIGFPGFPGDQDPSGPPGFPDGDGARGPKGY
* *****.*****.*****:*****.***.*** ***** |
| Canine_A4_Normal_5-21_Transl | KGDPASQFGPPGPKGEPGSPGCPQVGAPEQGLPGVQGLRGPGRPLG |
| Human_COL4A4_Translated_-_Fr | KGDPASHFGPPGPKGEPGSPGCPGHFGASGEQGLPGIIGPRGSPGRPGP |
| Mouse_COL4A4_Translated_-_Fr | KGDPASQCGPPGPKGEPGSPGYQRTGVPGEKGFPGDEGPRGPPGRPGQ
*****: ***** ***** *: *.** *: ** *:***** * |

| | |
|------------------------------|--|
| Canine_A4_Normal_5-21_Transl | GSSGPPGCPGNQGVPLTGPPGETGDPGPRGMMGDGPPGLPGIKGPSGS |
| Human_COL4A4_Translated_-_Fr | GSSGPPGCPGDHGMPLRGQPGEMGDGPRGLQGDGPIPGPPGIKGPSGS |
| Mouse_COL4A4_Translated_-_Fr | GSFGPPGCPGDGMPGLKGHPGEVGDGPRGDAGDFGRPGPAGVKGLGS |
| | ** *****: *:*** * *** ***** ** * ** .*:*** ** |
| Canine_A4_Normal_5-21_Transl | PGLNGLHGLKGQKGAKGASGLHEVGGPPGVGVPLKGEPDGPSPGISPP |
| Human_COL4A4_Translated_-_Fr | PGLNGLHGLKGQKGTGASGLHDVGGPPGVGIPGLKGERDGPSPGISPP |
| Mouse_COL4A4_Translated_-_Fr | PGLNGLHGLKGKGTGASGLLEMGPPGPMGMPGQKGEKGDGPSPGISPP |
| | *****:***** :*****:*** ** ***** |
| Canine_A4_Normal_5-21_Transl | GLSGERGPPPPGRPGAPGAGATGRAAEHVPDPGPPGDVGPPGPDGPR |
| Human_COL4A4_Translated_-_Fr | GPRGKKGPPGPPGSSGPPGAGATGRAPK-DIPDPGPPGDQGPDPGPR |
| Mouse_COL4A4_Translated_-_Fr | GLPGEGFGPPGRPGPPGAGAPGRAAKGDIIPDPGPPGDRGPPGPDGPR |
| | * **:~ ***** .*.*****.***.: .:***** ***** |
| Canine_A4_Normal_5-21_Transl | GAPGPPGPPGSDLLRGEFGDCGPPGPPGPPGPPGHQGFPGCDGKD |
| Human_COL4A4_Translated_-_Fr | GAPGPPGLPGSDLLRGEFGDCGLPGPPGPPGPPGPGYKGFPGCDGKD |
| Mouse_COL4A4_Translated_-_Fr | GVPGPPGSPGNVDLLKGDPGDCGLPGPPGSRGPPGPPGCGQPPGCDGKD |
| | *.***** ** .*****:***** ***** . ***** :* ***** |
| Canine_A4_Normal_5-21_Transl | QKGFPIGFPLQGPQLPLGEGKLLGIPGRRGHPGPPGSRGEPGPPADV |
| Human_COL4A4_Translated_-_Fr | QKGPMGFPGPQPHGFPGPGEKGLPGPPGRKGPTGLPGPRGEPGPPADV |
| Mouse_COL4A4_Translated_-_Fr | QKGPMGLPLGPPPLPGAPGEKGLPGPPGRKGFPVGPGRGEPGPPADV |
| | *****:*** ** *~***** * ****:* * ** ***** |
| Canine_A4_Normal_5-21_Transl | DSCPRIPGLPGVPGPRGPEGAMGVPGRGPPGPGCKGEPGLERRGEAGL |
| Human_COL4A4_Translated_-_Fr | DDCPRIPGLPGAGPMRGPEGAMGLPMRGPPGPGCKGEPGLDGRGVDGV |
| Mouse_COL4A4_Translated_-_Fr | DSCPRIPGLPGVPGPRGPEGAMGEPGRRGLPGPGCKGEPGPDGRRQDGI |
| | *.*****.~* ***** ** * ***** :**** * |
| Canine_A4_Normal_5-21_Transl | PGPPGPPGSTGDAGEAGCPGAPGPPGPIGDPGPRGFPGYLSGFLVLHS |
| Human_COL4A4_Translated_-_Fr | PGSPGPPGRKGDTEGEGYPPGPGPPGPIGDPGPKGFPGYLSGFLVLHS |
| Mouse_COL4A4_Translated_-_Fr | PGSPGPPGRKGDTEGAGCPGAPGPPGPTGDPGPKGFPGSLSGFLVLHS |
| | ** .***** .**:* * ** .***** *****:***** *.***** |
| Canine_A4_Normal_5-21_Transl | QTDGEPTCPAGMPRPWTGYSLLYLEGQEKAHNQDLGLAGSCLPMFSTLPP |
| Human_COL4A4_Translated_-_Fr | QTDQEPTCPLGMPRLWTGYSLLYLEGQEKAHNQDLGLAGSCLPVFSTLPP |
| Mouse_COL4A4_Translated_-_Fr | QTDQEPACFPVGMPLRLWTGYSLLYMEGQEKAHNQDLGLAGSCLPVFSTLPP |
| | *** **:* ** *****:*****:***** |
| Canine_A4_Normal_5-21_Transl | AYCNIHQVCHYARRNDRSYWLASAAPLPMPLSEEEIRPYISRCVCEAP |
| Human_COL4A4_Translated_-_Fr | AYCNIHQVCHYAQRNDRSYWLASAAPLPMPLSEEAIRPYVSRVCEAP |
| Mouse_COL4A4_Translated_-_Fr | AYCNIHQVCHYAQRNDRSYWLSSAAPLPMPLSEEEIRSYISRCVCEAP |
| | *****:*****:***** ***** *.***** |
| Canine_A4_Normal_5-21_Transl | APVVALHSQDRSIPPCPRSWRSLWIGYSFLMHTGAGDQGGGQALMSPGSC |
| Human_COL4A4_Translated_-_Fr | AQAVAVHSQDQSIPPCPQTRSLWIGYSFLMHTGAGDQGGGQALMSPGSC |
| Mouse_COL4A4_Translated_-_Fr | AQAVAVHSQDQSIPPCPRTWRSLWIGYSFLMHTGAGDQGGGQALMSPGSC |
| | * .**:****:*****:*****:***** |
| Canine_A4_Normal_5-21_Transl | LEDFRAAPFLECQGRQGTCHFFANEYSFWLTTVRPDLQFSSAPSPDTLKE |
| Human_COL4A4_Translated_-_Fr | LEDFRAAPFLECQGRQGTCHFFANKYSFWLTTVKADLQFSSAPAPDTLKE |
| Mouse_COL4A4_Translated_-_Fr | LEDFRAAPFVECQGRQGTCHFFANEYSFWLTTVNPDQLFASGSPDTLKE |
| | *****:*****:***** .*****.*.***** |
| Canine_A4_Normal_5-21_Transl | SQAQRQKISRCQVCIL-- |
| Human_COL4A4_Translated_-_Fr | SQAQRQKISRCQVCVKYS |
| Mouse_COL4A4_Translated_-_Fr | VQAQRKISRCQVCMKHS |
| | ****:*****: |

APPENDIX 7

Comparison of ECS-HN and normal COL4A3

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      30      40      50      60      70      80
ECS_A3  CGATGGTGTACCAGGATGCAATGGTTCTAAGGGTGAGCAAGGGTTTCCAGGCCTTCCAGG
        .....
NORMAL  CGATGGTGTACCAGGATGCAATGGTTCTAAGGGTGAGCAAGGGTTTCCAGGCCTTCCAGG
      30      40      50      60      70      80

      90     100     110     120     130     140
ECS_A3  GACACCAGGCTACCCAGGGATCCCGGGTGCTGTTGGCCTGAAAGGAGAAAAAGGTGCTCC
        .....
NORMAL  GACACCAGGCTACCCAGGGATCCCGGGTGCTGTTGGCCTGAAAGGAGAAAAAGGTGCTCC
      90     100     110     120     130     140

     150     160     170     180     190     200
ECS_A3  TGCCACAGAAGGTATAGAACTTGATGGAAGAGGTGACCCTGGGTTGCCAGGAGCTCCAGG
        .....
NORMAL  TGCCACAGAAGGTATAGAACTTGATGGAAGAGGTGACCCTGGGTTGCCAGGAGCTCCAGG
     150     160     170     180     190     200

     210     220     230     240     250     260
ECS_A3  ATTCCAGGGTTTACCRGGCCTTCCAGGCTTTCGGGACCTGCTGGTCCACCTGGCCCTCC
        .....
NORMAL  ATTCCAGGGTTTACCRGGCCTTCCAGGCTTTCGGGACCTGCTGGTCCACCTGGCCCTCC
     210     220     230     240     250     260

     270     280     290     300     310     320
ECS_A3  GGGATTCTTAGGCTTTCCAGGAGCCATGGGACCTCCAGGACCTAAGGGTCAAATGGGCGA
        .....
NORMAL  GGGATTCTTAGGCTTTCCAGGAGCCATGGGACCTCCAGGACCTAAGGGTCAAATGGGCGA
     270     280     290     300     310     320

     330     340     350     360     370     380
ECS_A3  TAAAGTGATAGGACAAAAAGGAGAGCGGGGTGTGAAAGGATTAACAGGACCTCCCGGACC
        .....
NORMAL  TAAAGTGATAGGACAAAAAGGAGAGCGGGGTGTGAAAGGATTAACAGGACCTCCCGGACC
     330     340     350     360     370     380

     390     400     410     420     430     440
ECS_A3  ACCAGGAACAGTTATTGTGACGCTAACCGGCCCTGATAACAGAACGGACCTYAAGGGGGA
        .....
NORMAL  ACCAGGAACAGTTATTGTGACGCTAACCGGCCCTGATAACAGAACGGACCTCAAGGGGGA
     390     400     410     420     430     440

     450     460     470     480     490     500
ECS_A3  GAAGGGAGACAAGGGAGCCATGGGGAAGCCTGGACCTCCTGGGCCCTTGGGACCGCCTGG
        .....
NORMAL  GAAGGGAGACAAGGGAGCCATGGGGAAGCCTGGACCTCCTGGGCCCTTGGGACCGCCTGG
     450     460     470     480     490     500

     510     520     530     540     550     560
ECS_A3  AGAATCCTATGGATCTGAAAAAGGTGCTCCTGGAGAACCTGGCCCGCAGGGAAAAACCTGG
        .....
NORMAL  AGAATCCTATGGATCTGAAAAAGGTGCTCCTGGAGAACCTGGCCCGCAGGGAAAAACCTGG
     510     520     530     540     550     560

     570     580     590     600     610     620
ECS_A3  CAAAGATGGTGCCCCCTGGTTTCCCTGGCACTGAGGGGCCCAAAGGCAACAGAGGCTTCCC
        .....
NORMAL  CAAAGATGGTGCCCCCTGGTTTCCCTGGCACTGAGGGGCCCAAAGGCAACAGAGGCTTCCC
     570     580     590     600     610     620

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        630          640          650          660          670          680
ECS_A3  TGGGTTACGGGGTGAAGACGGCATTAAAGGGGTGGAAAGGGGACATTGGCCCCGCCAGGATT
        .....
NORMAL  TGGGTTACGGGGTGAAGACGGCATTAAAGGGGTGGAAAGGGGACATTGGCCCCGCCAGGATT
        630          640          650          660          670          680

        690          700          710          720          730          740
ECS_A3  TCGTGGTCCAACAGAATATTATGATGCATACCAGGAAAAGGGTGATGAAGGAATTCCAGG
        .....
NORMAL  TCGTGGTCCAACAGAATATTATGATGCATACCAGGAAAAGGGTGATGAAGGAATTCCAGG
        690          700          710          720          730          740

        750          760          770          780          790          800
ECS_A3  CCCACCAGGCCCCAAAGGAGCTCGTGGCCCCCAGGGTCCAGCGGTCCCCCTGGAGTTCT
        .....
NORMAL  CCCACCAGGCCCCAAAGGAGCTCGTGGCCCCCAGGGTCCAGCGGTCCCCCTGGAGTTCT
        750          760          770          780          790          800

        810          820          830          840          850          860
ECS_A3  TGGAAGTGCTGGGTCATCAAAGCCTGGCCTCAGAGGAGCTCCTGGATCTCCAGGCATGAA
        .....
NORMAL  TGGAAGTGCTGGGTCATCAAAGCCTGGCCTCAGAGGAGCTCCTGGATCTCCAGGCATGAA
        810          820          830          840          850          860

        870          880          890          900          910          920
ECS_A3  AGGACGTAAAGGGGAACAAGGGCCCCCGGGAAGAATGCAGTGGGGCCTCCTGGGTCCCC
        .....
NORMAL  AGGACGTAAAGGGGAACAAGGGCCCCCGGGAAGAATGCAGTGGGGCCTCCTGGGTCCCC
        870          880          890          900          910          920

        930          940          950          960          970          980
ECS_A3  AGGTGTCTCGTTTACCAGGCCCCATAGGGCTGCCGGGATATCCAGGACCACCAGGTGA
        .....
NORMAL  AGGTGTCTCGTTTACCAGGCCCCATAGGGCTGCCGGGATATCCAGGACCACCAGGTGA
        930          940          950          960          970          980

        990          1000          1010          1020          1030          1040
ECS_A3  CATCGTGTTCGCAAAGGTCCACCTGGGGTTGGTGGACTCCCCGCCATGTTGGGTTTCC
        .....
NORMAL  CATCGTGTTCGCAAAGGTCCACCTGGGGTTGGTGGACTCCCCGCCATGTTGGGTTTCC
        990          1000          1010          1020          1030          1040

        1050          1060          1070          1080          1090          1100
ECS_A3  AGGCATCCCAGGAGTCGAGGGGCCCAAAGGGGAACCAGGCCTCTTGTGCGTACAGTGTGC
        .....
NORMAL  AGGCATCCCAGGAGTCGAGGGGCCCAAAGGGGAACCAGGCCTCTTGTGCGTACAGTGTGC
        1050          1060          1070          1080          1090          1100

        1110          1120          1130          1140          1150          1160
ECS_A3  TTATATCCCAGGGCGTCCAGGTCTCCAGGACTGCCAGGGTTGGATGGCATAAAAGGAAT
        .....
NORMAL  TTATATCCCAGGGCGTCCAGGTCTCCAGGACTGCCAGGGTTGGATGGCATAAAAGGAAT
        1110          1120          1130          1140          1150          1160

        1170          1180          1190          1200          1210          1220
ECS_A3  CCCAGGAGGACAAGGGGCAGCTGGCATTAAAGGAAGCCCAGGGTCCCAGGAAGTGCGGG
        .....
NORMAL  CCCAGGAGGACAAGGGGCAGCTGGCATTAAAGGAAGCCCAGGGTCCCAGGAAGTGCGGG
        1170          1180          1190          1200          1210          1220

        1230          1240          1250          1260          1270          1280
ECS_A3  TCTTCCAGGATTTCCAGGATTCCTGGGTGCTCCGGGTGCTCCAGGACTTAAAGGAGAAAA
        .....
NORMAL  TCTTCCAGGATTTCCAGGATTCCTGGGTGCTCCGGGTGCTCCAGGACTTAAAGGAGAAAA
        1230          1240          1250          1260          1270          1280

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      1290      1300      1310      1320      1330      1340
ECS_A3 AGGAGAAACATCTGAGCCTGAGGGACAGGTGGGTGCTCCAGGGGACCCCGGGCTCAGAGG
      .....
NORMAL AGGAGAAACATCTGAGCCTGAGGGACAGGTGGGTGCTCCAGGGGACCCCGGGCTCAGAGG
      1290      1300      1310      1320      1330      1340

      1350      1360      1370      1380      1390      1400
ECS_A3 CCATCCTGGAAGAAAGGGCTTGGATGGAATTCCTGGAACCTCCTGGAATTAAAGGACCACC
      .....
NORMAL CCATCCTGGAAGAAAGGGCTTGGATGGAATTCCTGGAACCTCCTGGAATTAAAGGACCACC
      1350      1360      1370      1380      1390      1400

      1410      1420      1430      1440      1450      1460
ECS_A3 AGGACCCAAAGGTGAACCGGCCCTGAGTGGTGAGAAGGGGACACGGGTCTCCAGGGGA
      .....
NORMAL AGGACCCAAAGGTGAACCGGCCCTGAGTGGTGAGAAGGGGACACGGGTCTCCAGGGGA
      1410      1420      1430      1440      1450      1460

      1470      1480      1490      1500      1510      1520
ECS_A3 TCCTGGGATCCCAGGGTCCCCAGGACCTGCAGGACCACCTGGACCACCAAACCTATGGACC
      .....
NORMAL TCCTGGGATCCCAGGGTCCCCAGGACCTGCAGGACCACCTGGACCACCAAACCTATGGACC
      1470      1480      1490      1500      1510      1520

      1530      1540      1550      1560      1570      1580
ECS_A3 ACAGGGAGAGCCTGGTCCAAAGGGCACCCAAGGAGTTCCTGGAGCCCCCTGGACCACCTGG
      .....
NORMAL ACAGGGAGAGCCTGGTCCAAAGGGCACCCAAGGAGTTCCTGGAGCCCCCTGGACCACCTGG
      1530      1540      1550      1560      1570      1580

      1590      1600      1610      1620      1630      1640
ECS_A3 AGAAGCCGGTCTTAAGGGAGAATTCGGTATTTTCGACACCAGTCCCAGGGCCCCCAGGACC
      .....
NORMAL AGAAGCCGGTCTTAAGGGAGAATTCGGTATTTTCGACACCAGTCCCAGGGCCCCCAGGACC
      1590      1600      1610      1620      1630      1640

      1650      1660      1670      1680      1690      1700
ECS_A3 TCCAGGGCCCCCTGGTTATGCTGGCCCCGAGGTCCACCTGGTATCCCTGGATCCATAGG
      .....
NORMAL TCCAGGGCCCCCTGGTTATGCTGGCCCCGAGGTCCACCTGGTATCCCTGGATCCATAGG
      1650      1660      1670      1680      1690      1700

      1710      1720      1730      1740      1750      1760
ECS_A3 AAAATGTGGTGATCCGGGTCTTCTGGGCCTGATGGTGAACCAGGAATTCAGGAATTGG
      .....
NORMAL AAAATGTGGTGATCCGGGTCTTCTGGGCCTGATGGTGAACCAGGAATTCAGGAATTGG
      1710      1720      1730      1740      1750      1760

      1770      1780      1790      1800      1810      1820
ECS_A3 CTTCCCTGGGCCCCCTGGACCTAAGGGAGACCAAGGTTTTCAGGAGCAAAGGAGCACC
      .....
NORMAL CTTCCCTGGGCCCCCTGGACCTAAGGGAGACCAAGGTTTTCAGGAGCAAAGGAGCACC
      1770      1780      1790      1800      1810      1820

      1830      1840      1850      1860      1870      1880
ECS_A3 GGGTTGTCCAGGAGAAATGGGGAAGCCCGGGTCACCTGGAGAACCAGGTCTCCAGGAGC
      .....
NORMAL GGGTTGTCCAGGAGAAATGGGGAAGCCCGGGTCACCTGGAGAACCAGGTCTCCAGGAGC
      1830      1840      1850      1860      1870      1880

      1890      1900      1910      1920      1930      1940
ECS_A3 CAAGGGAGAACCAAGGACTAGCCATACCTGGAGAACCAGGAATACCAGGTTTTCAGGAGA
      .....
NORMAL CAAGGGAGAACCAAGGACTAGCCATACCTGGAGAACCAGGAATACCAGGTTTTCAGGAGA
      1890      1900      1910      1920      1930      1940

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| | | | | | | |
|--------|--|------|------|------|------|------|
| | 1950 | 1960 | 1970 | 1980 | 1990 | 2000 |
| ECS_A3 | AAGAGGCAATTCTGGGGAAAATGGAGAAATTGGACTCCCTGGACTTCCAGGTCTCCCTGG | | | | | |
| | :: | | | | | |
| NORMAL | AAGAGGCAATTCTGGGGAAAATGGAGAAATTGGACTCCCTGGACTTCCAGGTCTCCCTGG | | | | | |
| | 1950 | 1960 | 1970 | 1980 | 1990 | 2000 |
| | | | | | | |
| | 2010 | 2020 | 2030 | 2040 | 2050 | 2060 |
| ECS_A3 | AATTCCAGGAACTGGAGGGCTTGATGGACCACCAGGGGATCCAGGGAAGCCTGGACCACC | | | | | |
| | :: | | | | | |
| NORMAL | AATTCCAGGAACTGGAGGGCTTGATGGACCACCAGGGGATCCAGGGAAGCCTGGACCACC | | | | | |
| | 2010 | 2020 | 2030 | 2040 | 2050 | 2060 |
| | | | | | | |
| | 2070 | 2080 | 2090 | 2100 | 2110 | 2120 |
| ECS_A3 | TGGAGAAAAGGACCCCAAGTTGCACAGAGGGTCCCAGAGGAGCCACGGACTTCC | | | | | |
| | :: | | | | | |
| NORMAL | TGGAGAAAAGGACCCCAAGTTGCACAGAGGGTCCCAGAGGAGCCACGGACTTCC | | | | | |
| | 2070 | 2080 | 2090 | 2100 | 2110 | 2120 |
| | | | | | | |
| | 2130 | 2140 | 2150 | 2160 | 2170 | 2180 |
| ECS_A3 | AGGATTAAATGGATTGGAAGGGCAACCAGGCAGAGAGGTGAAACAGGGCCAAAAGGAGA | | | | | |
| | :: | | | | | |
| NORMAL | AGGATTAAATGGATTGGAAGGGCAACCAGGCAGAGAGGTGAAACAGGGCCAAAAGGAGA | | | | | |
| | 2130 | 2140 | 2150 | 2160 | 2170 | 2180 |
| | | | | | | |
| | 2190 | 2200 | 2210 | 2220 | 2230 | 2240 |
| ECS_A3 | CCCAGGTATTCAGGCTTGATAGATCAGGCTTTCCTGGAGAACCTGGACCACCAGGAAT | | | | | |
| | :: | | | | | |
| Normal | CCCAGGTATTCAGGCTTGATAGATCAGGCTTTCCTGGAGAACCTGGACCACCAGGAAT | | | | | |
| | 2190 | 2200 | 2210 | 2220 | 2230 | 2240 |
| | | | | | | |
| | 2250 | 2260 | 2270 | 2280 | 2290 | 2300 |
| ECS_A3 | GCCAGGTCATCGAGGTGAGATAGGACCACCTGGCCAAAAGGATATCCAGGAAATCCGGG | | | | | |
| | :: | | | | | |
| Normal | GCCAGGTCATCGAGGTGAGATAGGACCACCTGGCCAAAAGGATATCCAGGAAATCCGGG | | | | | |
| | 2250 | 2260 | 2270 | 2280 | 2290 | 2300 |
| | | | | | | |
| | 2310 | 2320 | 2330 | 2340 | 2350 | 2360 |
| ECS_A3 | ATTTTTAGGACTACCAGGTGAAGATGGARTGATTGGGATGATGGGCTATCCAGGAAACAC | | | | | |
| | :: | | | | | |
| Normal | ATTTTTAGGACTACCAGGTGAAGATGGAGTGATTGGGATGATGGGCTATCCAGGAAACAC | | | | | |
| | 2310 | 2320 | 2330 | 2340 | 2350 | 2360 |
| | | | | | | |
| | 2370 | 2380 | 2390 | 2400 | 2410 | 2420 |
| ECS_A3 | TGGCCCTCCTGGGCCTCCCGGGATCCCAGGCACCCAGGACAGAGGGGTAGCTTTGGAAT | | | | | |
| | :: | | | | | |
| Normal | TGGCCCTCCTGGGCCTCCCGGGATCCCAGGCACCCAGGACAGAGGGGTAGCTTTGGAAT | | | | | |
| | 2370 | 2380 | 2390 | 2400 | 2410 | 2420 |
| | | | | | | |
| | 2430 | 2440 | 2450 | 2460 | 2470 | 2480 |
| ECS_A3 | TCCAGGTGCAAAGGGTGAGAAAGGGCCCCAGGAGCCAAGGGGAAGAAGGAGAGAAAGG | | | | | |
| | :: | | | | | |
| Normal | TCCAGGTGCAAAGGGTGAGAAAGGGCCCCAGGAGCCAAGGGGAAGAAGGAGAGAAAGG | | | | | |
| | 2430 | 2440 | 2450 | 2460 | 2470 | 2480 |
| | | | | | | |
| | 2490 | 2500 | 2510 | 2520 | 2530 | 2540 |
| ECS_A3 | ACCTATGGGGCCTTGTCGAATAACCAGCTCAGTGGGGGACAAAGGAGAACCAGGCCTCAA | | | | | |
| | :: | | | | | |
| Normal | ACCTATGGGGCCTTGTCGAATAACCAGCTCAGTGGGGGACAAAGGAGAACCAGGCCTCAA | | | | | |
| | 2490 | 2500 | 2510 | 2520 | 2530 | 2540 |
| | | | | | | |
| | 2550 | 2560 | 2570 | 2580 | 2590 | 2600 |
| ECS_A3 | AGGGTTTGCAAGGAAAGCCGGGTGAGAAAGGAAACAGAGGCATTCCAGGGTTACCAGGTTT | | | | | |
| | :: | | | | | |
| Normal | AGGGTTTGCAAGGAAAGCCGGGTGAGAAAGGAAACAGAGGCATTCCAGGGTTACCAGGTTT | | | | | |
| | 2550 | 2560 | 2570 | 2580 | 2590 | 2600 |

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      2610      2620      2630      2640      2650      2660
ECS_A3 GAAAGGATTCTGAAGGGCCACCTGGACCACCAGGCCACCAGGCCCGAGGAGATCCAGG
      .....
Normal GAAAGGATTCTGAAGGGCCACCTGGACCACCAGGCCACCAGGCCCGAGGAGATCCAGG
      2610      2620      2630      2640      2650      2660

      2670      2680      2690      2700      2710      2720
ECS_A3 CAGCATTGGGAATCCTGGAGAAGCAGGACCACGTGGAAGTCCAGGAAACATGGGGAACAT
      .....
Normal CAGCATTGGGAATCCTGGAGAAGCAGGACCACGTGGAAGTCCAGGAAACATGGGGAACAT
      2670      2680      2690      2700      2710      2720

      2730      2740      2750      2760      2770      2780
ECS_A3 GGGGATGCCAGGTCTTAAAGGACACAGGGGACCTTTGGGACTACCAGGTGTCTACTGGAAG
      .....
Normal GGGGATGCCAGGTCTTAAAGGACACAGGGGACCTTTGGGACTACCAGGTGTCTACTGGAAG
      2730      2740      2750      2760      2770      2780

      2790      2800      2810      2820      2830      2840
ECS_A3 ACCAGGCCTCCAGGTGTTACGGTCTCCAAGGAGACAAGGGAGAGCCAGGTTATTCAGC
      .....
Normal ACCAGGCCTCCAGGTGTTACGGTCTCCAAGGAGACAAGGGAGAGCCAGGTTATTCAGC
      2790      2800      2810      2820      2830      2840

      2850      2860      2870      2880      2890      2900
ECS_A3 AGGTACAAGGCCAGGACCACCGGGACTAAAGGGAGATCCAGGATTACCAGGTGCCATGGG
      .....
Normal AGGTACAAGGCCAGGACCACCGGGACTAAAGGGAGATCCAGGATTACCAGGTGCCATGGG
      2850      2860      2870      2880      2890      2900

      2910      2920      2930      2940      2950      2960
ECS_A3 AAGGAAAGGAGAAAAGAGGGCTACCTGGCACCCCTGGCCATTCGGGGCCTGCTGGAAGTGA
      .....
Normal AAGGAAAGGAGAAAAGAGGGCTACCTGGCACCCCTGGCCATTCGGGGCCTGCTGGAAGTGA
      2910      2920      2930      2940      2950      2960

      2970      2980      2990      3000      3010      3020
ECS_A3 GGGAGCCCCCTGGACATCCCGGAAGTCCTGGCCACCCAGGAAAGCCAGGCCCTGATGGAGA
      .....
Normal GGGAGCCCCCTGGACATCCCGGAAGTCCTGGCCACCCAGGAAAGCCAGGCCCTGATGGAGA
      2970      2980      2990      3000      3010      3020

      3030      3040      3050      3060      3070      3080
ECS_A3 TTTGGGGTTAAAAGGCATCAAAGGCTTCCCTGGGTCCCCAGGAGTCAAAGGACCTCCAGG
      .....
Normal TTTGGGGTTAAAAGGCATCAAAGGCTTCCCTGGGTCCCCAGGAGTCAAAGGACCTCCAGG
      3030      3040      3050      3060      3070      3080

      3090      3100      3110      3120      3130      3140
ECS_A3 ACCTCCAGGAATCCTAGGATCTCCTGGACCTATGGGCATGAGAGGCAACCAGGGCCGTGA
      .....
Normal ACCTCCAGGAATCCTAGGATCTCCTGGACCTATGGGCATGAGAGGCAACCAGGGCCGTGA
      3090      3100      3110      3120      3130      3140

      3150      3160      3170      3180      3190      3200
ECS_A3 TGGAATCCCTGGTCCCCCAGGAGAAAAGGGAGAAAACAGGTTTGCTGGGGGCACATCCAGG
      .....
Normal TGGAATCCCTGGTCCCCCAGGAGAAAAGGGAGAAAACAGGTTTGCTGGGGGCACATCCAGG
      3150      3160      3170      3180      3190      3200

      3210      3220      3230      3240      3250      3260
ECS_A3 CCCAGGAGGGATCCCTGGTCCTCCAGGAGCCAAAGGAGACAAGGGAGCCCCAGGCCTACC
      .....
Normal CCCAGGAGGGAGCCCTGGTCCTCCAGGAGCCAAAGGAGACAAGGGAGCCCCAGGCCTACC
      3210      3220      3230      3240      3250      3260

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      3270      3280      3290      3300      3310      3320
ECS_A3 AGGCCCCCAGGCAGGAAAGGGGCAGTGGGAGATGCCGGGCCACGGGGACCCACTGGCAT
      .....
Normal AGGCCCCCAGGCAGGAAAGGGGCAGTGGGAGATGCCGGGCCACGGGGACCCACTGGCAT
      3270      3280      3290      3300      3310      3320

      3330      3340      3350      3360      3370      3380
ECS_A3 GACGGGACTCCCAGGGCCACCAGGTTTTCCTGGTGCAGTCATCCCTGGCCAGAAAGGAAA
      .....
Normal GACGGGACTCCCAGGGCCACCAGGTTTTCCTGGTGCAGTCATCCCTGGCCAGAAAGGAAA
      3330      3340      3350      3360      3370      3380

      3390      3400      3410      3420      3430      3440
ECS_A3 TCAAGGTCCACCAGGCTTCAGAGGAAACCCAGGTGAGCCTGGTCTCCAGGCCCTCCAGG
      .....
Normal TCAAGGTCCACCAGGCTTCAGAGGAAACCCAGGTGAGCCTGGTCTCCAGGCCCTCCAGG
      3390      3400      3410      3420      3430      3440

      3450      3460      3470      3480      3490      3500
ECS_A3 GAGCCACGTGAAAGGCATAAAAGGAGACAAGGGACTCATGGGTGAGCCTGGCCCCAGAGG
      .....
Normal GAGCCACGTGAAAGGCATAAAAGGAGACAAGGGACTCATGGGTGAGCCTGGCCCCAGAGG
      3450      3460      3470      3480      3490      3500

      3510      3520      3530      3540      3550      3560
ECS_A3 TCTACCTGGAAGTGTGGGAGACAAGGGGCCACCGGGTCAACCGGGAGCACCAGGGAGCCC
      .....
Normal TCTACCTGGAAGTGTGGGAGACAAGGGGCCACCGGGTCAACCGGGAGCACCAGGGAGCCC
      3510      3520      3530      3540      3550      3560

      3570      3580      3590      3600      3610      3620
ECS_A3 AGGTCTGCCAGGGCTCAGGGGCGATCCTGGATTCTATGGATTTCCAGGTGTTAAAGGAGA
      .....
Normal AGGTCTGCCAGGGCTCAGGGGCGATCCTGGATTCTATGGATTTCCAGGTGTTAAAGGAGA
      3570      3580      3590      3600      3610      3620

      3630      3640      3650      3660      3670      3680
ECS_A3 GAAGGGTAACCCAGGATTTCGGGGACCAGCTGGACCTCCAGGGCAAATTTGGGCCAAAAGG
      .....
Normal GAAGGGTAACCCAGGATTTCGGGGACCAGCTGGACCTCCAGGGCAAATTTGGGCCAAAAGG
      3630      3640      3650      3660      3670      3680

      3690      3700      3710      3720      3730      3740
ECS_A3 ACCACCTGGTGTCCGTGGAGACCCTGGCACAGTTAAGATCATCTCCCTTCCAGGAAGCCC
      .....
Normal ACCACCTGGTGTCCGTGGAGACCCTGGCACAGTTAAGATCATCTCCCTTCCAGGAAGCCC
      3690      3700      3710      3720      3730      3740

      3750      3760      3770      3780      3790      3800
ECS_A3 AGGGCCACCTGGCCGTGCTGGAGGACCGGGGATGCCAGGAGAACCAGGGCCACCAGGGCC
      .....
Normal AGGGCCACCTGGCCGTGCTGGAGGACCGGGGATGCCAGGAGAACCAGGGCCACCAGGGCC
      3750      3760      3770      3780      3790      3800

      3810      3820      3830      3840      3850      3860
ECS_A3 ACCAGGAATCCTAGGACCCTGTGGACCAAGAGGTAAACCAGGCATGGATGGAATACCAGG
      .....
Normal ACCAGGAATCCTAGGACCCTGTGGACCAAGAGGTAAACCAGGCATGGATGGAATACCAGG
      3810      3820      3830      3840      3850      3860

      3870      3880      3890      3900      3910      3920
ECS_A3 AACTCCTGGGCCAATTGGAGAAAAAGGCAACAAAGGTTGTAAAGGCGAGCAAGGACCACC
      .....
Normal AACTCCTGGGCCAATTGGAGAAAAAGGCAACAAAGGTTGTAAAGGCGAGCAAGGACCACC
      3870      3880      3890      3900      3910      3920

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      3930      3940      3950      3960      3970      3980
ECS_A3 TGGATTAGATGGACTGCCAGGCTTGAAGGGGAGACCTGGAGACCCTGGACCACCTGCAAC
      .....
Normal TGGATTAGATGGACTGCCAGGCTTGAAGGGGAGACCTGGAGACCCTGGACCACCTGCAAC
      3930      3940      3950      3960      3970      3980

      3990      4000      4010      4020      4030      4040
ECS_A3 TGGGACCATGAAGAGGGGCTTTATCTTCACCAGGCATAGTCAGACCACAGTGATTCCCTTC
      .....
Normal TGGGACCATGAAGAGGGGCTTTATCTTCACCAGGCATAGTCAGACCACAGTGATTCCCTTC
      3990      4000      4010      4020      4030      4040

      4050      4060      4070      4080      4090      4100
ECS_A3 CTGTCCAGAAGGGACAGCGCCACTCTATAGTGGCTTTTCTTTCTTTTCATACAGGAAA
      .....
Normal CTGTCCAGAAGGGACAGCGCCACTCTATAGTGGCTTTTCTTTCTTTTCATACAGGAAA
      4050      4060      4070      4080      4090      4100

      4110      4120      4130      4140      4150      4160
ECS_A3 TGAACAAGCCCATGGACAAGACTTGGGAACCTCTCGGCAGCTGCCTGCAGCGATTACCAC
      .....
Normal TGAACAAGCCCATGGACAAGACTTGGGAACCTCTCGGCAGCTGCCTGCAGCGATTACCAC
      4110      4120      4130      4140      4150      4160

      4170      4180      4190      4200      4210      4220
ECS_A3 AATGCCATTCTTGTCTGTAAACATCAACAATGTCTGTAATTTGTCATCTCGAAATGACTA
      .....
Normal AATGCCATTCTTGTCTGTAAACATCAACAATGTCTGTAATTTGTCATCTCGAAATGACTA
      4170      4180      4190      4200      4210      4220

      4230      4240      4250      4260      4270      4280
ECS_A3 CTCATACTGGCTGTCAACACCAGCTCTGATGCCAATGGACATGGCTCCAATTACTGGCAG
      .....
Normal CTCATACTGGCTGTCAACACCAGCTCTGATGCCAATGGACATGGCTCCAATTACTGGCAG
      4230      4240      4250      4260      4270      4280

      4290      4300      4310      4320      4330      4340
ECS_A3 GGCCCTGGAGCCTTATATTAGCAGATGCACTGTCTGTGAAGGTCCTACGATTGCCATAGC
      .....
Normal GGCCCTGGAGCCTTATATTAGCAGATGCACTGTCTGTGAAGGTCCTACGATTGCCATAGC
      4290      4300      4310      4320      4330      4340

      4350      4360      4370      4380      4390      4400
ECS_A3 CATTACAGCCAAACCACTGACATTCCCTCATGTCCCAATGGCTGGATTCTCTCTGGAA
      .....
Normal CATTACAGCCAAACCACTGACATTCCCTCATGTCCCAATGGCTGGATTCTCTCTGGAA
      4350      4360      4370      4380      4390      4400

      4410      4420      4430      4440      4450      4460
ECS_A3 AGGATTTTCGTTTATCATGTTTCAAGTGCAGGTTCTGAGGGTGCTGGGCAAGCACTGGC
      .....
Normal AGGATTTTCGTTTATCATGTTTCAAGTGCAGGTTCTGAGGGTGCTGGGCAAGCACTGGC
      4410      4420      4430      4440      4450      4460

      4470      4480      4490      4500      4510      4520
ECS_A3 CTCCCCGGCTCCTGCCTGGAAGAGTTCCGAGCCAGTCCGTTTATASAATGTCACGGAAG
      .....
Normal CTCCCCGGCTCCTGCCTGGAAGAGTTCCGAGCCAGTCCGTTTATAGAATGTCACGGAAG
      4470      4480      4490      4500      4510      4520

      4530      4540      4550      4560      4570      4580
ECS_A3 AGGAACGTGCAACTACTATTCAAATTCCTACAGTTTCTGGTTGGCTTCGTAAACCCCA
      .....
Normal AGGAACGTGCAACTACTATTCAAATTCCTACAGTTTCTGGTTGGCTTCGTAAACCCCA
      4530      4540      4550      4560      4570      4580

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| | | | | | | |
|--------|--|------|------|------|------|------|
| | 4590 | 4600 | 4610 | 4620 | 4630 | 4640 |
| ECS_A3 | AAGAATGTTTCAGAAAACCTATTCCATCAACTGTGAAAGCTGAATCCATATGACTAGT | | | | | |
| | :: | | | | | |
| Normal | AAGAATGTTTCAGAAAACCTATTCCATCAACTGTGAAAGCTGAATCCATATGACTAGT | | | | | |
| | 4590 | 4600 | 4610 | 4620 | 4630 | 4640 |

APPENDIX 8

Comparison of ECS-HN and normal COL4A4

| | | | | | | |
|--------|--|-----|-----|-----|-----|-----|
| | 30 | 40 | 50 | 60 | 70 | 80 |
| Normal | GGATGTGAGAGTCTTCTGGAAGAGAAGAGAAACCTGACAATCTTTTAAAGTCCCAAGAC | | | | | |
| ESC A4 | GGATGTGAGAGTCTTCTGGAAGAGAAGAGAAACCTGACAATCTTTTAAAGTCCCAAGAC | | | | | |
| | 10 | 20 | 30 | 40 | 50 | 60 |
| | 90 | 100 | 110 | 120 | 130 | 140 |
| Normal | TTGACTGTGAGGTAATCCATGCACATGGCATCAGTAAAGTGTTTCATTTCAGGTGGATCAAG | | | | | |
| ESC A4 | TTGACTGTGAGGTAATCCATGCACATGGCATCAGTAAAGTGTTTCATTTCAGGTGGATCAAG | | | | | |
| | 70 | 80 | 90 | 100 | 110 | 120 |
| | 150 | 160 | 170 | 180 | 190 | 200 |
| Normal | CCCTTGCCAGAGATCCCTGGTCACTTATATTTATCCTCTTTTCTGTACAACATGCATAT | | | | | |
| ESC A4 | CCCTTGCCAGAGATCCCTGGTCACTTATATTTATCCTCTTTTCTGTACAACATGCATAT | | | | | |
| | 130 | 140 | 150 | 160 | 170 | 180 |
| | 210 | 220 | 230 | 240 | 250 | 260 |
| Normal | GGGAGTGGAAGAAGTTTGTGCGCCCTGTGGAGGAAGAGATTGCTCGGTGTGCCATTGC | | | | | |
| ESC A4 | GGGAGTGGAAGAAGTTTGTGCGCCCTGTGGAGGAAGAGATTGCTCGGTGTGCCATTGC | | | | | |
| | 190 | 200 | 210 | 220 | 230 | 240 |
| | 270 | 280 | 290 | 300 | 310 | 320 |
| Normal | TTTCTGAAAAAGGGTCTCGGGGTCAACCAGGACCGCGGGGCCACAGGGTCCCATTGGA | | | | | |
| ESC A4 | CTTCTGAAAAAGGGTCTCGGGGTCAACCAGGACCGCGGGGCCACAGGGTCCCATTGGA | | | | | |
| | 250 | 260 | 270 | 280 | 290 | 300 |
| | 330 | 340 | 350 | 360 | 370 | 380 |
| Normal | CCCCTGGGACTGCCAGGACCCACAGGAATTCCAGGAGAGAAAGGGATGAGAGGTGACAAG | | | | | |
| ESC A4 | CCCCTGGGACTGCCAGGACCCACAGGAATTCCAGGAGAGAAAGGGATGAGAAGTGACAAG | | | | | |
| | 310 | 320 | 330 | 340 | 350 | 360 |
| | 390 | 400 | 410 | 420 | 430 | 440 |
| Normal | TGGCCTCTCGGAGCAGCAGGTGACAAAGGTGACAAGGGTCCAACCTGGTGTTCCTGGATT | | | | | |
| ESC A4 | TGGCCTCTCGGAACAGCAGGTGACAAAGGTGACAAGGGTCCAACCTGGTGTTCCTGGATT | | | | | |
| | 370 | 380 | 390 | 400 | 410 | |
| | 450 | 460 | 470 | 480 | 490 | 500 |
| Normal | TCCAGGGTTGGATGGCATACTGGCCCCCGGGGCTCCTGGATCCAGAGGCAAGCCTGG | | | | | |
| ESC A4 | TCCAGGGTTGGATGGCATACTGGCCCCCGGGGCTCCTGGATCCAGAGGCAACCCTGG | | | | | |
| | 420 | 430 | 440 | 450 | 460 | 470 |
| | 510 | 520 | 530 | 540 | 550 | 560 |
| Normal | CATGCATGGCTACAATGGTTCACGAAGGTGATCCAGGGTTCCAGGAAAAAGAGGAGTTC | | | | | |
| ESC A4 | CATGCATGGCTACAATGGTTCACGAAGGTGATCCAGGGTTCCAGGAAAAAGAGGAGTTC | | | | | |
| | 480 | 490 | 500 | 510 | 520 | 530 |
| | 570 | 580 | 590 | 600 | 610 | 620 |
| Normal | CTGGCCAGGAGGCCCCCAGGCCTTCCTGGGGAAGGGGAGAAAAAGGAAATTCAGTGT | | | | | |
| ESC A4 | CTGGCCAGGAGGCCCCCAGGCCTTCCTGGGNAAGGGGAGAAAAAGGAAATTCAGTGT | | | | | |
| | 540 | 550 | 560 | 570 | 580 | 590 |

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        630      640      650      660      670      680
Normal TCATTTTAGGTGCCATTAAGGTATTCAGGGAGACAGGGGGACCCAGACCTGCCTGGCT
      .....
ESC A4 TCATTTTAGGTGCCATTAAGGTATTCAGGGAGACAGGGGGACCCAGACCTNCCTGGCT
      600      610      620      630      640      650

        690      700      710      720      730      740
Normal TGCCGGGACCGAAGGGGGCAGGAGGACCGGCAGGTCCCAGGGACAGCCAGGAGAGCCTG
      .....
ESC A4 TGCCGGGACCGAAGGGGGCAGGAGGACCGGCAGGNCCCAGGGGACAGCCAGGAGAGCCT
      660      670      680      690      700      710

        750      760      770      780      790      800
Normal GGTTAACAGGTGCTCCGGGCCATCCTGGGAGACCAGGCTTGAAGGGTAATCCTGGTGTGG
      .....
ESC A4 GGTTAACAGGTGCTCCGGGCCATCCTGGGAGACCAGGCTTGAAGGGTAATCCTGGTGTGG
      720      730      740      750      760      770

        810      820      830      840      850      860
Normal GAGTAAAGGGGCAAAATGGGAGACCCGGGTGAGGTTGGCCAGCAGGGTTCTCCTGGACCCA
      .....
ESC A4 GAGTAAAGGGGCAAAATGGGAGACCCGGGTGAGGTTGGCCAGCAGGGTTCTCCTGGACCCA
      780      790      800      810      820      830

        870      880      890      900      910      920
Normal CCTTATTGGTACAGCCACCTGATTCTGTCTGTATAAAGGAGAAAAGGGCATAAAAGGAA
      .....
ESC A4 CCTTATTGGTACAGCCACCTGATTCTGTCTGTATAAAGGAGAAAAGGGCATAAAAGGAA
      840      850      860      870      880      890

        930      940      950      960      970      980
Normal TGCCTGGTATGATTGGACCTCCAGGACTACCGGGACCCAGGGAGAACCTGGAATTGGAC
      .....
ESC A4 TGCCTGGTATGATTGGACCTCCAGGACTACCGGGACCCAGGGAGAACCTGGAATTGGAC
      900      910      920      930      940      950

        990      1000      1010      1020      1030      1040
Normal CAAAAGGAGAGAAGGGTATTCTTGGGTTCTCAGGACCTCGGGGTGATCCGGGTTCTATG
      .....
ESC A4 CAAAAGGAGAGAAGGGTATTCTTGGGTTCTCAGGACCTCGGGGTGATCCGGGTTCTATG
      960      970      980      990      1000      1010

        1050      1060      1070      1080      1090      1100
Normal GCTCTCCAGGTTTTCCAGGATTAAAGGGGAAACCAGGACTGTTTGGAGATCCTGGATCAT
      .....
ESC A4 GCTCTCCAGGTTTTCCAGGATTAAAGGGGAAACCAGGACTGTTTGGAGATCCTGGATCAT
      1020      1030      1040      1050      1060      1070

        1110      1120      1130      1140      1150      1160
Normal TTGGATTTGTGGCCCAAAGGGGGATCCTGGAGACCGTGGGTACCCAGGACCACCGGGTG
      .....
ESC A4 TTGGATTTGTGGCCCAAAGGGGGATCCTGGAGACCGTGGGTACCCAGGACCACCGGGTG
      1080      1090      1100      1110      1120      1130

        1170      1180      1190      1200      1210      1220
Normal TTCTGGTAACTCCATCTGTTCCTCAAGGCCCTCCAGGGGATCCGGGGCGCCCTGGCC
      .....
ESC A4 TTCTGGTAACTCCACCTGTTCCTCAAGGCCCTCCAGGGGATCCGGGGCGCCCTGGCC
      1140      1150      1160      1170      1180      1190

        1230      1240      1250      1260      1270      1280
Normal ACTATGGAGAAATGGGGTCCGTTGGACCACCTGGTCCCCCTGGTCCCCAGGTCCACCAG
      .....
ESC A4 ACTATGGAGAAATGGGGTCCGTTGGACCACCTGGTCCCCCTGGTCCCCAGGTCCACCAG
      1200      1210      1220      1230      1240      1250

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| | | | | | | |
|--------|--|------|------|------|------|------|
| | 2610 | 2620 | 2630 | 2640 | 2650 | 2660 |
| Normal | GGCAACCGGGGCTCCCTGGGGCCCCGGGGAGTCCAGGTGCCCCAGGTTGGAAAGGACAGC | | | | | |
| | :: | | | | | |
| ESC A4 | GGCAACCGGGGCTCCCTGGGGCCCCGGGGAGTCCAGGTGCCCCAGGTTGGAAAGGACAGC | | | | | |
| | 2580 | 2590 | 2600 | 2610 | 2620 | 2630 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 2670 | 2680 | 2690 | 2700 | 2710 | 2720 |
| Normal | GAGGGGATGTGGGGCCTCCGGGTCCAGCTGGAATGAAAGGCCTCCCTGGAGTCCCTGGAC | | | | | |
| | :: | | | | | |
| ESC A4 | GAGGGGATGTGGGGCCTCCGGGTCCAGCTGGAATGAAAGGCCTCCCTGGAGTCCCTGGAC | | | | | |
| | 2640 | 2650 | 2660 | 2670 | 2680 | 2690 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 2730 | 2740 | 2750 | 2760 | 2770 | 2780 |
| Normal | GGCCAGGGGAGAGGGACCCCTAGGATTCCCAGGAGTCCCAGGCCCTTAGGGGATGATG | | | | | |
| | :: | | | | | |
| ESC A4 | GGCCAGGGGAGAGGGACCCCTAGGATTCCCAGGAGTCCCAGGCCCTTAGGGGATGATG | | | | | |
| | 2700 | 2710 | 2720 | 2730 | 2740 | 2750 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 2790 | 2800 | 2810 | 2820 | 2830 | 2840 |
| Normal | GGCTGCCTGGTCTTCCAGGCCGAAGGGACCCAGGGGCTGCCTGGCTTCCCCGGTTTTTC | | | | | |
| | :: | | | | | |
| ESC A4 | GGCTGCCTGGTCTTCCAGGCCGAAGGGACCCAGGGGCTGCCTGGCTTCCCCGGTTTTTC | | | | | |
| | 2760 | 2770 | 2780 | 2790 | 2800 | 2810 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 2850 | 2860 | 2870 | 2880 | 2890 | 2900 |
| Normal | CAGGGGAGAGAGGAAAGCCTGGCCCAGAGGGACGCCCTGGCAGCAAGGGGGAACAGGAG | | | | | |
| | :: | | | | | |
| ESC A4 | CAGGGGAGAGAGGAAAGCCTGGCCCAGAGGGACGCCCTGGCAGCAAGGGGGAACAGGAG | | | | | |
| | 2820 | 2830 | 2840 | 2850 | 2860 | 2870 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 2910 | 2920 | 2930 | 2940 | 2950 | 2960 |
| Normal | AGGATGGTTCGGCCTGGCTTCTTCGAGACCAAGGGGTGAAAGGTGCCAAAGGAGCGAGAG | | | | | |
| | :: | | | | | |
| ESC A4 | AGGATGGTTCGGCCTGGCTTCTTCGAGACCAAGGGGTGAAAGGTGCCAAAGGAGCGAGAG | | | | | |
| | 2880 | 2890 | 2900 | 2910 | 2920 | 2930 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 2970 | 2980 | 2990 | 3000 | 3010 | 3020 |
| Normal | GACCCCCAGGAGATGAAGGAGAGATGGCGGTCTTTTCCCAAAAAGGGAAAACCGGGGAAC | | | | | |
| | :: | | | | | |
| ESC A4 | GACCCCCAGGAGATGAAGGAGAGATGGCGGTCTTTTCCCAAAAAGGGAAAACCGGGGAAC | | | | | |
| | 2940 | 2950 | 2960 | 2970 | 2980 | 2990 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 3030 | 3040 | 3050 | 3060 | 3070 | 3080 |
| Normal | CTGGACCTCCAGGAGATGATGGATTCCCAGGAGAAGAAGGTGATAAAGGCAATCCAGGGA | | | | | |
| | :: | | | | | |
| ESC A4 | CTGGACCTCCAGGAGATGATGGATTCCCAGGAGAAGAAGGTGATAAAGGCAATCCAGGGA | | | | | |
| | 3000 | 3010 | 3020 | 3030 | 3040 | 3050 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 3090 | 3100 | 3110 | 3120 | 3130 | 3140 |
| Normal | TGCCGGGGAGGAGAGAGAGCCTGGAAGATTTCGAGCACCTGGATTTCACAGAGGGGAGC | | | | | |
| | :: | | | | | |
| ESC A4 | TGCCGGGGAGGAGAGAGAGCCTGGAAGATTTCGAGCACCTGGATTTCACAGAGGGGAGC | | | | | |
| | 3060 | 3070 | 3080 | 3090 | 3100 | 3110 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 3150 | 3160 | 3170 | 3180 | 3190 | 3200 |
| Normal | CCGGGAGAACCGGGCAGCCAGGGCTTCTTGCCCCCAGGCCCCCAGGCTCACCTGGGC | | | | | |
| | :: | | | | | |
| ESC A4 | CCGGGAGAACCGGGCAGCCAGGGCTTCTTGCCCCCAGGCCCCCAGGCTCACCTGGGC | | | | | |
| | 3120 | 3130 | 3140 | 3150 | 3160 | 3170 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 3210 | 3220 | 3230 | 3240 | 3250 | 3260 |
| Normal | TGAGAGGGATTATTGGTTTTCCGGGATTTCCAGGTGACCAGGGTGAGCCAGGTTCTCCAG | | | | | |
| | :: | | | | | |
| ESC A4 | TGAGAGGGATTATTGGTTTTCCGGGATTTCCAGGTGACCAGGGTGAGCCAGGTTCTCCAG | | | | | |
| | 3180 | 3190 | 3200 | 3210 | 3220 | 3230 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 3270 | 3280 | 3290 | 3300 | 3310 | 3320 |
| Normal | GGTCTCCTGGACTTTCAGGAATTGATGGAATGAGAGGACCTAAAGGAAGCAAAGGTGACC | | | | | |
| | :: | | | | | |
| ESC A4 | GGTCTCCTGGACTTTCAGGAATTGATGGAATGAGAGGACCTAAAGGAAGCAAAGGTGACC | | | | | |
| | 3240 | 3250 | 3260 | 3270 | 3280 | 3290 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 3330 | 3340 | 3350 | 3360 | 3370 | 3380 |
| Normal | CTGCAAGTCAGTTTCGGCCACCTGGTCCAAAGGGTGAACCAGGTAGCCCTGGATGTCCAG | | | | | |
| | :: | | | | | |
| ESC A4 | CTGCAAGTCAGTTTCGGCCACCTGGTCCAAAGGGTGAACCAGGTAGCCCTGGATGTCCAG | | | | | |
| | 3300 | 3310 | 3320 | 3330 | 3340 | 3350 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 3390 | 3400 | 3410 | 3420 | 3430 | 3440 |
| Normal | GACAAGTTGGAGCACCTGGAGAGCAGGGCTTGCTGGTGTTCAGGGCTCAGAGGACCAC | | | | | |
| | :: | | | | | |
| ESC A4 | GACAAGTTGGAGCACCTGGAGAGCAGGGCTTGCTGGTGTTCAGGGCTCAGAGGACCAC | | | | | |
| | 3360 | 3370 | 3380 | 3390 | 3400 | 3410 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 3450 | 3460 | 3470 | 3480 | 3490 | 3500 |
| Normal | CTGGAAGGCCAGGATTACCTGGCTCCTCCGACCACCAGGGTGTCCAGGTAATCAAGGGG | | | | | |
| | :: | | | | | |
| ESC A4 | CTGGAAGGCCAGGATTACCTGGCTCCTCCGACCACCAGGGTGTCCAGGTAATCAAGGGG | | | | | |
| | 3420 | 3430 | 3440 | 3450 | 3460 | 3470 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 3510 | 3520 | 3530 | 3540 | 3550 | 3560 |
| Normal | TGCTTGGGCTGACAGGGCCTCCAGGAGAAACAGGGGATCCTGGGCCAAGAGGCATGATGG | | | | | |
| | :: | | | | | |
| ESC A4 | TGCTTGGGCTGACAGGGCCTCCAGGAGAAACAGGGGATCCTGGGCCAAGAGGCATGATGG | | | | | |
| | 3480 | 3490 | 3500 | 3510 | 3520 | 3530 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 3570 | 3580 | 3590 | 3600 | 3610 | 3620 |
| Normal | GAGATCCAGGGCCACCAGGTCTTCCAGGAATAAAAGGTCCCTCCGGGTCGCCGGGTCTGA | | | | | |
| | :: | | | | | |
| ESC A4 | GAGATCCAGGGCCACCAGGTCTTCCAGGAATAAAAGGTCCCTCCGGGTCGCCGGGTCTGA | | | | | |
| | 3540 | 3550 | 3560 | 3570 | 3580 | 3590 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 3630 | 3640 | 3650 | 3660 | 3670 | 3680 |
| Normal | ATGGCTTGATGGTTTAAAGGGTCAGAAAGGAGCCAAAGGTGCTTCAGGTTTGCACGAAG | | | | | |
| | :: | | | | | |
| ESC A4 | ATGGCTTGATGGTTTAAAGGGTCAGAAAGGAGCCAAAGGTGCTTCAGGTTTGCACGAAG | | | | | |
| | 3600 | 3610 | 3620 | 3630 | 3640 | 3650 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 3690 | 3700 | 3710 | 3720 | 3730 | 3740 |
| Normal | TGGGCCCACCCGGTCCAGTGGGCGTACCTGGGCTGAAAGGGGAGCCGGGAGACCTGGGA | | | | | |
| | :: | | | | | |
| ESC A4 | TGGGCCCACCCGGTCCAGTGGGCGTACCTGGGCTGAAAGGGGAGCCGGGAGACCTGGGA | | | | | |
| | 3660 | 3670 | 3680 | 3690 | 3700 | 3710 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 3750 | 3760 | 3770 | 3780 | 3790 | 3800 |
| Normal | GCCCAGGAATTTCTCCCCAGGCCTTTCTGGAGAAAGAGGCCCCCGGCCCCCAGGGA | | | | | |
| | :: | | | | | |
| ESC A4 | GCCCAGGAATTTCTCCCCAGGCCTTTCTGGAGAAAGAGGCCCCCGGCCCCCAGGGA | | | | | |
| | 3720 | 3730 | 3740 | 3750 | 3760 | 3770 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 3810 | 3820 | 3830 | 3840 | 3850 | 3860 |
| Normal | GACCTGGAGCCCCCTGGTCTGCAGGTGCCACAGGAAGAGCTGCTGAAGGTCACGTTCTTG | | | | | |
| | :: | | | | | |
| ESC A4 | GACCTGGAGCCCCCTGGTCTGCAGGTGCCACAGGAAGAGCTGCTGAAGGTCACGTTCTTG | | | | | |
| | 3780 | 3790 | 3800 | 3810 | 3820 | 3830 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 3870 | 3880 | 3890 | 3900 | 3910 | 3920 |
| Normal | ACCCAGGTCCACCCGAGATGTGGGTCTCCTGGCCCCGATGGTCCGAGAGGAGCGCCCCG | | | | | |
| | :: | | | | | |
| ESC A4 | ACCCAGGTCCACCCGAGATGTGGGTCTCCTGGCCCCGATGGTCCGAGAGGAGCGCCCCG | | | | | |
| | 3840 | 3850 | 3860 | 3870 | 3880 | 3890 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 3930 | 3940 | 3950 | 3960 | 3970 | 3980 |
| Normal | GGCCCCCAGGCCCCCTGGGAGCGTTGACCTTCTGAGAGGGGAACCAGGAGACTGTGGGC | | | | | |
| | :: | | | | | |
| ESC A4 | GGCCCCCAGGCCCCCTGGGAGCGTTGACCTTCTGAGAGGGGAACCAGGAGACTGTGGGC | | | | | |
| | 3900 | 3910 | 3920 | 3930 | 3940 | 3950 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 3990 | 4000 | 4010 | 4020 | 4030 | 4040 |
| Normal | CGCCGGGGCCTCCAGGTCCCCCGGGCCCACCCGGCCCTCCAGGACACCAAGGCTTCCCAG | | | | | |
| | :: | | | | | |
| ESC A4 | CGCCGGGGCCTCCAGGTCCCCCGGGCCCACCCGGCCCTCCAGGACACCAAGGCTTCCCAG | | | | | |
| | 3960 | 3970 | 3980 | 3990 | 4000 | 4010 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 4050 | 4060 | 4070 | 4080 | 4090 | 4100 |
| Normal | GATGCGATGGAAGACGGCCAGAAAGGACCAATAGGATTCCCGGGGCTGCAGGGGCCTC | | | | | |
| | :: | | | | | |
| ESC A4 | GATGCGATGGAAGACGGCCAGAAAGGACCAATAGGATTCCCGGGGCTGCAGGGGCCTC | | | | | |
| | 4020 | 4030 | 4040 | 4050 | 4060 | 4070 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 4110 | 4120 | 4130 | 4140 | 4150 | 4160 |
| Normal | AGGGACTTCTGGGCTCCCTGGGGAGAAGGGTCTGCTGGGCATTCCAGGCCGGCGAGGGC | | | | | |
| | :: | | | | | |
| ESC A4 | AGGGACTTCTGGGCTCCCTGGGGAGAAGGGTCTGCTGGGCATTCCAGGCCGGCGAGGGC | | | | | |
| | 4080 | 4090 | 4100 | 4110 | 4120 | 4130 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 4170 | 4180 | 4190 | 4200 | 4210 | 4220 |
| Normal | ACCCCGGTCTCCAGGTTCCAGAGGTGAACAGGGCCTCCTGCAGATGTGGATTCTGTGCC | | | | | |
| | :: | | | | | |
| ESC A4 | ACCCCGGTCTCCAGGTTCCAGAGGTGAACAGGGCCTCCTGCAGATGTGGATTCTGTGCC | | | | | |
| | 4140 | 4150 | 4160 | 4170 | 4180 | 4190 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 4230 | 4240 | 4250 | 4260 | 4270 | 4280 |
| Normal | CCCGCATCCCCGGGCTTCTGGGGTACCAGGCCCAAGAGGACCAGAAGGAGCCATGGGGG | | | | | |
| | :: | | | | | |
| ESC A4 | CCCGCATCCCCGGGCTTCTGGGGTACCAGGCCCAAGAGGACCAGAAGGAGCCATGGGGG | | | | | |
| | 4200 | 4210 | 4220 | 4230 | 4240 | 4250 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 4290 | 4300 | 4310 | 4320 | 4330 | 4340 |
| Normal | TCCCTGGAAGGAGAGGGCCCCCAGGACCAGGGTGCAAAGGAGAGCCGGGACTGGAGGGCA | | | | | |
| | :: | | | | | |
| ESC A4 | TCCCTGGAAGGAGAGGGCCCCCAGGACCAGGGTGCAAAGGAGAGCCGGGACTGGAGGGCA | | | | | |
| | 4260 | 4270 | 4280 | 4290 | 4300 | 4310 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 4350 | 4360 | 4370 | 4380 | 4390 | 4400 |
| Normal | GGAGGGGCGAGGCCGGCCTCCCAGGACCTCCGGGGCCTCCCGGAAGCACAGGGGACGCAG | | | | | |
| | :: | | | | | |
| ESC A4 | GGAGGGGCGAGGCCGGCCTCCCAGGACCTCCGGGGCCTCCCGGAAGCACAGGGGACGCAG | | | | | |
| | 4320 | 4330 | 4340 | 4350 | 4360 | 4370 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 4410 | 4420 | 4430 | 4440 | 4450 | 4460 |
| Normal | GAGAAGCCGTTGCCCGGAGCACCAGGGCCCCCGGGCCATTGGGGACCCCGGGCCCA | | | | | |
| | :: | | | | | |
| ESC A4 | GAGAAGCCGTTGCCCGGAGCACCAGGGCCCCCGGGCCATTGGGGACCCCGGGCCCA | | | | | |
| | 4380 | 4390 | 4400 | 4410 | 4420 | 4430 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 4470 | 4480 | 4490 | 4500 | 4510 | 4520 |
| Normal | GAGGGTTTGGGCCTGGATACCTCAGTGGCTTCCTCCTGGTTCTCCACAGTCAGACGGATG | | | | | |
| | :: | | | | | |
| ESC A4 | GAGGGTTTGGGCCTGGATACCTCAGTGGCTTCCTCCTGGTTCTCCACAGTCAGACGGATG | | | | | |
| | 4440 | 4450 | 4460 | 4470 | 4480 | 4490 |

| | | | | | | |
|--------|--|------|------|------|------|------|
| | 4530 | 4540 | 4550 | 4560 | 4570 | 4580 |
| Normal | GGGAGCCACCTGCCCCGCGGGCATGCCAGGCCCTGGACGGGCTACAGTCTGTTATACC | | | | | |
| | :: | | | | | |
| ESC A4 | GGGAGCCACCTGCCCCGCGGGCATGCCAGGCCCTGGACGGGCTACAGTCTGTTATACC | | | | | |
| | 4500 | 4510 | 4520 | 4530 | 4540 | 4550 |

```

          4590      4600      4610      4620      4630      4640
Normal  TGG AAGGACAGGAGAAGGCCACAATCAGGACCTTGGTCTGGCAGGGTCTTGCCTTCCCA
          .....
ESC A4  TGG AAGGACAGGAGAAGGCCACAATCAGGACCTTGGTCTGGCAGGGTCTTGCCTTCCCA
          4560      4570      4580      4590      4600      4610

          4650      4660      4670      4680      4690      4700
Normal  TGTTCAGCACCTTGCCCTTTGCCTACTGCAACATCCACCAAGTGTGCCACTACGCCC GGA
          .....
ESC A4  TGTTCAGCACCTTGCCCTTTGCCTACTGCAACATCCACCAAGTGTGCCACTACGCCC GGA
          4620      4630      4640      4650      4660      4670

          4710      4720      4730      4740      4750      4760
Normal  GAAACGACCGGTCTTACTGGCTGGCCAGTGTGCGCCCCTGCCCATGATGCCGCTCTCAG
          .....
ESC A4  GAAACGACCGGTCTTACTGGCTGGCCAGTGTGCGCCCCTGCCCATGATGCCGCTCTCAG
          4680      4690      4700      4710      4720      4730

          4770      4780      4790      4800      4810      4820
Normal  AGGAGGAGATCCGCCCCCTACATCAGCCGCTGTGCCGTGTGCGAGGCCCCAGCCCCCGTGG
          .....
ESC A4  AGGAGGAGATCCGCCCCCTACATCAGCCGCTGTGCCGTGTGCGAGGCCCCAGCCCCCGTGG
          4740      4750      4760      4770      4780      4790

          4830      4840      4850      4860      4870      4880
Normal  TGGCGCTGCACAGCCAGGACCGGTCCATCCCGCGTGTCCACGGAGCTGGAGGAGCCTCT
          .....
ESC A4  TGGCGCTGCACAGCCAGGACCGGTCCATCCCGCGTGTCCACGGAGCTGGAGGAGCCTCT
          4800      4810      4820      4830      4840      4850

          4890      4900      4910      4920      4930      4940
Normal  GGATCGGGTACTCGTTCCTGATGCACACAGGGGCTGGAGACCAAGGAGGAGGGCAGGCC
          .....
ESC A4  GGATCGGGTACTCGTTCCTGATGCACACAGGGGCTGGAGACCAAGGAGGAGGGCAGGCC
          4860      4870      4880      4890      4900      4910

          4950      4960      4970      4980      4990      5000
Normal  TCATGTCCCCCGGCAGCTGTCTGGAAGATTCCGAGCCGCACCGTTCTCGAATGCCAAG
          .....
ESC A4  TCATGTCCCCCGGCAGCTGTCTGGAAGATTCCGAGCCGCACCGTTCTCGAATGCCAAG
          4920      4930      4940      4950      4960      4970

          5010      5020      5030      5040      5050
Normal  GCCGACAGGGAAC TTGCCACTTTTTTGCAAATGAGTATAGCTTCTGGCTTCTGGCTGACGAC
          .....
ESC A4  GCCGACAGGGAAC TTGCCACTTTTTTGCAAATGAGTATAGCTTCTGGCTTCTGGCTGACGAC
          4980      4990      5000      5010      5020

```

APPENDIX 9

Effects of Aberrant COL4A3 Nucleotides

| Position | Change | Effect |
|----------|--------|------------------------------------|
| ECS 225 | R | All bases conserve proline (CCN) |
| ECS 440 | Y | All bases conserve leucine (CTN) |
| ECS 2340 | R | All bases conserve valine (GTN) |
| ECS 2507 | A-G | Conserves codon for glutamic acid |
| ECS 2575 | G-A | Conserves codon for glutamic acid |
| ECS 3250 | A-G | Conserves codon for glutamine |
| ECS 3338 | Y | All bases conserve threonine (ACN) |
| ECS 3220 | G-T | serine-isoleucine |
| ECS 3250 | G-A | arginine-lysine |
| ECS 3254 | C-A | Conserves codon for glycine |

APPENDIX 10

Effects of Aberrant COL4A4 Nucleotides

| Position | Change | Effect |
|----------|--------|-----------------------------------|
| ECS 99 | G-A | Conserves AAA codon for lysine |
| ECS 195 | N | Unknown |
| ECS 351 | G-A | Conserves GAA codon for glutamine |
| ECS 371 | G-A | Conserves GAA codon for glutamine |
| ECS 475 | G-C | Asparagine-Lysine |
| ECS 573 | N | Unknown |
| ECS 630 | N | Unknown |
| ECS 650 | N | All bases conserve leucine (CTN) |
| ECS 691 | N | All bases conserve glycine (GGN) |
| ECS 1179 | C-T | Conserves codon for leucine |
| ECS 1707 | C-T | Conserves codon for glycine |
| ECS 2078 | C-T | Conserves codon for asparagine |
| ECS 2099 | C-T | Conserves codon for proline |
| ECS 2100 | A-G | Conserves codon for glycine |
| ECS 4530 | C-T | Conserves codon for glycine |

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